



Abundance and viability of particle-attached and free-floating bacteria in dusty and nondust air

Wei Hu^{1,2}, Kotaro Murata^{2,3}, Chunlan Fan², Shu Huang¹, Hiromi Matsusaki², Pingqing Fu¹, Daizhou Zhang²

5 ¹ Institute of Surface-Earth System Science, Tianjin University, Tianjin, 300072, China

² Faculty of Environmental and Symbiotic Sciences, Prefectural University of Kumamoto, Kumamoto, 862-8502, Japan

³ Department of Physics, Tokyo Gakugei University, Tokyo, 184-8501, Japan

Correspondence to: Daizhou Zhang (dzzhang@pu-kumamoto.ac.jp)

10 **Abstract.** Widespread bacteria are a major proportion of bioaerosols and their coexistence with dust enables both types of aerosols more active in ice cloud formation and harmful to public health. However, the abundance and viability of particle-attached and free-floating bacteria in dusty air have not been quantitatively investigated. We researched this subject based on the fact that airborne bacterial cells are approximately 1 μm or smaller; therefore, particle-attached bacteria should occur in aerosol samples of particles larger than 1 μm , and free-floating bacteria should occur among particles smaller than 1 μm . Our
15 observations at a coastal site in Japan in spring, when the westerlies frequently blew dust there from the Asian continent, revealed that particle-attached bacteria in dust episodes, at the concentration of $3.2 \pm 2.1 \times 10^5$ cells m^{-3} , occupied 72 ± 9 % in the total bacteria. In contrast, the fraction was 56 ± 17 % during nondust periods and the concentration was $1.1 \pm 0.7 \times 10^5$ cells m^{-3} . The viability, defined as the ratio of viable cells to total cells, of particle-attached bacteria was 69 ± 19 % in dust episodes and 60 ± 22 % during nondust periods, both of which were considerably lower than the viabilities of free-floating bacteria (about
20 87 %) under either dusty or nondust conditions. The present cases suggest that dust particles carried substantial bacteria on their surfaces, more than half of which were viable, and spread these bacteria in the atmosphere. This implies that dust and bacteria have nonnegligible roles as internally mixed assemblages in various atmospheric processes and in linking geographically isolated microbial communities, as well as have synergistic effect on human health.



25 1 Introduction

Biological particles in the atmosphere have a significant potential effect on climate change (Ariya et al., 2009; Delort et al., 2010; Möhler et al., 2007; Zhang et al., 2017), efficiently link microbial communities between continents, islands and oceans (Fröhlich-Nowoisky et al., 2016; Morris et al., 2011; Caliz et al., 2018), and pose risks to human health (Polymenakou et al., 2008; Reinmuth-Selzle et al., 2017). Representing a high fraction of primary biological particles, airborne bacteria are emitted into the atmosphere from various sources, among which desert dust is a major source (Morris et al., 2011; Pöschl and Shiraiwa, 2015; Pöschl et al., 2010). The cooccurrence of dust and high concentrations of bacteria has been observed frequently in different locations, indicating the widespread nature and dissemination of bacteria with dust at local, regional and even global scales (Griffin, 2007; Hara and Zhang, 2012; Iwasaka et al., 2009). Limited available observations have revealed the coexistence of mineral and biological contents in ice crystals (Creamean et al., 2013; Pratt et al., 2009), and laboratory experiments have demonstrated that the ice nucleation ability of dust particles is enhanced by biological components, including bacteria in the particles (Boose et al., 2019; Tobo et al., 2019; Conen et al., 2011). Recent toxicological studies with mouse exposure found that the internal mixture of dust and pathogenic bacteria exacerbated pneumonia (He et al., 2012). In addition, the attachment of bacteria to dust particles is expected to largely alter the fate of bacterial cells in the air due to protection by the dust particles from harsh environmental conditions (Bowers et al., 2013) and enhanced gravitational settling (Zhang, 2008). All these results reflect that the adherence of bacterial cells to dust particles, i.e., the particle-attached state, and the viability or metabolic capability of bacterial cells are key factors affecting the roles and fate of airborne bacteria in the evolution, development and conservation of the natural environment.

Quantitative data on the mutual state of airborne bacteria and dust particles in dusty air are no doubt very scientifically interesting (Schuerger et al., 2018). However, quantitative data are rare because of a lack of available and confident methods for such research, leaving unidentifiable uncertainties in both field observations and model simulations exploring the activities and roles of bacterial cells in atmospheric processes. The cell size distributions for bacteria separated from soils have been investigated previously (Portillo et al., 2013). Whereas, airborne bacteria should have different survival mechanisms, dispersal and size distribution from bacteria in soils because of the aerosolization process from Earth surfaces and harsh atmospheric stressors. We quantified the fractions of particle-attached and free-floating bacterial cells in dusty and nondust air based on the fact that airborne bacterial cells are usually $\sim 1 \mu\text{m}$ or smaller than $1 \mu\text{m}$ (Delort et al., 2010; Després et al., 2012; Pósfai et al., 2003; Burrows et al., 2009; Hara et al., 2011); thus, particle-attached bacteria should be trapped in aerosol samples of particles larger than $1 \mu\text{m}$, and free-floating bacteria should be located among particles smaller than $1 \mu\text{m}$.

By utilizing 8-stage Andersen cascade impactors, size-segregated aerosol samples were collected at a southwestern coastal site of Japan in the spring of 2013–2016, when the middle latitude westerly wind in the Northern Hemisphere frequently brought dust from the Asian continent to the observation site. Viable and nonviable bacteria in each sample were counted using the LIVE/DEAD BacLight bacterial viability assay to estimate bacterial concentrations (Murata and Zhang, 2013, 2016).



Bacteria detected in samples of particles larger than 1.1 μm (the cutoff size of the sampler stages) were considered particle-attached bacteria, and those in the stages of particles smaller than 1.1 μm were considered free-floating bacteria. An analysis of method confidence showed that uncertainties due to the sample collection were small (Figs. S4 and S5 in the Supplement).
60 In this study, we focus on comparisons of the quantitative results of particle-attached and free-floating bacteria in the air and the viability of these bacteria under dust and nondust conditions.

2 Methods

2.1 Sample collection and cell enumeration

Aerosol samples were collected on the platform of a building (32.324°N, 129.993°E; 15 m above ground level and 23
65 m above sea level) on the seaside of Amakusa Island, southwestern Japan (Fig. S1) during several observational campaigns in the spring of 2013–2016. Dust plumes from the Asian continent, called Asian dust, frequently pass this area in spring. There are limited fishery and agriculture activities and few anthropogenic sources of air pollutants around the area, making the site suitable for investigating airborne bacteria in the Asian continental outflow (Murata and Zhang, 2016).

