



1 **Title:** The relative importance of photodegradation and biodegradation of terrestrially derived  
2 dissolved organic carbon across four lakes of differing trophic status

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48 **Abstract**

49 Outgassing of carbon dioxide (CO<sub>2</sub>) from freshwater ecosystems comprises 12-25% of the total  
50 carbon flux from soils and bedrock. This CO<sub>2</sub> is largely derived from both biodegradation and  
51 photodegradation of terrestrial dissolved organic carbon (DOC) entering lakes from wetlands and  
52 soils in the watersheds of lakes. In spite of the significance of these two processes in regulating  
53 rates of CO<sub>2</sub> outgassing, their relative importance remains poorly understood in lake ecosystems.  
54 In this study, we used groundwater from the watersheds of one subtropical and three temperate  
55 lakes of differing trophic status to simulate the effects of increases in terrestrial DOC from storm  
56 events. We assessed the relative importance of biodegradation and photodegradation in oxidizing  
57 DOC to CO<sub>2</sub>. We measured changes in DOC concentration, the optical characteristics of the  
58 DOC (SUVA<sub>320</sub> and S<sub>r</sub>), dissolved oxygen, and dissolved inorganic carbon (DIC) in short-term  
59 experiments from May-August, 2016. In all lakes, photodegradation led to larger changes in  
60 DOC and DIC concentrations and optical characteristics than biodegradation. A descriptive  
61 discriminant analysis showed that in brown-water lakes, photodegradation led to the largest  
62 declines in DOC concentration. In these brown-water systems, ~30% of the DOC was processed  
63 by sunlight and ~2% was photo mineralized. In addition to documenting the importance of  
64 photodegradation in lakes, these results also highlight how lakes in the future may respond to  
65 changes in DOC inputs.

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## 68 **Introduction**

69           Lakes are closely linked to their surrounding terrestrial ecosystems. As the lowest point  
70 in the landscape, they receive a significant influx of terrestrially-derived dissolved organic  
71 carbon (DOC) and nutrients (Williamson et al., 2009; Wilkinson et al., 2013). Climate and land  
72 use changes are altering the link between lakes and their surrounding landscapes by  
73 strengthening the flow of material during extreme rain events and large wildfires, or weakening  
74 it during extended periods of drought (Strock et al., 2016; Williamson et al., 2016). Long-term  
75 changes in DOC concentrations are variable and appear to be regionally controlled. In  
76 northeastern North American and western European lakes, there has been as much as a doubling  
77 of DOC concentrations due to recovery from anthropogenic acidification and climate change  
78 (Monteith et al., 2007; Williamson et al., 2015; de Wit et al., 2016). However, DOC  
79 concentrations in Greenland lakes (Saros et al., 2015) and the Mississippi River (Duan et al.,  
80 2017) have been decreasing. A long-term study of the Florida Everglades showed that some  
81 study sites were decreasing in DOC concentration, but the majority of sites were not changing  
82 (Julian et al., 2017). As DOC inputs into aquatic ecosystems have increased, stabilized, or  
83 decreased, long-term studies have focused on understanding the mechanisms behind the change,  
84 but less research has addressed the fate of DOC once it enters a lake.

85           By attenuating light in the water column and also providing a source of energy, DOC  
86 serves an important role in lakes by regulating the balance between photosynthesis and  
87 respiration (Williamson et al., 1999), and thus the flux of CO<sub>2</sub> to the atmosphere (Cole et al.,  
88 1994). Previous studies indicated that most lakes are net heterotrophic, where the breakdown of  
89 organic carbon exceeds production (Kling et al., 1991; Cole et al., 1994). Estimates suggest that  
90 lakes respire about half of the annual 2 gigaton flux of carbon to the oceans each year as CO<sub>2</sub>



91 (Cole et al., 1994; Tranvik et al., 2009; Tranvik, 2014). The traditional paradigm has been that  
92 the dominant mechanism causing the release of excess CO<sub>2</sub> from lakes is bacterial respiration of  
93 DOC (biodegradation), with photomineralization accounting for only 10% of bacterial rates  
94 (Granéli et al., 1996; del Giorgio et al., 1997; Jonsson et al., 2001). However, research on over  
95 200 Arctic lakes, rivers, and streams revealed that sunlight dominated the processing of DOC,  
96 and photomineralization rates were on average 5x greater than dark bacterial respiration rates  
97 (Cory et al., 2014). In addition, the source of inland water CO<sub>2</sub> remains uncertain (Raymond et  
98 al., 2013; Lapierre et al., 2013; Weyhenmeyer et al., 2015) and predicting DOC reactivity has  
99 been challenging (Evans et al., 2017). Quantifying the dominant degradation pathways for  
100 terrestrial DOC from a range of lakes will improve estimates of carbon fluxes, particularly  
101 mineralization rates that currently have a high degree of uncertainty (Hanson et al., 2014).

102 Many past studies have focused on testing the effects of photodegradation and  
103 biodegradation on DOC quantity individually, but they have not simultaneously evaluated how  
104 these two processes alter the absorbance characteristics of DOC, hereafter referred to as colored  
105 dissolved organic matter (CDOM) (Granéli et al., 1996; Koehler et al., 2014; Vachon et al.,  
106 2016a). The effects of sunlight on DOC are not isolated to only increasing mineralization rates;  
107 photodegradation can also decrease the color and molecular weight of DOC, which can increase  
108 light availability and the subsequent bacterial respiration of DOC (Bertilsson and Tranvik, 2000;  
109 Amado et al., 2003; Chen and Jaffé, 2016). Cory et al. (2014) found the dominant degradation  
110 process for Arctic lakes to be partial photodegradation, suggesting that in lakes, sunlight-driven  
111 changes in CDOM without undergoing complete mineralization may dominate DOC processing.

112 Since light attenuation varies so strongly among lakes of differing trophic status, testing  
113 the relative importance of DOC processing via photodegradation or biodegradation with



114 mechanistic experiments is needed. Previous research on DOC degradation has primarily  
115 occurred in high DOC lakes, but in clear-water lakes, 1% of surface UV-A and  
116 photosynthetically active radiation (PAR), which are the primary wavelengths active in  
117 photodegradation (Osburn et al., 2001), may reach up to 5-7 m for UV-A and 12-14 m for PAR  
118 in oligotrophic lakes and depths of >45-50 m in some of the clearest lakes in the world, such as  
119 Lake Tahoe (Rose et al., 2009a; Rose et al., 2009b). Geographic location and time of year  
120 influence the amount of solar radiation lakes receive. In the subtropics, PAR and UV light have  
121 high intensity across the spectrum year-around, whereas in temperate regions those wavelengths  
122 are strongest during the summer months.

123 Watershed land use and lake trophic status have also been shown to influence DOC  
124 composition and reactivity (Lu et al., 2013; Hosen et al., 2014; Larson et al., 2014; Evans et al.,  
125 2017). DOC from forested systems was more reactive when compared to disturbed environments  
126 and had different optical properties (Lu et al., 2012; Williams et al., 2015; Evans et al., 2017).  
127 Studies examining how terrestrial DOC inputs are processed in lakes are needed, especially with  
128 the increasing frequency of extreme rain events (Rahmstorf and Coumou, 2011; Westra et al.,  
129 2014; Fischer and Knutti, 2015). Future climate change projections suggest that for northern  
130 ecosystems a 10% increase in precipitation could lead to a 30% increase in the mobilization of  
131 soil organic matter (de Wit et al., 2016). Extreme rain events deliver fresh DOC not exposed to  
132 prior sunlight into lakes, which can lead to significant reductions in light availability, as well as  
133 increases in thermal stability and lake heterotrophy (Jennings et al., 2012; Klug et al., 2012; de  
134 Eyto et al., 2016; Zwart et al., 2016). As DOC concentrations change globally, understanding the  
135 processes that determine the fate of DOC will help predict the systems most likely to release  
136 more CO<sub>2</sub>.



137 Here our aim was to 1) determine the relative importance of photodegradation and  
138 biodegradation for altering terrestrial DOC quantity and CDOM from lakes of varying trophic  
139 status, 2) quantify the percentage of the initial DOC pool that was photomineralized, partially-  
140 photodegraded, biodegraded or remained unprocessed, and 3) compare the effects of  
141 photodegradation on DOC quantity and CDOM across four lakes to understand differences in  
142 how terrestrial DOC from the watersheds of different lake types responds to photodegradation.  
143 Since lakes are closely linked to their surrounding landscape (i.e. soils and vegetation), we  
144 collected terrestrial DOC from the watershed of three temperate lakes and one subtropical lake,  
145 all varying in trophic status. This soil organic matter represents the current and future inputs of  
146 organic material. We studied changes in the concentration of DOC, dissolved inorganic carbon  
147 (DIC), and dissolved oxygen (DO) and measured changes in CDOM. We hypothesized that  
148 photodegradation would be more important than biodegradation in all lakes, but the strongest  
149 responses to sunlight would be observed in the brown-water lakes.