Aerosol samples were collected onto 0.2 μm pore polycarbonate filters (47 mm; Merck Millipore Ltd., Cork, Ireland)
70 with 8-stage Andersen cascade samplers (Model AN-200; Tokyo Dylec Corp., Japan). The flow rate of the samplers was 28.3 L min^{-1} . Aerosol particles were collected onto 8 filters according to the particle aerodynamic diameter ranges of >11, 7.0–11, 4.7–7.0, 3.3–4.7, 2.1–3.3, 1.1–2.1, 0.65–1.1 and 0.43–0.65 μm . The collection time of one set of samples was from approximately 3 to 24 h. Details on the sample collection are given in Table S1 and Fig. S2 in the Supplement.

Before the collection of each sample set, all stages of the sampler were cleaned carefully, and the plates for the filters
75 were rinsed and wiped with 70% ethanol in a clean hood to avoid contamination. A blank control for each set of samples was prepared, i.e., a blank filter was set in the sampler without sample collection. After sample collection, the filters were sealed in Petri dishes and stored at -20°C until analysis.

The viable and nonviable bacterial cells (Fig. S3) on the filters were enumerated using the LIVE/DEAD BacLight bacterial viability assay with an epifluorescence microscope (EFM; Eclipse 80i, Nikon Corp., Tokyo, Japan) as described
80 previously (Murata and Zhang, 2016, 2013). Fluorescent green and red/orange/yellow cells with spherical shape and size close to or smaller than 1 μm in diameter were counted as viable and nonviable bacteria, respectively. The cell concentrations in the size-segregated particles in the air were estimated based on cell counts and the sampling of air volumes following the subtraction of the blank controls. The viability of a group of bacterial cells was defined as the ratio of the viable bacterial cells to total bacterial cells. The procedure for the experimental operation and the formulations for the estimation of cell
85 concentrations are given in the Supplement (Text S1 in the Supplement).



The collection efficiency of airborne bacterial cells with Andersen samplers was evaluated by comparing the results using BioSamplers (SKC Inc., Eighty-Four, PA, US) and in-line filter holders (47 mm, Millipore Corp., Billerica, MA, US). The results show that the total bacterial concentration results of the Andersen sampler were generally consistent with those of the BioSamplers and the holders (Fig. S4).

90 2.2 Separation of particle-attached and free-floating bacteria

In this study, bacteria in the samples of stages with particles larger than 1.1 μm were considered particle-attached bacteria, and bacteria in the samples of stages with particles ranging from 0.43–1.1 μm were considered free-floating bacteria. The resuspension of bacteria trapped by upper stages and falling onto lower stages during sample collection may cause uncertainties in the size distribution of bacteria-associated particles and the separation of particle-attached and free-
95 floating bacteria.

The uncertainties were investigated in the laboratory (Text S2 in the Supplement). The fractions and concentrations of particle-attached bacteria obtained by the present method were potentially underestimated. But the underestimation did not significantly affect the size distributions of particle-attached bacteria, and, in particular, the underestimation of the concentrations of particle-attached bacterial cells was less than 10% on average (Fig. S5). In addition, the concentrations of
100 total bacterial cells quantified using the Andersen samplers were consistent ($100\pm 15\%$) with those quantified using the holders. This result indicates that bacteria smaller than 0.43 μm , which trapped with difficulty by the Andersen samplers, were a minor fraction of the free-floating bacteria.

2.3 Atmospheric conditions

During the observation periods, the number concentrations of size-segregated airborne particles
105 (>0.3 , >0.5 , >1.0 , >2.0 , and >5.0 μm in diameter) were monitored with optical particle counters (OPC, KC-01D in 2013 and KC-01E in 2014–2016, Rion Co., Ltd, Tokyo, Japan). In this study, fine particles are in the range of 0.3–1.0 μm , and those larger than 1.0 μm are referred to as coarse particles. Meteorological conditions, including temperature, pressure, relative humidity, precipitation, and wind speed and direction, were monitored with a weather transmitter (WXT520, Vaisala Inc., Helsinki, Finland). Airborne particle number concentrations and meteorological data during the observation periods are
110 summarized in Fig. S2 and Table S2.

On the basis of surface pressure and weather charts in the days before and after sample collection (Figs. S2 and S6), the air parcels on the synoptic scales from which samples were collected were categorized into four groups: prefront, postfront, approaching anticyclone, and anticyclone (Table S1). Details of the categorization are available in Murata and Zhang (2016).



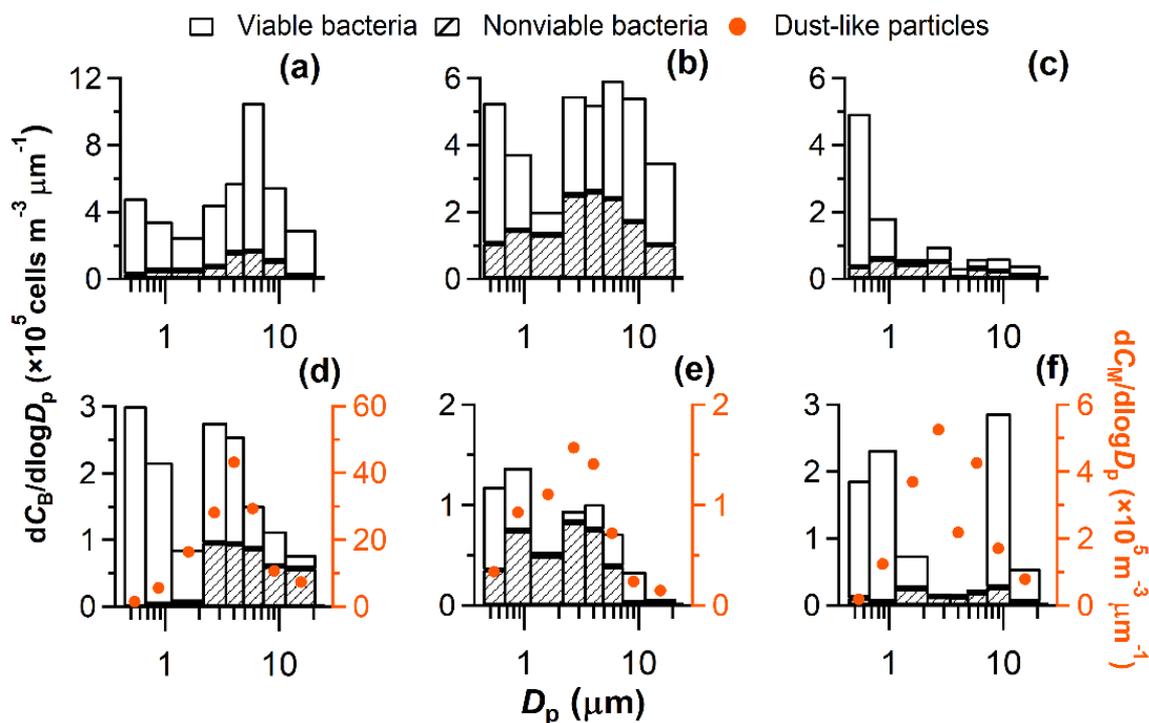
115 Dust episodes were identified by significant increases in coarse particle concentrations ($>1\ \mu\text{m}$), the forecast for
Asian dust distributions in the east Asian region (<http://www-cfors.nies.go.jp/~cfors/>; Fig. S7), and the backward trajectory
of air masses calculated with the NOAA/HYSPLIT model (http://ready.arl.noaa.gov/HYSPLIT_traj.php). During dust
events, the coarse particle concentration largely increased at the study site (Zhang et al., 2003). Dust particles were present in
the postfront air and sometimes in the approaching anticyclone air. The results of backward trajectory analysis during dusty
120 and nondust episodes are shown in Fig. S8.