150

## 151 **1. Methods**

### 152 **1.1 Study Sites and Samplers**

153 Groundwater samples were collected from the watersheds immediately adjacent to four  
154 lakes used in this study (Table 1). All of the lakes are small, with a surface area  $\leq 0.48$  km<sup>2</sup> and a  
155 maximum depth ranging from 12.5 m in Lake Waynewood to 24 m in Lake Giles. The three  
156 temperate lakes (Giles – oligotrophic; Lacawac – brown-water; Waynewood – eutrophic) are in  
157 close proximity, located on the Pocono Plateau in northeastern Pennsylvania. Lake Annie  
158 (brown-water) is a subtropical, sinkhole lake located on the Lake Wales Ridge in south-central  
159 Florida. These lakes were selected because of their variability in the dominant vegetation types



160 in their watersheds that lead to differences in DOC concentration and quality (Table 1). Annie,  
161 Giles, and Lacawac are all seepage lakes within protected watersheds, and there have been no  
162 significant changes in land use or land cover over the past thirty years. The watersheds of Giles  
163 and Lacawac have > 90% cover of mixed and northern hardwood-conifer forests, with oak trees  
164 dominating the watershed at Giles, while hemlocks represent the highest proportion of  
165 Lacawac's watershed (Moeller et al., 1995). Annie is surrounded by well-drained sandy soils and  
166 the major vegetation types include a mixed-scrub community, pinelands, and oak forests (Gaiser,  
167 2009). Both Annie and Lacawac are brown-water lakes with moderate DOC concentrations and  
168 lower transparency (Table 1). A higher percentage of wetlands (7% for Annie and 25% for  
169 Lacawac) in their watersheds likely contribute to their darker color compared to the other lakes  
170 (Moeller et al., 1995; H. Swain *unpublished data*). Waynewood is the most eutrophic lake and  
171 has the largest watershed with runoff from dairy farms upstream that feeds into the lake through  
172 an inlet stream. The forest surrounding Waynewood is similarly dominated by oak and hemlock  
173 trees, but there is overall less total forest cover in the watershed than Lacawac and Giles, and  
174 there are more homes adjacent to the lake (Moeller et al., 1995).

175



176 **Table 1.** Summary characteristics of the four study lakes in May-August 2013–2016 (mean ±  
 177 SD). Abbreviations: Chl-*a* (chlorophyll-a), DOC (dissolved organic carbon), GW DOC (initial  
 178 groundwater DOC), PAR (photosynthetically active radiation, 400-700 nm), UV-A (ultraviolet  
 179 A radiation, 380 nm), UV-B (ultraviolet B radiation, 320 nm), RT (residence time).

Lake	Lat. (°)	Long. (°)	Lake area (km <sup>2</sup> )	Max. depth (m)	Chl- <i>a</i> (µg L <sup>-1</sup> ) ± (SD)	Lake DOC (mg L <sup>-1</sup> ) ± (SD)	GW DOC (mg L <sup>-1</sup> ) ± (SD)	pH ± (SD)	1% UV-B depth (m) ± (SD)	1% UV-A depth (m) ± (SD)	1% PAR depth (m) ± (SD)	RT (yr)
Lacawac	41° 22' N	75° 17' W	0.21	13	1.9 (1.4)	5.2 (0.8)	59.4 (6.1)	6.6 <sup>+</sup>	0.4 (0.1)	0.9 (0.2)	5.7 (0.6)	3.3
Annie	27° 12' N	81° 20' W	0.36	20.7	4.0 (1.5)	9.4 (2.5)	20.7 (0.5)	5.5 (0.3)	0.5*	1.3*	4.5 (1.6)	2
Wayne-wood	41° 23' N	75° 21' W	0.28	12.5	5.3 (3.7)	6.4 (1.0)	7.6 (0.3)	7.5 <sup>+</sup>	0.3 (0.1)	0.7 (0.2)	4.3 (0.9)	0.42
Giles	41° 22' N	75° 5' W	0.48	24	1.1 (0.7)	2.3 (0.3)	6.0 (0.6)	6.2 <sup>+</sup> (0.3)	2.0 (0.5)	4.7 (1.2)	14.4 (2.1)	5.6

180 \*Indicates estimates from a single profile in March 2012. <sup>+</sup>pH data in Lacawac and Waynewood  
 181 are from 2015 only and from 2015-2016 in Giles.

182  
 183 Samplers were used to collect groundwater as a proxy for terrestrial DOC runoff entering  
 184 the lakes. The samplers were installed in close proximity to the Pocono lakes near small inlet  
 185 streams in sandy or bog areas on 6 July 2015 (~1 year prior to experiments). The groundwater  
 186 sampler consisted of 1m sections of 7.6cm diameter PVC pipe installed to a depth of 60-81cm  
 187 below ground. 0.5cm holes were drilled in the sides with a fine mesh covering the holes to let  
 188 shallow groundwater in but exclude large particulates. At Lake Annie, a groundwater sampler  
 189 was installed on 17 March 2016 on the south side of the lake near a small, intermittent inlet  
 190 stream. The groundwater sampler near Lake Annie was a 3m section of PVC pipe installed  
 191 slightly deeper to 2m below ground to allow continuous access to groundwater during the dry  
 192 season.



193 On 7 May 2016, 10 L of water was collected using a peristaltic pump from the  
194 groundwater samplers at all of the Pocono lakes in acid-washed 18 L bottles. Groundwater  
195 samples from Annie were collected from the sampler monthly (25 April, 31 May, 27 June, and 1  
196 Aug 2016) prior to starting the experiments and shipped overnight on ice to Pennsylvania. All  
197 groundwater samples were kept cold (4 °C) and dark until filtered to avoid sunlight exposure  
198 prior to the start of the experiments. Samples for the May experiments were filtered on May 8,  
199 2016 through a 0.7 µm Whatman GF/F filter. The remaining 8 L of groundwater for the June,  
200 July, and August experiments for each Pocono lake were filtered in a similar manner over the  
201 next 14 days. Samples were kept cold and dark until the experiments started. Samples for June,  
202 July, and August were re-filtered with a 0.7 µm Whatman GF/F filter prior to the start of those  
203 experiments. The initial DOC concentration of the groundwater for each lake varied at the start  
204 of each experiment, but it was always higher than the in-lake DOC concentration (Table 1).

205

## 206 **1.2 Sampling Design and Variables Analyzed**

207 To determine the relative importance of photodegradation and biodegradation for  
208 processing DOC, we designed three treatments in a manner similar to Cory et al., (2014): 1)  
209 photodegradation only, 2) biodegradation only, and 3) control. From each treatment, five  
210 different variables were measured including DOC concentration, DIC concentration, DO  
211 concentration, SUVA<sub>320</sub>, and S<sub>r</sub>. The different variables measured in each treatment required the  
212 use of different containers for the sample water. Samples for DOC analysis (concentration and  
213 CDOM) were deployed in acid-washed, muffled 35 mL quartz tubes sealed with silicone  
214 stoppers. The quartz tubes had an average transmittance of 96% of solar UV-A and 87% of solar  
215 UV-B, which allowed for an accurate representation of *in-situ* solar radiation levels (SFig. 1,



216 Morris and Hargreaves, 1997). However, the quartz tubes were not gas tight, so samples for  
217 dissolved inorganic carbon (DIC) and dissolved oxygen (DO) analysis were deployed in gas tight  
218 borosilicate vials (Labco, Ceredigion, UK). The borosilicate vials had a volume of 12 mL but  
219 were filled to 10 mL due to safety concerns with mercury chloride (see below). A clean 10 mL  
220 pipette was used to carefully transfer water into the borosilicate vials. Borosilicate glass has a  
221 sharp cut-off at 320 nm and transmits <5% UV-B, but it transmits an average of 63% of UV-A  
222 radiation and 90% of PAR (SFig. 1, Reche et al., 1999).

223 Water samples for all of the treatments were initially filtered through ashed 0.7  $\mu\text{m}$   
224 Whatman GF/F filters one day prior to the start of each monthly experiment. For the  
225 photodegradation and control treatments detailed below, samples for DO and DIC analysis were  
226 treated with 0.35 mL of 1% mercury chloride ( $\text{HgCl}_2$ ) to kill the microbial community. Samples  
227 for DOC concentration and CDOM analysis ( $\text{SUVA}_{320}$  and  $S_r$ ) for the same treatments were  
228 sterile filtered with a 0.2  $\mu\text{m}$  membrane filter (Sterivex MilliporeSigma, Burlington, MA USA)  
229 pre-rinsed with 100 mL of DI water and 50 mL of sample water instead of using  $\text{HgCl}_2$  because  
230 adding  $\text{HgCl}_2$  altered the optical scans. Sterile filtering has previously been shown to remove the  
231 majority of microbes present, and water samples remained sterile for one week following this  
232 procedure (Moran et al., 2000; Fasching and Battin, 2011). For the biodegradation treatment,  
233 water samples were inoculated with 100  $\mu\text{L}$  of unfiltered groundwater that was collected 1 day  
234 prior to the start of each monthly experiment. By adding a fresh inoculum of groundwater each  
235 month, we aimed to re-stimulate the microbial community and assess the short-term response of  
236 biodegradation. Treatments were deployed in triplicate for each lake (i.e. 3 DOC quartz tubes, 3  
237 DO borosilicate vials, and 3 DIC borosilicate vials for each treatment). Here, we included a  
238 summary of the three experimental treatments that were designed as follows:



239 a) *Photodegradation Only*: Water for DOC concentration and CDOM analysis (SUVA<sub>320</sub>  
240 and S<sub>r</sub>) was sterile filtered and stored in quartz tubes (n = 3 replicates). Water for DIC  
241 and DO analysis was treated with 1% HgCl<sub>2</sub> and stored in borosilicate vials (n = 6  
242 replicates; 3 replicates for DIC and 3 replicates for DO analysis).

243 b) *Biodegradation Only*: Water for all analyses was inoculated with 100 µL of unfiltered  
244 groundwater. Water samples for DOC concentration and CDOM analysis were stored in  
245 quartz tubes (n = 3 replicates). Water samples for DIC and DO analysis were stored in  
246 borosilicate vials (n = 6 replicates; 3 replicates for DIC and 3 replicates for DO analysis).  
247 Both the quartz tubes and borosilicate vials were wrapped with multiple layers of  
248 aluminum foil to eliminate light exposure.

249 c) *Control*: Water for DOC concentration and CDOM analysis was sterile filtered and  
250 stored in quartz tubes (n = 3 replicates). Water for DIC and DO analysis was treated with  
251 1% HgCl<sub>2</sub> and stored in borosilicate vials (n = 6 replicates; 3 replicates for DIC and 3  
252 replicates for DO analysis). All samples were wrapped in aluminum foil (dark).

253

254 The experimental treatments for each lake were deployed for seven days at the surface of  
255 Lake Lacawac in May, June, July, and August of 2016 (for exact sampling dates see SI, Table 1).  
256 Samples were kept at the lake surface using floating racks, and samples from each lake were  
257 randomly distributed across the racks. The deployment design ensured that samples stayed at the  
258 surface and dipped no deeper than 2 cm in the water column. After the one-week exposure, racks  
259 were collected from the surface of Lake Lacawac and samples were immediately transferred into  
260 coolers and returned to the lab. We assessed the response of terrestrially derived DOC to  
261 photodegradation and biodegradation by measuring changes in the concentrations of DOC, DIC,



262 and DO, and the absorbance properties ( $SUVA_{320}$  and  $S_r$ ) of the CDOM. All samples were  
263 analyzed within 72 hours of collection.

264 Dissolved organic carbon concentrations were analyzed using a Shimadzu TOC-V<sub>CPH</sub>  
265 Total Organic Analyzer with an ASI-V auto sampler. External acidification was used for each  
266 sample and triplicate measurements were performed following the methods of Sharp (1993).  
267 Dissolved inorganic carbon concentrations (as CO<sub>2</sub>) were measured with a Shimadzu GC-8A  
268 Gas Chromatograph using helium as the carrier gas. Samples were acidified using 0.1 N H<sub>2</sub>SO<sub>4</sub>  
269 and then stripped with nitrogen gas prior to injection. Dissolved oxygen was measured using a  
270 modified Winkler titration (Parson et al., 1984). Samples for gas measurements (DO and DIC)  
271 were kept in a 21°C water bath for 30 minutes prior to analysis. These samples were well mixed  
272 just prior to analysis. The absorbance properties of CDOM were analyzed using a Shimadzu UV  
273 1800 scanning spectrophotometer at 25°C. Raw absorbance scans were generated from 800 to  
274 200 nm using a 1 cm cuvette and were blank corrected with ultra-pure DI water. From the  
275 absorbance scans, the spectral slope ratio ( $S_r$ ) was calculated following Helms et al., (2008). The  
276 DOC specific ultraviolet absorbance at 320 nm ( $SUVA_{320}$ ) was calculated following methods in  
277 Williamson et al., (2014).  $S_r$  can be used as a proxy for the molecular weight of the DOC, while  
278  $SUVA_{320}$  can be used as a proxy for DOC color and aromatic carbon content (Helms et al., 2008,  
279 Williamson et al., 2014).

280 Due to differences between the borosilicate vials and quartz tubes, the DIC and DO  
281 samples were spectrally corrected for the amount of light they received (SI, SFig. 1). Total  
282 cumulative energy exposure over the monthly incubations was calculated from a BSI Model  
283 GUV-521 (Biospherical Instruments, San Diego, CA) radiometer with cosine irradiance sensors  
284 that have a nominal bandwidth of 8 nm for 305 nm, 320 nm, 340 nm, 380 nm, and 400-700 nm



285 (PAR). Daily irradiance for UV-B, UV-A, and PAR were calculated using 15-minute averages of  
286 1-second readings from a GUV radiometer located near Lake Lacawac over the 7-day  
287 experiments. The area under the curve was calculated by multiplying the measurement frequency  
288 (900 sec) by the average of two adjacent time step readings. These values were then summed  
289 over the exposure period to calculate the total cumulative energy exposure for each sample.  
290 Readings from a profiling BIC sensor (Biospherical Instruments, San Diego, CA) were then used  
291 to calculate the percent of the deck cell at the surface rack incubation depth (0.02 m) in Lake  
292 Lacawac.

293

### 294 ***1.3 Explanation of Calculations and Statistical Analysis***

295 To determine the fate of terrestrial DOC in the four lakes, we used the measured changes  
296 (i.e. final – control) in DOC and DIC concentrations to identify four pools of DOC:  
297 photomineralized, partially photodegraded, biodegraded, and unprocessed. Each pool was  
298 converted to a carbon basis, and we assumed a conversion of 0.5 moles CO<sub>2</sub> for each mole of  
299 DOC consumed (Cory et al., 2014). The amount of carbon photomineralized (converted to CO<sub>2</sub>)  
300 was calculated as the concentration of DIC produced [DIC\*2] by sunlight (i.e. carbon that was  
301 completely oxidized by sunlight). The amount of carbon partially photodegraded represents the  
302 remainder of the carbon pool that was processed by sunlight (but not completely oxidized to  
303 CO<sub>2</sub>) and was calculated as the total DOC processed by sunlight minus the amount  
304 photomineralized [Total Photodegraded – Photomineralized]. The amount of carbon biodegraded  
305 was calculated as the concentration of DOC lost in the biodegradation treatments. The  
306 unprocessed carbon was calculated as the fraction of the carbon pool that was not processed by  
307 either sunlight or microbes [Control DOC – Photomineralized – Partially Photodegraded –



308 Biodegraded]. Each process was determined for each lake and each month. Here we report the  
309 average response across all four months for each DOC pool.

310 While we carried out monthly experiments (May-August), here we report the average  
311 response across the open-water season (i.e. all four months) to provide a more complete picture  
312 of DOC processing. The downside of this approach is that it potentially increases variation in  
313 variables associated with DOC processing, since such processing may vary across the season.  
314 However, there was not a strong seasonal response to photodegradation or biodegradation in all  
315 of our study variables (SI Fig. 3). Furthermore, the majority of the terrestrial DOC was collected  
316 on a single date and time (except for Lake Annie).

317 Final treatments were compared relative to the dark and killed (1% HgCl<sub>2</sub>) control  
318 treatments, as those samples were deployed at the surface of the lake with the photodegradation  
319 and biodegradation treatments. We used a t-test to determine whether the photodegradation  
320 samples for all of the variables were significantly different from the biodegradation samples (n =  
321 12 for each treatment) in each lake. Photodegradation and biodegradation samples were analyzed  
322 separately using a one-way ANOVA to assess differences between lakes. A post-hoc Tukey's  
323 multiple comparison test (Sigma Plot 14.0) was used to determine if there were significant  
324 differences in the response variables between the lakes to the photodegradation and  
325 biodegradation treatments. A descriptive discriminant analysis (DDA) was used to classify the  
326 four lakes based on changes in DOC, DIC, DO, SUVA<sub>320</sub>, and S<sub>r</sub> measurements due to  
327 photodegradation. Since these five measures are likely to be highly correlated with one another,  
328 DDA is a good choice since it considers these relationships simultaneously in the analysis  
329 (Sherry 2006). In this case, DDA, works by producing linear combinations of the five measured  
330 variables (DOC, DIC, DO, SUVA<sub>320</sub>, and S<sub>r</sub>). The first linear combination provides the best



331 separation of the four lakes, followed by subsequent linear combinations for axes that are  
332 orthogonal (Sherry, 2006). Linear combinations are weighted more heavily by variables that are  
333 better able to discriminate between the lakes. In the figures and tables below, we report these  
334 data as either average measured changes (i.e. concentrations) or average percent changes and  
335 have indicated where appropriate. Data for this experiment were analyzed in either Sigma Plot  
336 14.0 (Fig. 1, Table 2) or Systat version 10.2 (Fig. 4).

337

## 338 **2. Results**

339 Throughout the results and discussion, the use of the lake names is to present the data in a  
340 meaningful manner, but it is important to recognize that the actual water samples originated from  
341 groundwater samples adjacent to each lake.

342

### 343 ***2.1 Photodegradation and biodegradation responses in each lake***

344 Photodegradation altered DOC quantity and CDOM significantly more than  
345 biodegradation for terrestrial DOC from the watersheds of all four lakes (Table 2, Fig. 1). For the  
346 photodegradation only treatments, exposure to sunlight resulted in significant production of DIC  
347 and increases in  $S_r$ , as well as significant decreases in DO, DOC, and  $SUVA_{320}$  relative to the  
348 biodegradation treatments. The only significant effect of biodegradation on terrestrial DOC was  
349 a reduction in DO concentrations compared to the dark control (Fig. 1c). In all other cases, the  
350 biodegradation treatments were not significantly different than the control, and the average  
351 percent change was close to 0.

352 The terrestrial DOC from the brown-water lakes (Lacawac and Annie) typically followed  
353 similar patterns to each other, while the terrestrial DOC from the oligotrophic and eutrophic



354 lakes (Giles and Waynewood) responded more similarly to each other. In the brown-water lakes,  
355 we observed a stronger response in DOC quantity (i.e. DOC, DIC, and DO), while the changes in  
356 DOC quantity were much more muted in the oligotrophic and eutrophic lakes. The responses of  
357  $S_r$  changes in each lake due to sunlight did not differ significantly. All four lakes showed a strong  
358 response to changes in terrestrial CDOM (i.e.  $SUVA_{320}$  and  $S_r$ ).  
359



360 **Table 2.** A summary of the mean ( $\pm$  SD) final concentration of DOC, DIC, DO,  $SUVA_{320}$  and  $S_r$   
 361 in photodegradation (Photo), biodegradation (Bio), and control experimental treatments in  
 362 groundwater samples from the watersheds of lakes Lacawac, Annie, Giles, and Waynewood. The  
 363 mean ( $\pm$  SD) initial concentration for each variable is also depicted. The P/B column list the  
 364 results of a t-test to determine whether photodegradation samples were significantly different  
 365 from the biodegradation samples ( $n = 12$  for each treatment for the four months). Bolded values  
 366 indicate the Photo treatments that were statistically different from the Bio treatments ( $p < 0.05$ ).