3 Results

3.1 Concentrations of airborne bacteria in segregated size ranges

The concentrations of bacterial cells, including viable and nonviable cells, generally showed a bimodal number-size
distribution during dust episodes (e.g., Fig. 1a, b, d, f). Most of the bacteria were present in particle fractions with aerodynamic
125 size (D_p) ranges larger than $2\ \mu\text{m}$ (i.e., 2.1–3.3, 3.3–4.7 and 4.7–7.0 μm ; Fig. S9). These sizes are larger than the size of
individual airborne bacterial cells (approximately $1\ \mu\text{m}$ or smaller), indicating that the bacteria did not float individually in the
air but were combined with other particles, i.e., the bacteria were particle-attached. There were also many bacterial cells in the
size ranges smaller than $1.1\ \mu\text{m}$, i.e., free-floating bacterial cells. Their concentration was comparable to or lower than the
concentrations of bacteria in the larger size ranges (Figs. 1 and S9).

130 In contrast to dust episodes, during nondust periods, the number-size distribution of bacteria largely varied and did not
show any trend with respect to other particles or weather conditions. In seven cases during nondust periods (Samples 9, 11,
12, 13, 14, 21, and 23; Fig. S9), the bacteria appeared mainly in size ranges smaller than $1.1\ \mu\text{m}$ and accumulated the most in
the size range of 0.43–0.65 μm (e.g., Fig. 1c), indicating the predominance of free-floating bacteria. During most of the other
nondust periods (Samples 6, 7, 8, 16, 19, 20, 22, 24, and 25), the distributions of bacteria were similar to those during the dust
135 periods, although the concentrations were much lower than or comparable to those in the dust episodes (e.g., Fig. 1e). There
were two exceptional cases in nondust periods that had a mono-modal distribution, with peaks at 3.3–4.7 μm (Sample 15) or
larger than $11\ \mu\text{m}$ (Sample 18) (Fig. S9).



140 **Figure 1.** Concentrations of viable and nonviable bacteria (C_B) and mineral dust-like particles (C_M) in size-segregated airborne particles: (a) 19 March 2013, dusty; (b) 14 April 2013, dusty; (c) 21 March 2014, nondust; (d) 22 March 2015, dusty; (e) 22 March 2016, nondust; (f) 31 May 2016, dusty. The results of all sampling periods are depicted in Fig. S9 in the Supplement.

3.2 Concentration of particle-attached and free-floating bacteria

145 On average, the concentration of total bacterial cells, $4.4 \pm 2.6 \times 10^5$ cells m^{-3} , during dust episodes was much higher than that during nondust periods, $2.0 \pm 1.0 \times 10^5$ cells m^{-3} (Table 1). This high difference in concentration is consistent with the results of previous studies (Hara and Zhang, 2012; Yamaguchi et al., 2014). The concentrations of particle-attached bacterial cells during dust episodes and nondust periods were $3.2 \pm 2.1 \times 10^5$ and $1.1 \pm 0.7 \times 10^5$ cells m^{-3} , respectively. The percentage of particle-attached bacteria during dust periods, $72 \pm 9\%$, was much higher than that during nondust periods, $56 \pm 17\%$ (ANOVA test, $P < 0.05$).
 150 These results signify that dust particles carry a substantial amount of bacterial cells on their surfaces to remote downstream areas.



155

Table 1 Concentration and viability of total, free-floating, and particle-attached bacteria under dusty and nondust conditions. The concentration of coarse particles ($>1 \mu\text{m}$) and the ratio of particle-attached bacteria to coarse particles are also listed. The percentages of free-floating and particle-attached bacteria are given in parentheses.

Weather condition (Number of cases)	Coarse particles (10^6 m^{-3})	Total bacteria		Free-floating bacteria		Particle-attached bacteria (PAB)		
		Concentration ($10^5 \text{ cells m}^{-3}$)	Viability (%)	Concentration (10^5 cells m^{-3})	Viability (%)	Concentration (10^5 cells m^{-3})	Viability (%)	PAB/Coarse particles (%)
Dusty (9)	3.2 ± 2.5	4.4 ± 2.6	74 ± 17	1.2 ± 0.7 (28 ± 9)	87 ± 14	3.2 ± 2.1 (72 ± 9)	69 ± 19	16 ± 17
Nondust (18)	1.2 ± 0.5	2.0 ± 1.0	75 ± 13	0.9 ± 0.7 (44 ± 17)	87 ± 12	1.1 ± 0.7 (56 ± 17)	60 ± 22	10 ± 7
All (27)	1.8 ± 1.8	2.8 ± 2.0	74 ± 14	1.0 ± 0.7 (39 ± 16)	87 ± 12	1.8 ± 1.7 (61 ± 16)	63 ± 21	12 ± 11

160

On the other hand, the percentage of free-floating bacterial cells was higher than 70% in some cases during nondust periods (Table S2). In particular, the percentage ranged from 35% to 73% ($49 \pm 15\%$ on average) under anticyclone weather conditions, when the air parcels were from marine areas rather than from continental areas and moved stagnantly (Fig. S8). Therefore, there were a substantial amount of free-floating bacteria, and they were frequently the most common bacteria in nondust air.