Analysis	Treatment	Lacawac (Mean $\pm$ SD)	P/B p-value	Annie (Mean $\pm$ SD)	P/B p-value	Giles (Mean $\pm$ SD)	P/B p-value	Waynewood (Mean $\pm$ SD)	P/B p-value
DOC ( $\mu\text{moles L}^{-1}$ )	Photo	3600 $\pm$ 330	<b>p &lt; 0.001</b>	1270 $\pm$ 211	<b>p &lt; 0.001</b>	692 $\pm$ 123	<b>p &lt; 0.001</b>	883 $\pm$ 73.3	<b>p &lt; 0.001</b>
	Bio	4910 $\pm$ 674		1810 $\pm$ 45.7		608 $\pm$ 99.0		765 $\pm$ 93.8	
	Control	5110 $\pm$ 628		1820 $\pm$ 76.9		630 $\pm$ 102		783 $\pm$ 73.8	
DIC ( $\mu\text{moles L}^{-1}$ )	Photo	54 $\pm$ 8.2	<b>p &lt; 0.001</b>	41.9 $\pm$ 11.4	<b>p &lt; 0.001</b>	20.4 $\pm$ 1.9	<b>p &lt; 0.001</b>	32.2 $\pm$ 7.3	<b>p = 0.02</b>
	Bio	16.1 $\pm$ 5.0		25.3 $\pm$ 7.2		17.7 $\pm$ 3.0		27.1 $\pm$ 8.0	
	Control	13.8 $\pm$ 4.6		30.4 $\pm$ 18.2		15.3 $\pm$ 2.1		27.8 $\pm$ 3.5	
DO ( $\mu\text{moles L}^{-1}$ )	Photo	278 $\pm$ 62.4	<b>p &lt; 0.001</b>	419 $\pm$ 25.9	<b>p &lt; 0.001</b>	536 $\pm$ 35.6	p = 0.09	522 $\pm$ 49.0	<b>p &lt; 0.001</b>
	Bio	556 $\pm$ 46.4		533 $\pm$ 42.2		556 $\pm$ 34.3		577 $\pm$ 76.9	
	Control	660 $\pm$ 29.4		656 $\pm$ 32.1		688 $\pm$ 60.9		702 $\pm$ 57.3	
$SUVA_{320}$ ( $\text{m}^{-1}/\text{mg L}^{-1}$ )	Photo	4.3 $\pm$ 0.4	<b>p &lt; 0.001</b>	2.4 $\pm$ 0.4	<b>p &lt; 0.001</b>	2.4 $\pm$ 0.2	<b>p &lt; 0.001</b>	1.8 $\pm$ 0.2	<b>p &lt; 0.001</b>
	Bio	5.3 $\pm$ 0.2		3.8 $\pm$ 0.1		4.8 $\pm$ 0.3		3.2 $\pm$ 0.2	
	Control	5.1 $\pm$ 0.2		3.8 $\pm$ 0.1		4.7 $\pm$ 0.2		3.2 $\pm$ 0.1	
$S_r$	Photo	1.1 $\pm$ 0.0	<b>p &lt; 0.001</b>	1.3 $\pm$ 0.1	<b>p &lt; 0.001</b>	1.4 $\pm$ 0.1	<b>p &lt; 0.001</b>	1.2 $\pm$ 0.1	<b>p &lt; 0.001</b>
	Bio	0.7 $\pm$ 0.1		0.8 $\pm$ 0.0		0.9 $\pm$ 0.1		0.8 $\pm$ 0.1	
	Control	0.7 $\pm$ 0.1		0.8 $\pm$ 0.0		0.9 $\pm$ 0.1		0.9 $\pm$ 0.1	

367

368 Sunlight caused average ( $\pm$  SD) DOC losses relative to the control treatments of  $30.5 \pm$   
 369  $11.5\%$  and  $28.9 \pm 8.3\%$  in Lacawac and Annie, respectively (Fig. 1a). In Giles and Waynewood,  
 370 we observed an average of  $9.6 \pm 6.5\%$  and  $13.4 \pm 6.2\%$  increase in DOC concentration,  
 371 respectively following exposure to sunlight. When we compared lakes within each treatment,  
 372 there were no significant differences in DOC concentration due to sunlight in Giles vs.  
 373 Waynewood, whereas Annie and Lacawac were significantly different from the prior two lakes  
 374 and from each other (ANOVA:  $F_{1,3} = 70.9$ ,  $p < 0.001$ ).

375 Decreases in DOC concentration due to photodegradation could lead to mineralization  
 376 (i.e. DIC production; Fig. 1b) and therefore oxidation (i.e. DO consumption; Fig. 1c). We



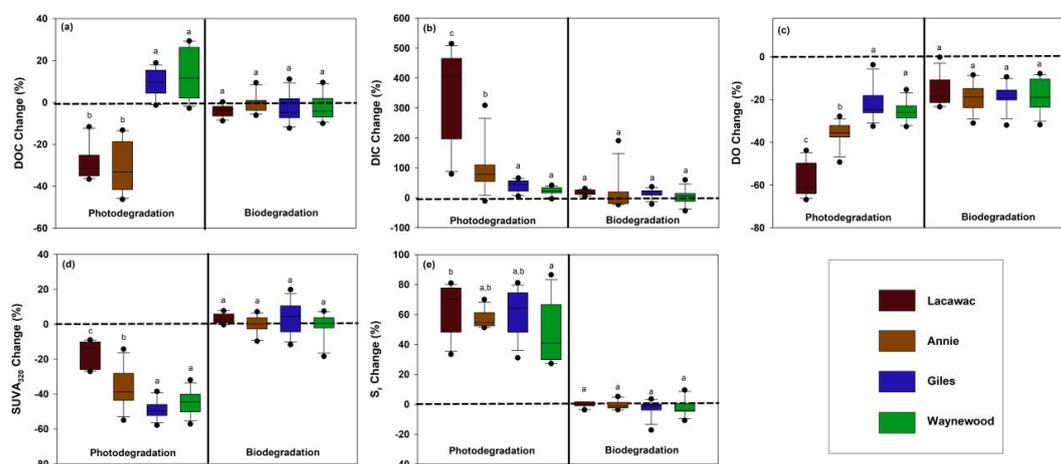
377 observed the production of DIC due to sunlight in all of our lakes (Fig. 1b). In Lacawac and  
378 Annie, the average ( $\pm$  SD) percent increases in DIC relative to the control treatments were  $350 \pm$   
379  $160\%$  and  $96.0 \pm 79.0\%$ , respectively. The average percent increases relative to controls in Giles  
380 and Waynewood were  $40.7 \pm 19.4\%$  and  $23.2 \pm 12.7\%$  respectively. The DIC percent change  
381 was similar between Giles and Waynewood, and both were statistically different from Annie and  
382 Lacawac. The percent DIC change in Lacawac was significantly higher than Annie (ANOVA:  
383  $F_{1,3} = 36.4$ ,  $p < 0.001$ ).

384 In all lakes, both photodegradation and biodegradation led to decreases in DO  
385 concentrations (Fig. 1c). Average DO losses due to biodegradation for all four lakes ranged from  
386 15 to 18%. DO losses due to photodegradation were more variable. The average DO loss from  
387 sunlight in Lacawac and Annie was  $58.2 \pm 7.8\%$  and  $35.9 \pm 5.4\%$ , respectively. In Giles and  
388 Waynewood, we observed average DO losses of  $21.6 \pm 7.9\%$  and  $25.6 \pm 4.7\%$  respectively.  
389 While the largest losses of DO due to sunlight were observed in Annie and Lacawac, there was  
390 no significant difference between Annie and Waynewood. Giles and Lacawac were significantly  
391 different from the other two lakes and from each other (ANOVA:  $F_{1,3} = 73.9$ ,  $p < 0.001$ ).

392 Changes in CDOM due to biodegradation were minimal in all of the lakes (Fig. 1d & 1e).  
393 In contrast, photodegradation caused significant changes in all of the lakes, but the magnitude of  
394 the change varied by lake.  $SUVA_{320}$  decreased in all lakes due to sunlight, but the largest changes  
395 were observed in the oligotrophic and eutrophic lakes (Fig. 1d). Average  $SUVA_{320}$  values  
396 decreased between 16.8% in Lacawac and 48.9% in Giles. The response in Annie and  
397 Waynewood were similar, whereas Lacawac and Giles were significantly different from the prior  
398 two lakes and each other (ANOVA:  $F_{1,3} = 39.7$ ,  $p < 0.001$ ). In all lakes,  $S_r$  increased due to  
399 sunlight (Fig. 1e). Average percent increases for the lakes ranged from 46.4% in Waynewood to



400 65.1% in Lacawac. For  $S_r$ , the response between Lacawac and Waynewood were significantly  
401 different, but those lakes were no different compared to the remaining lakes (ANOVA:  $F_{1,3} = 3.1$ ,  
402  $p = 0.04$ ).  
403