165

The number ratio of particle-attached bacteria to particles in the size range larger than $1.1 \mu\text{m}$ was $12 \pm 11\%$ on average (Table S2). Except for two periods when the ratios were 35% and 59%, respectively, the ratio was approximately stable ($9 \pm 5\%$ on average for the other periods), regardless of dust episodes and nondust periods (Table S2). That is, assuming that a bacteria-attached coarse particle harbors at least one bacterial cell, coarse particles including mineral dust particles with attached bacteria usually made up less than 9% of the total coarse particles. Maki et al. (2008) reported that the mineral particles with attached bacteria made up approximately 10% of the total mineral particles, with the remaining mineral particles possessing few or no bacterial cells at 800-m height above the ground in an Asian dust source region, Dunhuang, China.

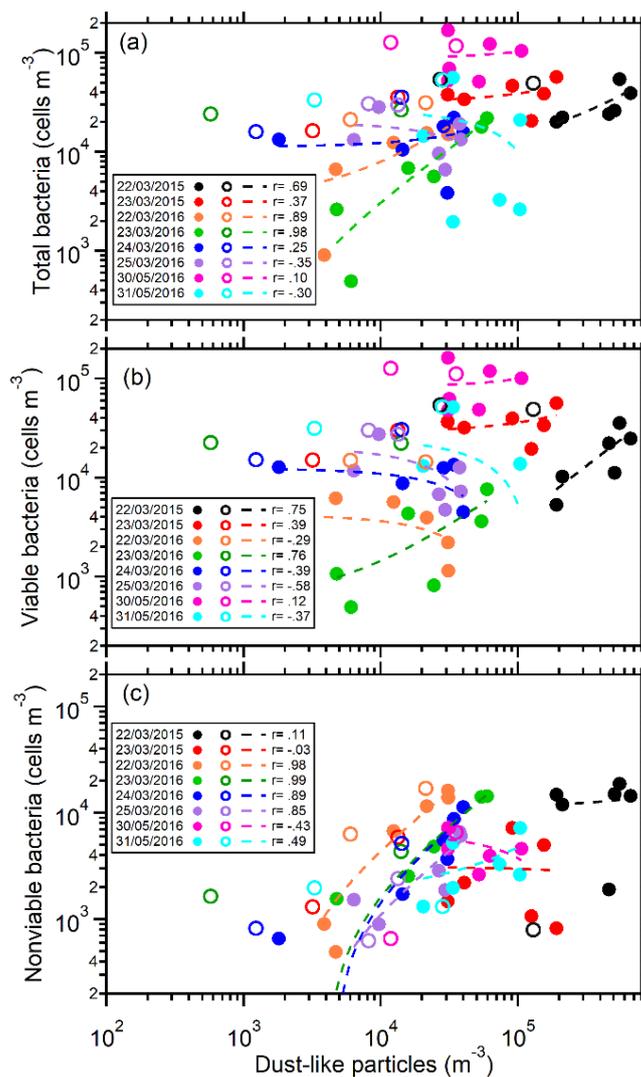
170

The number-size distributions of bacterial cells and mineral dust-like particles (insoluble and with irregular shapes; Fig. S3) in the microscope fields of some samples were compared. In most cases, the distributions (mode sizes) of particles and bacteria in the size ranges larger than $1.1 \mu\text{m}$ showed very good consistency (Figs. 1 and S9). The concentration of bacteria was usually closely correlated with the mineral dust-like particles in size-segregated samples (Fig. 2). These results further confirm that bacteria observed in the large size ranges were closely associated with airborne coarse particles, i.e., they were particle attached. In some cases, the mode size ranges of the bacterial cells and the dust-like particles were inconsistent (Fig. S9), likely because the number of bacteria on the surface of each coarse particle largely varied or there were less dust-like

175



particles (Sample 26). Dust-like particles were rarely observed in the size ranges smaller than $1.1 \mu\text{m}$ (Fig. S9), further indicating that the bacteria observed in those size ranges were predominantly free-floating.



180 **Figure 2.** Relationship between bacteria and mineral dust-like particles in size-segregated aerosols. (a) Total bacteria, (b) viable bacteria, and (c) nonviable bacteria. Solid and open circles represent particles larger and smaller than $1.1 \mu\text{m}$, respectively. Dashed lines represent unweighted linear fits for particles larger than $1.1 \mu\text{m}$.



3.3 Viabilities of particle-attached and free-floating bacteria

The viability of particle-attached bacteria varied over a wide range from 18% to 98% ($63 \pm 21\%$ on average), and the
185 viability of free-floating bacteria was between 56% and 99% ($87 \pm 12\%$), higher than the viability of particle-attached bacteria
(Tables 1 and S2). The attachment of airborne bacteria to larger particles is expected to be favorable for retaining the viability
or cultivability of cells and may indirectly increase the diversity of bacterial communities because of the possible protection
of bacterial cells from harsh atmospheric conditions (Bowers et al., 2013; Prospero et al., 2005; Lighthart, 2000).

However, we found that the viability of particle-attached bacteria was much lower than that of free-floating bacteria,
190 regardless of weather conditions (Table S2). This result indicates that a fraction of the particle-attached bacterial cells were
either nonviable when they were blown into the air with the dust or had experienced atmospheric stressors for several days
during long-distance transport and changed from a viable to a nonviable state. This is also likely the reason for the poor
correlation (Pearson correlation $r = 0.349$, $p = 0.075$) between the viability of particle-attached bacteria and the ratio of particle-
attached bacteria to coarse particles (Tables 1 and S2). In contrast, a large fraction of free-floating bacteria were viable. A
195 fraction of these bacteria were likely from local areas, with a residence time shorter than that of the particle-attached bacteria
transported from the Asian continent. In terms of concentration, viable particle-attached bacteria were usually more abundant
than viable free-floating bacteria in dust episodes (Figs. 1 and S9).

The viability ($74 \pm 17\%$) of total bacteria in dusty episodes was close to the viability ($75 \pm 13\%$) of total bacteria during
nondust periods (Table 1). The viability of particle-attached bacteria ($69 \pm 19\%$) during dust periods was slightly higher than
200 that ($60 \pm 22\%$) during nondust periods. The majority of particle-attached bacteria were viable.

Free-floating bacteria exhibited a quite high viability, and the viabilities of the bacteria in dusty ($87 \pm 14\%$) and nondust
($87 \pm 12\%$) air were similar. The concentration of viable free-floating bacteria was 3.8×10^4 – 1.5×10^5 cells m^{-3} , which was
lower than that of particle-attached bacteria (6.2×10^4 – 5.1×10^5 cells m^{-3}). An increase in viable free-floating bacteria on the
order of 10^5 cell m^{-3} was observed when the weather was fine and the air masses moved slowly from marine areas, favoring
205 the accumulation of bacteria emitted from local areas (Fig. S8).

4 Discussion

4.1 Comparison with literature data

There are few data on airborne bacterial cells available for comparison with the present study. Observations in the
multiphase atmosphere with culture-dependent methods revealed that approximately 60–90% or even more culturable airborne
210 bacteria were present in the size range of particles larger than $1.1 \mu m$ (Agarwal, 2017; Burrows et al., 2009; Montero et al.,
2016; Raisi et al., 2013), and the median aerodynamic diameter of particles containing culturable bacteria was approximately
2–4 μm at diverse sites (Lighthart, 2000; Raisi et al., 2013; Shaffer and Lighthart, 1997; Tong and Lighthart, 2000). These results



indicate the predominance of culturable particle-attached bacteria in the air, which is approximately in line with the results under dusty and nondust conditions of this study.

215 Early studies with single-particle analysis frequently encountered the mode size of biological aerosol particles in the
size range smaller than 1 μm (Matthias-Maser et al., 1999; Matthias-Maser and Jaenicke, 1995, 2000). In contrast, recent real-
time measurements using ultraviolet aerodynamic particle sizer spectrometers and wideband integrated bioaerosol sensor
techniques revealed the mode size of fluorescent biological aerosol particles (FBAP) to be approximately 2–6 μm , and the
particles were mainly attributed to fungal spores (Pöschl et al., 2010; Savage et al., 2017; Yue et al., 2017; Huffman et al., 2010).
220 However, the abundant particle-attached bacteria identified in this study in size ranges larger than 2 μm indicate dust-particle-
attached bacteria should not compose small fractions of real-time FBAP results in the relevant size ranges. In addition, the
mode at or smaller than 1 μm observed in real-time FBAP studies is likely consistent with the presence of free-floating bacterial
cells in the present study, but the comparison and discussion on the data are not confident because of the large uncertainties
caused by the low counting efficiency and accuracy in submicron size ranges of the instruments used in the studies (Yue et al.,
225 2017; Huffman et al., 2010).

4.2 Ice cloud formation

Dust particles from desert areas are constantly spread at local, regional and global scales in the atmosphere. These
particles transport microorganisms across continents and oceans to remote downstream areas (Griffin, 2007; Schuerger et al.,
2018). It has been shown that bacteria in the air are more effective ice nuclei at temperatures as warm as -2°C than abiotic
230 particles (Ariya et al., 2009; Burrows et al., 2013; Fröhlich-Nowoisky et al., 2016; Möhler et al., 2007). Biological particles
coexisting with dust particles have been detected in ice residues sampled from clouds (Creamean et al., 2013; Pratt et al., 2009),
and the coexistence of dust and bacterial cells increases the ability of particles to act as ice nuclei for ice crystal formation
(Tobo et al., 2019). Proteins in bacteria are ice nucleation active sites and are well protected when bacteria adhere to mineral
dust surfaces (Conen et al., 2011). The attachment of bacteria to dust particles possibly increases the number of sites for ice
235 nucleation and consequently the ice nucleation ability of dust particles (Boose et al., 2019; Conen et al., 2011). The present
results show that up to one-tenth or more dust particles could be bacteria carriers, and the concentration of particle-attached
bacteria, i.e., the number of bacteria-dust contact sites in dust episodes, was on average 3 times larger than that during nondust
periods (Table 1). The occurrence of dust in remote downstream areas will significantly increase not only the concentration of
bacterial cells but also the concentration of dust-bacteria mixture particles and the number of ice nucleation active sites. This
240 phenomenon could provide important sources of nuclei for ice cloud formation under saturated meteorological conditions for
icing, particularly in remote elevated air, where the concentrations of aerosol particles able to act as nuclei are usually very
low (Creamean et al., 2013).



4.3 Ecosystem conservation and development

245 More than 60% of particle-attached bacteria and approximately 87% of free-floating bacteria in the dusty air remained
viable. Airborne bacteria can multiply more easily after they settle into water (lakes, rivers and oceans) and soil surfaces than
in the atmosphere. As a consequence, their dissemination via the atmosphere has the potential to alter the microbial
biogeography, biogeochemistry and ecosystem services of downstream areas. Moreover, a recent study on phosphorus in
aerosol particles in Asian continental outflow revealed that natural dust particles supplied higher ratios of bioavailable
phosphorus than other types of particles as nutrients for the primary production in marine ecosystems, and the phosphorus was
250 presumed to be from the biological particles in dust plumes (Shi et al., 2019). The dissemination of bacteria with dust in the
air is much more efficient than that via other routes, such as rivers, because dust in the atmosphere can travel globally within
two weeks (Uno et al., 2009). Therefore, the wide dispersal of atmospheric dust is an efficient link between bacterial
communities in geographically isolated ecosystems. This linking function is likely the key process that constantly blurs the
distinctions between closely related microbial species in distant areas. Thus, the diversities of microorganisms have a
255 geographically weak gradient at the global scale, and are functions of habitat properties but not of historical/evolutionary
factors (Fenchel and Finlay, 2004).

4.4 Health effects

Allergenic and toxic bacteria inhaled and deposited on the surface of upper respiratory tracts and lungs are suggested
to provoke severe adverse health effects, regardless of whether the bacteria are viable, dead or cell fragments (Fröhlich-
260 Nowoisky et al., 2016; Després et al., 2007). Dust particles carrying biological materials, including bacteria with pathogenic,
allergenic, and adjuvant activity, can cause and aggravate respiratory disorders (Reinmuth-Selzle et al., 2017). The size
distribution of bacteria-related particles in the air is particularly meaningful because the movement and deposition of the
particles in the airways are size-dependent. Particles larger than 0.5 μm are deposited by sedimentation and impaction mainly
in the head airways, and particles smaller than 0.5 μm can reach the lower airways by diffusion (Fröhlich-Nowoisky et al.,
265 2016). According to the size distribution of the airborne bacteria-related particles in this study (Figs. 1 and S9), the deposition
fraction and abundance of particle-attached bacteria are much higher than those of individual cells in both the upper and the
lower airways. Polymenakou et al. (2008) reported that a large fraction of airborne bacteria at respiratory particle sizes (< 3.3
 μm) during an intense dust event were phylogenetic neighbors to human pathogens. He et al. (2012) suggested that Asian dust
caused the exacerbation of pneumonia induced by *Klebsiella pneumoniae* due to the enhanced production of pro-inflammatory
270 mediators in alveolar macrophages. Therefore, free-floating bacterial cells are likely to more easily influence the deep parts
than the upper parts of respiratory airways, while the negative influence of particle-attached bacteria, particularly under dust
conditions, is expected to be more serious in the upper parts than in the deep parts of respiratory airways.