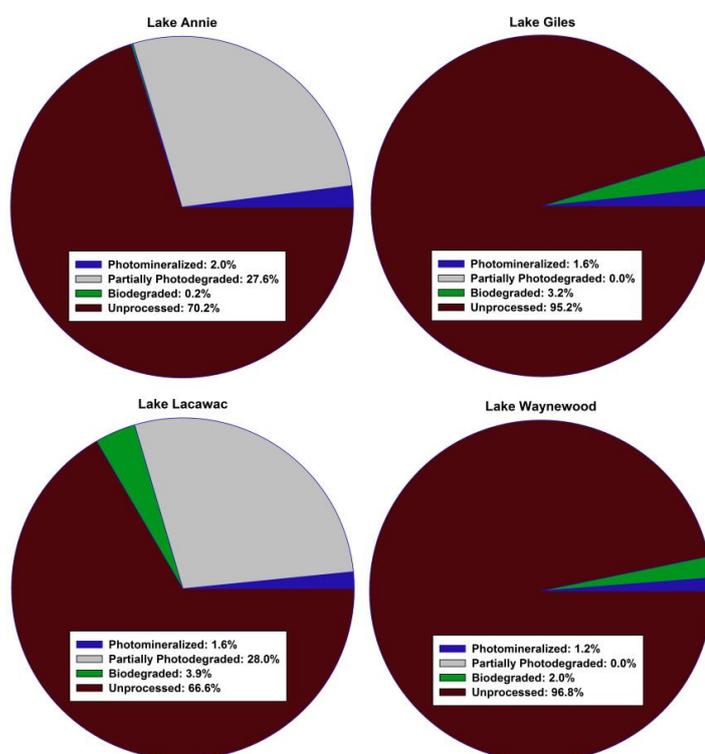
404  
405 **Figure 1.** The monthly average percent change from the dark and killed control treatments  
406 (dashed line) in each lake for photodegradation (left) and biodegradation (right) for (a) DOC, (b)  
407 DIC, (c) DO, (d)  $SUVA_{320}$ , and (e)  $S_r$ . Statistical differences ( $p < 0.05$ ) between lakes are  
408 indicated by different letters above each boxplot. For each boxplot  $n = 12$  replicates.  
409

## 410 2.2 Fate of DOC

411 Of the four pools of carbon we identified in the groundwater samples entering our study  
412 lakes, we found the average amount of carbon processed by sunlight ranged from 1.2% to ~30%  
413 (Fig. 2). Carbon in Giles and Waynewood ( $< 2\%$ ) showed little response to sunlight, whereas the  
414 response in Annie and Lacawac (~30%) was much higher over the 7-day experiments. The  
415 dominant pathway through which sunlight interacted with DOC was through partial  
416 photodegradation in these latter two lakes. About 2% of the carbon pool was photomineralized in  
417 the brown water lakes. The amount of carbon processed via biodegradation was minimal in all



418 lakes (ranging from 0.2–4%). The fraction of the unprocessed carbon pool ranged from a low of  
419 66% for Lacawac to a high of 97% for Waynewood. An average of 3.4 to 34% of the carbon  
420 pool was processed in one week.



421

422 **Figure 2.** A summary of the average fate of carbon in the groundwater samples from our study  
423 lakes (see methods section for explanation of calculations). All terms were converted to a carbon  
424 basis. Photomineralized describes the amount of carbon completely mineralized to CO<sub>2</sub> by  
425 sunlight. Partially photodegraded describes the amount of carbon processed by sunlight minus  
426 the amount photomineralized. Biodegraded describes the amount of carbon lost through  
427 biodegradation. Unprocessed carbon describes the remaining carbon that was not processed by  
428 photodegradation or biodegradation.

429

### 430 **2.3 DOC response by lake trophic status**

431 For the descriptive discriminant analysis (DDA) to classify the lakes, we found that the  
432 five metrics were strongly correlated with one another (Table 3). In general, the changes in DOC,



433 DIC, and DO were more strongly correlated with one another than with SUVA<sub>320</sub> and S<sub>r</sub> and vice  
434 versa (Table 3). We will refer to the changes in DOC, DIC, and DO as “DOC quantity” and the  
435 changes in SUVA<sub>320</sub> and S<sub>r</sub> as “CDOM” for brevity.

436

437 **Table 3.** Pearson correlations between the measured changes in the five metrics: DOC, DIC, DO,  
438 SUVA<sub>320</sub>, and S<sub>r</sub>.

	DOC	DIC	DO	SUVA <sub>320</sub>
DIC	-0.934			
DO	0.869	-0.837		
SUVA <sub>320</sub>	-0.705	-0.671	-0.666	
S <sub>r</sub>	-0.027	0.021	0.163	-0.319

439

440 DDA produced three functions (axes) with canonical correlations of 0.961, 0.753, and  
441 0.181 (Fig. 3). Collectively, the entire model was significant (Wilks'  $\lambda = 0.032$ ;  $F_{15, 108} = 17.79$ ;  $p$   
442  $< 0.001$ ). Effect size was calculated following Sherry and Henson (2010) as  $1 - \text{Wilks' } \lambda$ , and  
443 therefore the overall model explains 96.8% of the variation among lakes. Functions 1 through 3  
444 and 2 through 3 were significant ( $p < 0.001$  for both). Function 3 was not significant ( $p = 0.710$ )  
445 and therefore is not discussed further. Functions 1 through 3 collectively explain 92.4% of the  
446 shared variance while functions 2 through 3 collectively explain 56.7% of the shared variance.

447 Function 1 represents a new variate that is a linear combination of the changes in the five  
448 variables that best discriminates the lakes from one another. This new variate is composed  
449 mainly of DOC, with a function coefficient of 0.465 and a structure coefficient of 0.821 (Table  
450 4). Of note are also DIC, DO, and SUVA<sub>320</sub> that had smaller function coefficients ( $< 0.45$ ), but  
451 had large structure coefficients ( $> 0.45$ ). This result suggests that Function 1 is mainly related to  
452 DOC quantity. Function 2, also a new variate that is a linear combination of the five measured  
453 changes, is composed mainly of SUVA<sub>320</sub> (function coefficient = 0.985 and structure coefficient

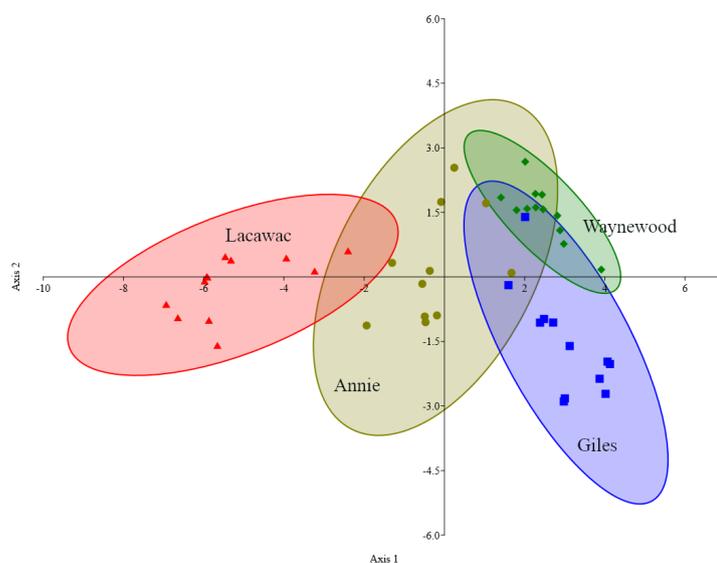


454 = 0.719; Table 4). Function 2 is orthogonal to Function 1 and together they discriminate the four  
 455 lakes (Fig. 3).

456  
 457 **Table 4.** The solution for changes in measured independent variables that predict the dependent  
 458 variable, lake. Structure coefficients ( $r_s$ ) and communality coefficients greater than  $|0.45|$  are in  
 459 bold. Coeff = standardized canonical function coefficient;  $r_s$  = structure coefficient;  $r_s^2$  = squared  
 460 structure coefficient.  
 461

Variable	Function 1			Function 2		
	Coeff.	$r_s$	$r_s^2$ (%)	Coeff.	$r_s$	$r_s^2$ (%)
DOC	0.465	<b>0.821</b>	67.40	0.639	0.278	40.83
DIC	-0.337	<b>-0.703</b>	11.36	-0.059	-0.216	0.35
DO	0.440	<b>0.679</b>	19.36	-0.124	0.009	1.54
SUVA <sub>320</sub>	-0.139	<b>-0.473</b>	1.93	0.985	<b>0.719</b>	97.02
S <sub>r</sub>	0.244	0.068	5.95	-0.238	-0.434	5.66

462



463

464 **Figure 3.** Canonical plot scores and 95% confidence ellipses from descriptive discriminant  
 465 analysis of the measured changes (i.e. treatment minus control) in the five variables (DOC, DIC,  
 466 DO, SUVA<sub>320</sub>, and S<sub>r</sub>) and four lakes: Annie (olive circles), Giles (blue squares), Lacawac (red  
 467 triangles), and WayneWood (green diamonds). Only photodegradation samples were included in  
 468 this analysis.



469 DDA correctly classified 89.4% of the samples to their collection site (Fig. 3). One  
470 sample from Annie was incorrectly assigned to Waynewood, two samples from Giles were  
471 incorrectly assigned to Waynewood, and two samples from Lacawac were incorrectly assigned  
472 to Annie. All of the Waynewood samples were correctly classified.

473

### 474 **3. Discussion**

#### 475 **3.1 Comparing the relative importance of photodegradation and biodegradation**

476 Despite a large number of studies examining the effects of either photodegradation or  
477 biodegradation on DOC processing, very few have conducted simultaneous *in-situ* experiments  
478 of the relative importance of both processes for transforming DOC from the watersheds of a  
479 range of different lakes. Our results indicate that sunlight was the primary process in the surface  
480 waters responsible for degrading terrestrial DOC from the watershed of all four lakes.  
481 Biodegradation played a minimal role in changing the DOC quantity and CDOM. We observed  
482 decreases in DOC, DO, and SUVA<sub>320</sub> due to sunlight and saw increases in DIC and S<sub>r</sub>. The loss  
483 of DOC, as well as a shift to more photobleached, and lower molecular weight organic material  
484 is consistent with prior studies on these lakes that evaluated just the effects of sunlight (Morris  
485 and Hargreaves, 1997). Exceptions to DOC loss due to photodegradation occurred in Giles and  
486 Waynewood. In these lakes, we observed an increase in average DOC concentrations. In Giles,  
487 there was significant production of DOC in June and July. In Waynewood, significant production  
488 occurred in May and July. We speculate that this production may be due to the lysing of any  
489 microbes remaining in solution. Increases may also be attributed to interactions with iron. We  
490 have no measurable evidence, but a number of samples from Giles and Waynewood contained a



491 red precipitate at the conclusion of the one-week experiments. Previously iron-bound DOC could  
492 have been released back into the water.