5 Conclusions

In this study, we aimed to quantify the particle-attached and free-floating bacteria in dusty and nondust air in southwestern Japan using the fluorescent enumeration of bacterial cells in size-segregated aerosol samples. The bacteria showed bimodal number-size distributions during dust episodes, while the distributions largely varied during nondust periods. Particle-attached bacteria in dust episodes, with a concentration of $3.2 \pm 2.1 \times 10^5$ cells m^{-3} , occupied $72 \pm 9\%$ of the total bacteria. In contrast, this percentage was $56 \pm 17\%$ during nondust periods, with a concentration of $1.1 \pm 0.7 \times 10^5$ cells m^{-3} . The results indicate that dust particles conveyed substantial bacterial cells on their surfaces. Viable particle-attached bacteria were more abundant than viable free-floating bacteria in dusty air, which is compatible with the previous results that larger particles harbor more viable and/or culturable bacteria than smaller particles.

The viability (approximately $63 \pm 21\%$) of particle-attached bacteria was much lower than that ($87 \pm 12\%$) of free-floating bacteria, likely because atmospheric stressors along with long-distance transport inhibited the survival of particle-attached bacteria and the entrainment of locally originating free-floating bacteria. High concentrations and viabilities of free-floating bacteria were observed in stagnant air, mostly under anticyclone conditions, suggesting that locally emitted bacteria accounted for the major fractions.

The present results, quantitatively showing the state of airborne bacteria in association with particles, i.e., particle-attached and free-floating bacteria, could have broad implications in the disciplines of atmospheric sciences, ecology, public health and climate. In addition, the methods used in this study are low cost and easily available but are time- and labor-intensive. Verification of the status of airborne bacteria using efficient techniques, such as *in situ* electron microscopy, and the exploration of the compositions, functions and activities of particle-attached and free-floating bacteria in the atmosphere, are necessary to deepen our understanding of the related fields.

Data availability. All data are available from the corresponding author upon request.

Supplement. The supplement related to this article is available online.

Author contributions: DZ and WH designed research; WH, KM, CF and SH performed research; WH, KM and DZ analyzed data and wrote the paper; HM and PF reviewed and commented on the paper.

Competing interests. The authors declare that they have no conflict of interest.

Acknowledgments. This work was supported by the Japan Society for the Promotion of Science KAKENHI (JP16H02492, 17K18811), and the National Natural Science Foundation of China (41805118, 41977183). We thank Yuka Horikawa, Megumi Mukogawa and Miki Miyamoto for their assistance with sampling and analysis.



References

- Agarwal, S.: Seasonal variability in size-segregated airborne bacterial particles and their characterization at different source-sites, *Environ. Sci. Pollut. Res.*, 24, 13519-13527, 10.1007/s11356-017-8705-2, 2017.
- 305 Ariya, P. A., Sun, J., Eltouny, N. A., Hudson, E. D., Hayes, C. T., and Kos, G.: Physical and chemical characterization of bioaerosols – Implications for nucleation processes, *International Reviews in Physical Chemistry*, 28, 1-32, 10.1080/01442350802597438, 2009.
- Boose, Y., Baloh, P., Plötze, M., Ofner, J., Grothe, H., Sierau, B., Lohmann, U., and Kanji, Z. A.: Heterogeneous ice nucleation on dust particles sourced from nine deserts worldwide – Part 2: Deposition nucleation and condensation freezing, *Atmos. Chem. Phys.*, 19, 1059-1076, 10.5194/acp-19-1059-2019, 2019.
- 310 Bowers, R. M., Clements, N., Emerson, J. B., Wiedinmyer, C., Hannigan, M. P., and Fierer, N.: Seasonal variability in bacterial and fungal diversity of the near-surface atmosphere, *Environ. Sci. Technol.*, 47, 12097-12106, 10.1021/es402970s, 2013.
- Burrows, S. M., Elbert, W., Lawrence, M. G., and Pöschl, U.: Bacteria in the global atmosphere–Part 1: Review and synthesis of literature data for different ecosystems, *Atmos. Chem. Phys.*, 9, 9263-9280, 10.5194/acp-9-9263-2009, 2009.
- 315 Burrows, S. M., Hoose, C., Pöschl, U., and Lawrence, M. G.: Ice nuclei in marine air: biogenic particles or dust?, *Atmos. Chem. Phys.*, 13, 245-267, 10.5194/acp-13-245-2013, 2013.
- Caliz, J., Triado-Margarit, X., Camarero, L., and Casamayor, E. O.: A long-term survey unveils strong seasonal patterns in the airborne microbiome coupled to general and regional atmospheric circulations, *Proc Natl Acad Sci U S A*, 10.1073/pnas.1812826115, 2018.
- 320 Conen, F., Morris, C. E., Leifeld, J., Yakutin, M. V., and Alewell, C.: Biological residues define the ice nucleation properties of soil dust, *Atmos. Chem. Phys.*, 11, 9643-9648, 10.5194/acp-11-9643-2011, 2011.
- Creamean, J. M., Suski, K. J., Rosenfeld, D., Cazorla, A., DeMott, P. J., Sullivan, R. C., White, A. B., Ralph, F. M., Minnis, P., Comstock, J. M., Tomlinson, J. M., and Prather, K. A.: Dust and biological aerosols from the Sahara and Asia influence precipitation in the western U.S., *Science*, 339, 1572-1578, 10.1126/science.1227279, 2013.
- 325 Delort, A.-M., Väitilingom, M., Amato, P., Sancelme, M., Parazols, M., Mailhot, G., Laj, P., and Deguillaume, L.: A short overview of the microbial population in clouds: potential roles in atmospheric chemistry and nucleation processes, *Atmos. Res.*, 98, 249-260, 10.1016/j.atmosres.2010.07.004, 2010.
- Després, V. R., Nowoisky, J. F., Klose, M., Conrad, R., Andreae, M. O., and Pöschl, U.: Characterization of primary biogenic aerosol particles in urban, rural, and high-alpine air by DNA sequence and restriction fragment analysis of ribosomal RNA genes, *Biogeosciences*, 4, 1127-1141, 10.5194/bg-4-1127-2007, 2007.
- 330 Després, V. R., Huffman, J. A., Burrows, S. M., Hoose, C., Safatov, A. S., Buryak, G., Fröhlich-Nowoisky, J., Elbert, W., Andreae, M. O., and Pöschl, U.: Primary biological aerosol particles in the atmosphere: a review, *Tellus B Chem. Phys. Meteorol.*, 64, 15598, 10.3402/tellusb.v64i0.15598, 2012.
- Fenchel, T., and Finlay, B. J.: The ubiquity of small species: patterns of local and global diversity, *BioScience*, 54, 777-784, 10.1641/0006-3568(2004)054[0777:TUOSSP]2.0.CO;2, 2004.
- 335 Fröhlich-Nowoisky, J., Kampf, C. J., Weber, B., Huffman, J. A., Pöhlker, C., Andreae, M. O., Lang-Yona, N., Burrows, S. M., Gunthe, S. S., Elbert, W., Su, H., Hoor, P., Thines, E., Hoffmann, T., Després, V. R., and Pöschl, U.: Bioaerosols in the Earth system: climate, health, and ecosystem interactions, *Atmos. Res.*, 182, 346-376, 10.1016/j.atmosres.2016.07.018, 2016.
- 340 Griffin, D. W.: Atmospheric movement of microorganisms in clouds of desert dust and implications for human health, *Clin. Microbiol. Rev.*, 20, 459-477, 10.1128/CMR.00039-06, 2007.
- Hara, K., Zhang, D., Yamada, M., Matsusaki, H., and Arizono, K.: A detection of airborne particles carrying viable bacteria in an urban atmosphere of Japan, *Asian J. Atmos. Environ.*, 5, 152-156, 10.5572/ajae.2011.5.3.152, 2011.
- Hara, K., and Zhang, D.: Bacterial abundance and viability in long-range transported dust, *Atmos. Environ.*, 47, 20-25, 10.1016/j.atmosenv.2011.11.050, 2012.
- 345 He, M., Ichinose, T., Yoshida, S., Yamamoto, S., Inoue, K., Takano, H., Yanagisawa, R., Nishikawa, M., Mori, I., Sun, G., and Shibamoto, T.: Asian sand dust enhances murine lung inflammation caused by *Klebsiella pneumoniae*, *Toxicol Appl Pharmacol*, 258, 237-247, 10.1016/j.taap.2011.11.003, 2012.



- Huffman, J. A., Treutlein, B., and Pöschl, U.: Fluorescent biological aerosol particle concentrations and size distributions measured with an Ultraviolet Aerodynamic Particle Sizer (UV-APS) in Central Europe, *Atmos. Chem. Phys.*, 10, 3215-3233, 10.5194/acp-10-3215-2010, 2010.
- Iwasaka, Y., Shi, G.-Y., Yamada, M., Kobayashi, F., Kakikawa, M., Maki, T., Naganuma, T., Chen, B., Tobo, Y., and Hong, C.: Mixture of Kosa (Asian dust) and bioaerosols detected in the atmosphere over the Kosa particles source regions with balloon-borne measurements: possibility of long-range transport, *Air Qual. Atmos. Health*, 2, 29-38, 10.1007/s11869-009-0031-5, 2009.
- Lighthart, B.: Mini-review of the concentration variations found in the alfresco atmospheric bacterial populations, *Aerobiologia*, 16, 7-16, 10.1023/A:1007694618888, 2000.
- Maki, T., Susuki, S., Kobayashi, F., Kakikawa, M., Yamada, M., Higashi, T., Chen, B., Shi, G., Hong, C., and Tobo, Y.: Phylogenetic diversity and vertical distribution of a halobacterial community in the atmosphere of an Asian dust (KOSA) source region, Dunhuang City, *Air Qual. Atmos. Health*, 1, 81-89, 10.1007/s11869-008-0016-9, 2008.
- Matthias-Maser, S., and Jaenicke, R.: The size distribution of primary biological aerosol particles with radii > 0.2 μm in an urban/rural influenced region, *Atmos. Res.*, 39, 279-286, 10.1016/0169-8095(95)00017-8, 1995.
- Matthias-Maser, S., Brinkmann, J., and Schneider, W.: The size distribution of marine atmospheric aerosol with regard to primary biological aerosol particles over the South Atlantic Ocean, *Atmos. Environ.*, 33, 3569-3575, 10.1016/S1352-2310(98)00121-6, 1999.
- Matthias-Maser, S., and Jaenicke, R.: The size distribution of primary biological aerosol particles in the multiphase atmosphere, *Aerobiologia*, 16, 207-210, <https://doi.org/10.1023/A:1007607614544> 2000.
- Möhler, O., DeMott, P., Vali, G., and Levin, Z.: Microbiology and atmospheric processes: the role of biological particles in cloud physics, *Biogeosciences*, 4, 1059-1071, 10.5194/bg-4-1059-2007, 2007.
- Montero, A., Dueker, M. E., and O'Mullan, G. D.: Culturable bioaerosols along an urban waterfront are primarily associated with coarse particles, *PeerJ*, 4, e2827, 10.7717/peerj.2827, 2016.
- Morris, C. E., Sands, D. C., Bardin, M., Jaenicke, R., Vogel, B., Leyronas, C., Ariya, P. A., and Psenner, R.: Microbiology and atmospheric processes: research challenges concerning the impact of airborne micro-organisms on the atmosphere and climate, *Biogeosciences*, 8, 17-25, 10.5194/bg-8-17-2011, 2011.
- Murata, K., and Zhang, D.: Applicability of LIVE/DEAD BacLight stain with glutaraldehyde fixation for the measurement of bacterial cell concentration and viability in the air, *Aerosol Air Qual. Res.*, 13, 1755-1767, 10.4209/aaqr.2012.10.0293, 2013.
- Murata, K., and Zhang, D.: Concentration of bacterial aerosols in response to synoptic weather and land-sea breeze at a seaside site downwind of the Asian continent, *J. Geophys. Res. Atmos.*, 121, 11636-11647, 10.1002/2016jd025028, 2016.
- Polymenakou, P. N., Mandalakis, M., Stephanou, E. G., and Tselepidis, A.: Particle size distribution of airborne microorganisms and pathogens during an Intense African dust event in the eastern Mediterranean, *Environ. Health Perspect.*, 116, 292-296, 10.1289/ehp.10684, 2008.
- Portillo, M. C., Leff, J. W., Lauber, C. L., and Fierer, N.: Cell size distributions of soil bacterial and archaeal taxa, *Appl. Environ. Microbiol.*, 79, 7610-7617, 2013.
- Pöschl, U., Martin, S. T., Sinha, B., Chen, Q., Gunthe, S. S., Huffman, J. A., Borrmann, S., Farmer, D. K., Garland, R. M., Helas, G., Jimenez, J. L., King, S. M., Manzi, A., Mikhailov, E., Pauliquevis, T., Petters, M. D., Prenni, A. J., Roldin, P., Rose, D., Schneider, J., Su, H., Zorn, S. R., Artaxo, P., and Andreae, M. O.: Rainforest aerosols as biogenic nuclei of clouds and precipitation in the Amazon, *Science*, 329, 1513-1516, 10.1126/science.1191056, 2010.
- Pöschl, U., and Shiraiwa, M.: Multiphase chemistry at the atmosphere-biosphere interface influencing climate and public health in the anthropocene, *Chem. Rev.*, 115, 4440-4475, 10.1021/cr500487s, 2015.
- Pósfai, M., Li, J., Anderson, J. R., and Buseck, P. R.: Aerosol bacteria over the Southern Ocean during ACE-1, *Atmos. Res.*, 66, 231-240, 10.1016/s0169-8095(03)00039-5, 2003.
- Pratt, K. A., DeMott, P. J., French, J. R., Wang, Z., Westphal, D. L., Heymsfield, A. J., Twohy, C. H., Prenni, A. J., and Prather, K. A.: In situ detection of biological particles in cloud ice-crystals, *Nat. Geosci.*, 2, 398-401, 10.1038/ngeo521, 2009.
- Prospero, J. M., Blades, E., Mathison, G., and Naidu, R.: Interhemispheric transport of viable fungi and bacteria from Africa to the Caribbean with soil dust, *Aerobiologia*, 21, 1-19, 10.1007/s10453-004-5872-7, 2005.