493 Dissolved oxygen was the lone variable where biodegradation led to decreases in DO  
494 relative to the controls, but the differences between lakes were not significant. We attributed the  
495 changes in DO to the “sloppy feeding” of bacteria, where they produce DOC through exudates  
496 and then assimilate it (Evans et al., 2017). The above results are similar to observations in Arctic  
497 and tropical waters in that photodegradation was more important than biodegradation on short  
498 time scales (Cory et al., 2014; Chen and Jaffé, 2014; Amado et al., 2003). Interestingly, we  
499 found that terrestrial DOC from the watersheds of lakes of different trophic status was processed  
500 differently, resulting in DIC production and DOC degradation for the brown-water lakes  
501 (Lacawac and Annie), but greater changes in  $SUVA_{320}$  for the oligotrophic and eutrophic lakes  
502 (Giles and Waynewood). This highlights the need to account for lake trophic status in predicting  
503 DOC processing and  $CO_2$  emissions from lakes.

504

### 505 ***3.2 Dominant degradation process***

506 Based on our study design we were able to identify four pools of carbon:  
507 photomineralized, partially photodegraded, biodegraded, and unprocessed. The dominant  
508 degradation pathway across all lakes was partial photodegradation (i.e. loss of DOC, but no  
509 mineralization), although the size of each carbon pool varied by lake. In the brown-water lakes,  
510 ~28% of the total carbon pool was partially photodegraded and ~2% was photomineralized. In  
511 the oligotrophic and eutrophic lakes ~1.4% of the carbon was photodegraded and none of the  
512 carbon was photomineralized.



513 Observations in Toolik Lake showed 70% of the total carbon pool being processed by  
514 sunlight during the open water period (~3 months) (Cory et al., 2014). Other estimates have  
515 found that photomineralization of DOC accounts for only 8-14% of total water column CO<sub>2</sub>  
516 production (Granéli et al., 1996; Jonsson et al., 2001; Koehler et al., 2014; Vachon et al., 2016b).  
517 We observed 30% of the carbon pool being processed by sunlight within one week in our lakes  
518 and this was restricted to the brown-water lakes. Similar to Toolik Lake, the dominant  
519 degradation process was partial photodegradation. Partial photodegradation can alter CDOM and  
520 stimulate subsequent bacterial respiration. Degradation of CDOM can have important effects for  
521 downstream ecosystems if it can be further processed and released as CO<sub>2</sub> or instead is buried or  
522 exported downstream (Weyhenmeyer et al., 2012; Catalan et al., 2016; Chen and Jaffe, 2014;  
523 Biddanda and Cotner, 2003). It is thus important to include all sunlight-driven degradation  
524 processes to fully account for its relative importance.

525 Differences between the responses observed in the Arctic and our temperate/subtropical  
526 lakes are most likely explained by the initial concentration and quality of terrestrially derived  
527 DOC. In the Arctic, glacial meltwater can be highly photolabile and dominated by seasonal  
528 inputs of DOC from shallow or deep soils (Cory et al., 2014; Spencer et al., 2014; and Kaiser et  
529 al., 2017). In temperate regions, DOC tends to contain more humic and fulvic acids derived from  
530 soils, which may be less photolabile than Arctic DOC. Additionally, we did not integrate our  
531 results over the entire water column because the samples were analyzed on the surface of a single  
532 lake. Over the entire water column, photodegradation could have processed additional carbon. In  
533 clear-water lakes, DOC may be photodegraded down to the 1% UV-A attenuation depth (Osburn  
534 et al., 2001), which ranged from 0.7-4.7 m in our study lakes (Table 1).

535



### 536 **3.3 Response of lakes to photodegradation**

537           With an increase in extreme precipitation events, terrestrial DOC inputs are likely to  
538 increase in many aquatic ecosystems (Rahmstorf and Coumou, 2011; Westra et al., 2014). By  
539 using groundwater as a proxy of terrestrial inputs from the watersheds of different types of lakes,  
540 we simulated the effects of storm events and compared the sensitivity of different terrestrial  
541 DOC sources to photodegradation. Interestingly, we found DOC from the watersheds of  
542 oligotrophic and eutrophic lakes showed stronger changes in CDOM, compared to DOC from the  
543 watersheds of the brown-water lakes that showed significantly larger changes in DOC quantity.  
544 This difference may be due to the more allochthonous nature of the brown-water DOC, which is  
545 highly photolabile, resulting in greater changes in DOC quantity due to its ability to absorb UV  
546 radiation (Bertilsson and Tranvik, 2000). The less allochthonous and more microbially derived  
547 DOC from the watersheds of the eutrophic and oligotrophic lakes may be less photolabile with  
548 fewer UV-absorbing chromophores. Results of the DDA may be helpful in predicting changes in  
549 other lakes based on their trophic status.  $SUVA_{320}$  is the variable most likely to change due to  
550 photodegradation in eutrophic and oligotrophic lakes. In contrast, DOC concentration is the  
551 variable most likely to change in brown-water lakes due to photodegradation. Both results (DOC  
552 and  $SUVA_{320}$ ) highlight how lakes of varying trophic status respond to photodegradation. These  
553 results can be used to predict how lakes not included in this study will respond to increased DOC  
554 concentrations (i.e. browning).

555           Across our study lakes, changes in DIC production scaled linearly with initial  
556 groundwater DOC concentration. Lacawac had the highest initial DOC concentration ( $59.4 \pm$   
557  $6.1$ ) and the highest average DIC production, while Giles had the lowest initial DOC  
558 concentration ( $6.0 \pm 0.6$ ) and the lowest average DIC production. This suggests that DOC



559 concentration plays a critical role in determining the fate of DOC (Leech et al., 2014; Lapierre et  
560 al., 2013). Recent research has also reported that residence time controls organic carbon  
561 decomposition across a wide range of freshwater ecosystems (Catalan et al., 2016, Evans et al.,  
562 2017). However, extreme precipitation events may shorten the residence time of lakes,  
563 effectively flushing out fresh DOC and preventing significant in-lake degradation from occurring  
564 (de Wit et al., 2018). For the terrestrial DOC from the oligotrophic and eutrophic lakes, a  
565 significant fraction was not degraded, which may mean that terrestrial inputs from these  
566 watersheds undergoes less immediate in-lake processing and instead is exported downstream.  
567 Our results indicate that differences in the fate and processing of DOC from the watersheds of a  
568 range of lake types have important implications for determining which lakes may release more  
569 CO<sub>2</sub> versus export DOC downstream (Weyhenmeyer et al., 2012; Zwart et al., 2015;  
570 Weyhenmeyer and Conley, 2017).

571       Even though we observed similar responses to photodegradation in the brown-water lakes  
572 (Fig. 1), the magnitude of the response varied and may have been related to the initial DOC  
573 concentration. Initial concentrations (mg L<sup>-1</sup>) of terrestrial DOC from Lacawac ( $59.4 \pm 6.1$ ) were  
574 almost 3x higher than Annie ( $20.7 \pm 0.5$ ). Average DOC losses for both lakes due to  
575 photodegradation were ~35%. The main difference between Lacawac and Annie was the DIC  
576 percent change due to photodegradation (Fig. 1). Average percent increases in DIC for Lacawac  
577 were close to 400%, whereas in Annie it was ~85%. Despite the fact that both Annie and  
578 Lacawac are brown-water lakes, their different DIC production rates indicate that certain types of  
579 terrestrial DOC may be more photolabile than others and capable of outgassing large amounts of  
580 CO<sub>2</sub>. The DDA analysis did also pick out the separation between Lacawac and Annie primarily  
581 on axis 1 (DOC). The responses in Annie shared similarities with the other 3 lakes while



582 Lacawac only overlapped with Annie. When put in the context of the entire DOC pool for each  
583 lake, photomineralization accounted for 2% of the carbon loss. We anticipated that terrestrial  
584 DOC from subtropical lakes would undergo additional microbial processing due to the higher  
585 temperatures year-round. In a comparison between boreal Swedish and tropical Brazilian lakes,  
586 Graneli et al., (1998) also found strong similarities in changes of DOC concentrations and DIC  
587 production between lakes from the different latitudes. A weak significant correlation between  
588 DOC concentration and DIC production has also been observed in Amazon clear water systems  
589 (Amado et al., 2003)

590

## 591 **Conclusions**

592 Here we showed that photodegradation can be more important than biodegradation in  
593 processing watershed inputs of terrestrial DOC on short time scales in the surface waters of a  
594 lake. The responses that we observed varied with lake trophic status. Quantitative changes in  
595 DOC, DIC, and DO were strongest in the terrestrial DOC from the watersheds of the brown-  
596 water lakes, whereas the largest changes in  $SUVA_{320}$  were observed in the terrestrial DOC from  
597 the watersheds of the eutrophic and oligotrophic lakes. Consistent with prior studies, we found  
598 that sunlight can impact not only changes in the concentration, but also the absorbance properties  
599 of the DOC pool. We observed a range of 1.2 to 34% of the carbon pool processed in one week.  
600 As DOC concentrations increase in some aquatic ecosystems, the potential for increased  $CO_2$   
601 outgassing due to photo-mineralization also increases. On short time scales, sunlight had  
602 important impacts on our study lakes. Future studies should focus on additional lakes, longer  
603 timescales, and integrating DIC production throughout the water column.



604 Over the next century, DOC concentrations in northern boreal lakes are projected to  
605 increase by 65% (Larsen et al., 2011). Thus, understanding the fate of terrestrial sourced organic  
606 material will be essential for predicting the ecological consequences for lakes and downstream  
607 ecosystems (Solomon et al., 2015; Williamson et al., 2015; Finstad et al., 2016). Improving  
608 estimates of organic carbon processing in lakes will be an important component of creating more  
609 complete carbon budgets (Hanson et al., 2004; 2014) and global estimates of CO<sub>2</sub> emissions can  
610 be more accurately scaled to reflect the ability of lakes to act as CO<sub>2</sub> sinks or sources as  
611 browning continues (Lapierre et al., 2013, Evans et al., 2017).

612

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621 declare no competing interests.

622

### 623 **Data Availability:**

624 Data and metadata will be made available in the Environmental Data Initiative repository. Data  
625 archiving will be led by C. Dempsey and J. Brenttrup.

626



627 **Author Contribution Statement**

628 CMD, JAB, and CEW designed the study with help from LBK, EEG, and HMS. CMD, JAB,  
629 SM, and HMS collected the water samples and ran the experiments. DPM provided the  
630 analytical equipment for measuring DIC and DOC. CMD and JAB analyzed the data, and CMD  
631 and MTG conducted the statistical and DDA analyses. CMD and JAB wrote the manuscript with  
632 contributions from all of the authors.

633



## References

- 634  
635  
636 Amado, A.M., Farjalla, V.F., Esteves, F.A., and Bozelli, R.L. DOC photo-oxidation in clear  
637 water Amazonian aquatic ecosystems. *Amazonia* 17 (3/4): 513-523, 2003.
- 638 Bertilsson, S. and L. J. Tranvik. Photochemical transformation of dissolved organic matter in  
639 lakes. *Limnology and Oceanography* 45: 753–762. doi:10.4319/lo.2000.45.4.0753, 2000.
- 640 Biddanda, B.A. and Cotner, J.B. Enhancement of Dissolved Organic Matter Bioavailability by  
641 Sunlight and Its Role in the Carbon Cycle of Lakes Superior and Michigan. *Journal of Great  
642 Lakes Research* 29 (2): 228-241, 2003.
- 643 Catalan, N., R. Marcé, D. N. Kothawala, and L. J. Tranvik. Organic carbon decomposition rates  
644 controlled by water retention time across inland waters. *Nature Geoscience* 9: 501–504.  
645 doi:10.1038/ngeo2720, 2006.
- 646 Cole, J. J., N. F. Caraco, G. W. Kling, and T. K. Kratz. Carbon dioxide supersaturation in the  
647 surface waters of lakes. *Science* 265: 1568–1570, 1994.
- 648 Cory, R. M., C. P. Ward, B. C. Crump, and G. W. Kling. Sunlight controls water column  
649 processing of carbon in arctic fresh waters. *Science* 345: 925–928.  
650 doi:10.1126/science.1253119, 2014.
- 651 Chen, M. and R. Jaffe. Photo- and bio-reactivity patterns of dissolved organic matter from  
652 biomass and soil leachates and surface waters in a subtropical wetland. *Water Research* 61:  
653 181-190. DOI: 10.1016/j.watres.2014.03.075, 2014.
- 654 Chen, M., Jaffé, R. Quantitative assessment of photo- and bio-reactivity of chromophoric and  
655 fluorescent dissolved organic matter from biomass and soil leachates and from surface  
656 waters in a subtropical wetland. *Biogeochemistry* 129: 273–289.  
657 <https://doi.org/10.1007/s10533-016-0231-7>, 2016.
- 658 de Eyto, E., E. Jennings, E. Ryder, K. Sparber, M. Dillane, C. Dalton, and R. Poole. Response of  
659 a humic lake ecosystem to an extreme precipitation event: physical, chemical, and biological  
660 implications. *Inland Waters* 6: 483–498. doi:10.5268/IW-6.4.875, 2016.
- 661 de Wit, H. A., S. Valinia, G. A. Weyhenmeyer, and others. Current Browning of Surface Waters  
662 Will Be Further Promoted by Wetter Climate. *Environ. Sci. Technol. Lett.* 3: 430–435.  
663 doi:10.1021/acs.estlett.6b00396, 2016.
- 664 de Wit, H.A., Couture, R.M., Jackson-Blake, L., Futter, M.N., Valinia, S., Austnes, K., Guerrero,  
665 J., and Lin, Y. Pipes or chimneys? For carbon cycling in small boreal lakes, precipitation  
666 matters most. *Limnology and Oceanography Letters* 3:275-284, 2018.
- 667 del Giorgio, P. A., J. J. Cole, and A. Cimleris. Respiration rates in bacteria exceed  
668 phytoplankton production in unproductive aquatic systems. *Nature* 385: 148–151, 1997.
- 669 Duan, S., He, Y., Kaushai, S.S., Bianchi, T.S., Ward, N.D., and Guo, L. Impact of wetland  
670 decline on decreasing dissolved organic carbon concentrations along the Mississippi River  
671 continuum. *Frontiers in Marine Science* 3:280. doi: 10.3389/fmars.2016.00280, 2017.
- 672 Evans, C.D., Futter, M.N., Moldan, F., Valinia, S., Frogbrook, Z., and Kothawala, D.N.  
673 Variability in organic carbon reactivity across lake residence time and trophic gradients.  
674 *Nature Geosciences* 3: 832-837. doi: 10.1038/NGEO3051, 2017.
- 675 Fasching, C., and T. J. Battin. Exposure of dissolved organic matter to UV-radiation increases  
676 bacterial growth efficiency in a clear-water Alpine stream and its adjacent groundwater.  
677 *Aquatic Sciences* 74: 143–153. doi:10.1007/s00027-011-0205-8, 2011.
- 678 Finstad, A. G., T. Andersen, S. Larsen, K. Tominaga, S. Blumentrath, H. A. de Wit, H.  
679 Tømmervik, and D. O. Hessen. From greening to browning: Catchment vegetation



- 680 development and reduced S-deposition promote organic carbon load on decadal time scales  
681 in Nordic lakes. *Sci. Rep.* 6: 31944–9. doi:10.1038/srep31944, 2016.
- 682 Fischer, E. M., and R. Knutti. Anthropogenic contribution to global occurrence of heavy  
683 precipitation and high-temperature extremes. *Nature Climate Change* 5: 560–564.  
684 doi:10.1038/nclimate2617, 2015.
- 685 Gaiser, E.E., Deyrup, N.D., Bachmann, R.W., Battoe, L.E. and Swain, H.M. Effects of climate  
686 variability on transparency and thermal structure in subtropical, monomictic Lake Annie,  
687 Florida. *Fundamental and Applied Limnology*. Vol 175 (3): 217-230, 2009.
- 688 Granéli, W., M. Lindell, and L. Tranvik. Photo-oxidative production of dissolved inorganic  
689 carbon in lakes of different humic content. *Limnology and Oceanography* 41: 698–706.  
690 doi:10.4319/lo.1996.41.4.0698, 1996.
- 691 Granéli, W., Lindell, M., de Faria, B.M., and de Assis Esteves, F. Photoproduction of dissolved  
692 inorganic carbon in temperate and tropical lakes – dependence on wavelength band and  
693 dissolved organic carbon concentration. *Biogeochemistry* 43: 175-195, 1998.
- 694 Hanson, P. C., A. I. Pollard, D. L. Bade, K. Predick, S. R. Carpenter, and J. A. Foley. A model of  
695 carbon evasion and sedimentation in temperate lakes. *Global Change Biology* 10: 1285–  
696 1298. doi:10.1111/j.1529-8817.2003.00805.x, 2004.
- 697 Hanson, P. C., M. L. Pace, S. R. Carpenter, J. J. Cole, and E. H. Stanley. Integrating Landscape  
698 Carbon Cycling: Research Needs for Resolving Organic Carbon Budgets of Lakes.  
699 *Ecosystems* 18: 363–375. doi:10.1007/s10021-014-9826-9, 2014.
- 700 Helms, J. R., Stubbins, A., Ritchie, J. D., Minor, E. C., Kieber, D. J., and Mopper, K. Absorption  
701 spectral slopes and slope ratios as indicators of molecular weight, source, and  
702 photobleaching of chromophoric dissolved organic matter, *Limnol. Oceanogr.*, 53, 955–969,  
703 2008.
- 704 Hosen, J.D., O.T. McDonough, C.M. Febria, and M.A. Palmer. Altered stream dissolved organic  
705 matter composition and bioavailability with urbanization. *Environmental Science &*  
706 *Technology*. 48:7817-7824, 2014.
- 707 Jennings, E., S. Jones, L. Arvola, and others. Effects of weather-related episodic events in lakes:  
708 an analysis based on high-frequency data. *Freshwater Biology* 57: 589–601.  
709 doi:10.1111/j.1365-2427.2011.02729.x, 2012.
- 710 Jonsson, A., M. Meili, A.-K. Bergström, and M. Jansson. Whole-lake mineralization of  
711 allochthonous and autochthonous organic carbon in a large humic lake (Oerträesket, N.  
712 Sweden). *Limnology and Oceanography* 46: 1691–1700, 2001.
- 713 Julian II, P., Gerber, S., Wright, A.L., Gu, B., and Osborne, T.Z. Carbon pool trends and  
714 dynamics within a subtropical peatland during long-term restoration. *Ecological Processes*  
715 6:43. DOI 10.1186/s13717-017-0110-8, 2017.
- 716 Kaiser, K., Candedo-Oropeza, M., McMahon, R., and Amon, R.M. Origins and transformations  
717 of dissolved organic matter in large Arctic rivers. *Scientific Reports* 7, 13064, 2017.
- 718 Kling, G. W., G. W. Kipphut, and M. C. Miller. Arctic lakes and rivers as gas conduits to the  
719 atmosphere: implications for tundra carbon budgets. *Science* 251:298-301, 1991.
- 720 Klug, J. L., D. C. Richardson, H. A. Ewing, and others. Ecosystem Effects of a Tropical Cyclone  
721 on a Network of Lakes in Northeastern North America. *Environmental Science &*  
722 *Technology* 46: 11693–11701. doi:10.1021/es302063v, 2012.
- 723 Koehler, B., T. Landelius, G. A. Weyhenmeyer, N. Machida, and L. J. Tranvik. Sunlight-induced  
724 carbon dioxide emissions from inland waters. *Global Biogeochemical Cycles* 28: 696–711.  
725 doi:10.1002/2014GB004850, 2014.