- Raisi, L., Aleksandropoulou, V., Lazaridis, M., and Katsivela, E.: Size distribution of viable, cultivable, airborne microbes and their relationship to particulate matter concentrations and meteorological conditions in a Mediterranean site, *Aerobiologia*, 29, 233-248, 10.1007/s10453-012-9276-9, 2013.
- 400 Reinmuth-Selzle, K., Kampf, C. J., Lucas, K., Lang-Yona, N., Frohlich-Nowoisky, J., Shiraiwa, M., Lakey, P. S. J., Lai, S., Liu, F., Kunert, A. T., Ziegler, K., Shen, F., Sgarbanti, R., Weber, B., Bellinghausen, I., Saloga, J., Weller, M. G., Duschl, A., Schuppan, D., and Pöschl, U.: Air Pollution and Climate Change Effects on Allergies in the Anthropocene: Abundance, Interaction, and Modification of Allergens and Adjuvants, *Environ. Sci. Technol.*, 51, 4119-4141, 10.1021/acs.est.6b04908, 2017.
- 405 Savage, N. J., Krentz, C. E., Könemann, T., Han, T. T., Mainelis, G., Pöhlker, C., and Huffman, J. A.: Systematic characterization and fluorescence threshold strategies for the wideband integrated bioaerosol sensor (WIBS) using size-resolved biological and interfering particles, *Atmos. Meas. Tech.*, 10, 4279-4302, 10.5194/amt-10-4279-2017, 2017.
- Schuerger, A. C., Smith, D. J., Griffin, D. W., Jaffe, D. A., Wawrik, B., Burrows, S. M., Christner, B. C., Gonzalez-Martin, C., Lipp, E. K., Schmale Iii, D. G., and Yu, H.: Science questions and knowledge gaps to study microbial transport and survival in Asian and African dust plumes reaching North America, *Aerobiologia*, 34, 425-435, 10.1007/s10453-018-9541-7, 2018.
- 410 Shaffer, B. T., and Lighthart, B.: Survey of culturable airborne bacteria at four diverse locations in Oregon: urban, rural, forest, and coastal, *Microb. Ecol.*, 34, 167-177, 10.1007/s002489900046, 1997.
- Shi, J., Wang, N., Gao, H., Baker, A. R., Yao, X., and Zhang, D.: Phosphorus solubility in aerosol particles related to particle sources and atmospheric acidification in Asian continental outflow, *Atmos. Chem. Phys.*, 19, 847-860, 10.5194/acp-19-847-2019, 2019.
- 415 Tobo, Y., Adachi, K., DeMott, P. J., Hill, T. C. J., Hamilton, D. S., Mahowald, N. M., Nagatsuka, N., Ohata, S., Uetake, J., Kondo, Y., and Koike, M.: Glacially sourced dust as a potentially significant source of ice nucleating particles, *Nat. Geosci.*, 12, 253-258, 10.1038/s41561-019-0314-x, 2019.
- 420 Tong, Y., and Lighthart, B.: The annual bacterial particle concentration and size distribution in the ambient atmosphere in a rural area of the Willamette Valley, Oregon, *Aerosol Sci. Technol.*, 32, 393-403, 10.1080/027868200303533, 2000.
- Uno, I., Eguchi, K., Yumimoto, K., Takemura, T., Shimizu, A., Uematsu, M., Liu, Z., Wang, Z., Hara, Y., and Sugimoto, N.: Asian dust transported one full circuit around the globe, *Nat. Geosci.*, 2, 557-560, 10.1038/ngeo583, 2009.
- 425 Yamaguchi, N., Ichijo, T., Baba, T., and Nasu, M.: Long-range transportation of bacterial cells by Asian dust, *Genes and Environment*, 36, 145-151, 10.3123/jemsge.2014.015, 2014.
- Yue, S., Ren, H., Fan, S., Wei, L., Zhao, J., Bao, M., Hou, S., Zhan, J., Zhao, W., Ren, L., Kang, M., Li, L., Zhang, Y., Sun, Y., Wang, Z., and Fu, P.: High abundance of fluorescent biological aerosol particles in winter in Beijing, China, *ACS Earth Space Chem.*, 1, 493-502, 10.1021/acsearthspacechem.7b00062, 2017.
- 430 Zhang, D., Iwasaka, Y., Shi, G., Zang, J., Matsuki, A., and Trochkin, D.: Mixture state and size of Asian dust particles collected at southwestern Japan in spring 2000, *J. Geophys. Res. Atmos.*, 108, 4760, 10.1029/2003jd003869, 2003.
- Zhang, D.: Effect of sea salt on dust settling to the ocean, *Tellus B Chem. Phys. Meteorol.*, 60B, 641-646, 10.1111/j.1600-0889.2008.00358.x, 2008.
- 435 Zhang, D., Murata, K., Hu, W., Yuan, H., Li, W., Matsusaki, H., and Kakikawa, M.: Concentration and viability of bacterial aerosols associated with weather in Asian continental outflow: current understanding, *Aerosol Sci. Eng.*, 1, 66-77, 10.1007/s41810-017-0008-y, 2017.