- 726 Lapiere, J.-F., F. Guillemette, M. Berggren, and P. A. del Giorgio. Increases in terrestrially  
727 derived carbon stimulate organic carbon processing and CO<sub>2</sub> emissions in boreal aquatic  
728 ecosystems. *Nat Comms* 4. doi:10.1038/ncomms3972, 2013.
- 729 Larsen, S., T. Andersen, and D. O. Hessen. Climate change predicted to cause severe increase of  
730 organic carbon in lakes. *Global Change Biology* 17: 1186–1192. doi:10.1111/j.1365-  
731 2486.2010.02257.x, 2011.
- 732 Larson, J., Frost, P., Xenopoulos, M., Williams, C., Morales-Williams, A., Vallazza, J., Nelson,  
733 J., and Richardson, W. Relationships Between Land Cover and Dissolved Organic Matter  
734 Change Along the River to Lake Transition. *Ecosystems*. 17. 10.1007/s10021-014-9804-2,  
735 2014.
- 736 Leech, D., M. Snyder, and R. Wetzel. Alterations in the photomineralization of allochthonous  
737 DOM related to elevated atmospheric CO<sub>2</sub>. *Inland Waters* 4: 147–156. doi:10.5268/IW-  
738 4.2.626, 2014.
- 739 Lu, Y. H., J. E. Bauer, E. A. Canuel, Y. Yamashita, R. M. Chambers, and R. Jaffe´.  
740 Photochemical and microbial alteration of dissolved organic matter in temperate headwater  
741 streams associated with different land use, *J. Geophys. Res. Biogeosci.*, 118, 566–580,  
742 doi:10.1002/jgrg.20048, 2013.
- 743 Madronich, S. UV radiation in the natural and perturbed atmosphere. Pages 17–69 in M. Tevini,  
744 editor. *Environmental effects of UV (ultraviolet) radiation*. Lewis Publisher, Boca Raton,  
745 Florida, 1993.
- 746 Moeller, R.E., Williamson, C.E., Hargreaves, B.R., and Morris, D.P. *Limnology of lakes*  
747 *Lacawac, Giles, and Waynewood 1989-1993: An introduction to the core lakes of the*  
748 *Pocono comparative lakes program*. Lehigh University, 1995.
- 749 Monteith, D. T., J. L. Stoddard, C. D. Evans, and others. Dissolved organic carbon trends  
750 resulting from changes in atmospheric deposition chemistry. *Nature* 450: 537–540.  
751 doi:10.1038/nature06316, 2007.
- 752 Moran, M. A., W. M. Sheldon Jr, and R. G. Zepp. Carbon loss and optical property changes  
753 during long-term photochemical and biological degradation of estuarine dissolved organic  
754 matter. *Limnology and Oceanography* 45: 1254–1264. doi:10.4319/lo.2000.45.6.1254, 2000.
- 755 Morris, D.P. and Hargreaves, B.R. The role of photochemical degradation of dissolved organic  
756 carbon in regulating the UV transparency of three lakes on the Pocono Plateau. *Limnology*  
757 *and Oceanography* 42(2): 239-249, 1997.
- 758 Osburn, C. L., H. E. Zagarese, D. P. Morris, B. R. Hargreaves, and W. E. Cravero. Calculation of  
759 spectral weighting functions for the solar photobleaching of chromophoric dissolved organic  
760 matter in temperate lakes. *Limnology and Oceanography* 46: 1455–1467.  
761 doi:10.4319/lo.2001.46.6.1455, 2001.
- 762 Parsons, T. R., Y. Maita, and C. M. Lalli. *A manual for chemical and biological methods for*  
763 *seawater analysis*, Pergamon Press, 1984.
- 764 Rahmstorf, S., and D. Coumou. Increase of extreme events in a warming world. *Proceedings of*  
765 *the National Academy of Sciences of the United States of America* 108: 17905–17909.  
766 doi:10.1073/pnas.1101766108, 2011.
- 767 Raymond, P. A., D. L. Hartmann, R. Lauerwald, and others. Global carbon dioxide emissions  
768 from inland waters. *Nature* 503: 355–359. doi:10.1038/nature12760, 2013.
- 769 Reche, I., Pace, M.L, and Cole, J.J. Relationship of trophic and chemical conditions to  
770 photobleaching of dissolved organic matter in lake ecosystems. *Biogeochemistry* 44: 259-  
771 280, 1999.



- 772 Rose, K.C., Williamson, C.E., Saros, J.E., Sommaruga, R., and Fisher, J.M. Differences in UV  
773 transparency and thermal structure between alpine and subalpine lakes: implications for  
774 organisms. *Photochemical and Photobiological Sciences*. Volume 8(9): 1244-56.  
775 doi:10.1039/b9056, 2009a.
- 776 Rose, K.C., Williamson, C.E., Schladow, S. G., Winder, M., and Oris, J.T. Patterns of spatial and  
777 temporal variability of UV transparency in Lake Tahoe, California-Nevada. *Journal of*  
778 *Geophysical Research Biogeosciences*. Volume 114(G2). doi:10.1029/2008JG000816, 2009,  
779 2009b.
- 780 Saros, J. E., C. L. Osburn, R.M.Northington, S.D. Birkel, J.D. Auger, C. A. Stedmon, and N. J.  
781 Anderson. Recent decrease in DOC concentrations in Arctic lakes of southwest Greenland,  
782 *Geophys. Res. Lett.*, 42, 6703–6709, doi:10.1002/2015GL065075, 2015.
- 783 Sharp, J.H., and others. Procedures subgroup report. *Marine Chemistry*. Volume 41. 1993. pp 37-  
784 49, 1993.
- 785 Sherry, A. Discriminant analysis in counseling psychology research. *The Counseling*  
786 *Psychologist* 34: 661-683, 2006.
- 787 Sherry, A., and R. K. Henson. Conducting and interpreting canonical correlation analysis in  
788 personality research: A user-friendly primer. *Journal of Personality Assessment* 84: 37-48,  
789 2010.
- 790 Solomon, C. T., S. E. Jones, B. C. Weidel, and others. Ecosystem Consequences of Changing  
791 Inputs of Terrestrial Dissolved Organic Matter to Lakes: Current Knowledge and Future  
792 Challenges. *Ecosystems* 18: 376–389. doi:10.1007/s10021-015-9848-y, 2015.
- 793 Spencer, R.G.M., Guo, W. Raymond, P.A., Dittmar, T., Hood, E., Fellman, J., and Stubbins, A.  
794 Source and biolability of ancient dissolved organic matter in glacier and lake ecosystems on  
795 the Tibetan Plateau. *Geochimica et Cosmochimica Acta*. Vol 142:1-64-74, 2014.
- 796 Strock, K. E., J. E. Saros, S. J. Nelson, S. D. Birkel, J. S. Kahl, and W. H. McDowell. Extreme  
797 weather years drive episodic changes in lake chemistry: implications for recovery from  
798 sulfate deposition and long-term trends in dissolved organic carbon. *Biogeochemistry* 127:  
799 353–365. doi:10.1007/s10533-016-0185-9, 2016.
- 800 Tranvik, L. J., J. A. Downing, J. B. Cotner, and others. Lakes and reservoirs as regulators of  
801 carbon cycling and climate. *Limnology and Oceanography* 54: 2298–2314, 2009.
- 802 Tranvik, L. Carbon cycling in the Arctic. *Science* 345: 870–870. doi:10.1126/science.1258235,  
803 2014.
- 804 Vachon, D., C. T. Solomon, and P. A. del Giorgio. Reconstructing the seasonal dynamics and  
805 relative contribution of the major processes sustaining CO<sub>2</sub> emissions in northern lakes.  
806 *Limnology and Oceanography* 62: 706–722. doi:10.1002/lno.10454, 2016a.
- 807 Vachon, D., J.-F. Lapierre, and P. A. del Giorgio. Seasonality of photochemical dissolved  
808 organic carbon mineralization and its relative contribution to pelagic CO<sub>2</sub> production in  
809 northern lakes. *J. Geophys. Res. Biogeosci.* 121: 864–878. doi:10.1002/2015JG003244,  
810 2016b.
- 811 Westra, S., H. J. Fowler, J. P. Evans, and others. Future changes to the intensity and frequency of  
812 short-duration extreme rainfall. *Reviews of Geophysics* 52: 522–555.  
813 doi:10.1002/2014RG000464, 2014.
- 814 Weyhenmeyer, G. A., M. Fröberg, E. Karlun, M. Khalil, D. Kothawala, J. Temnerud, and L. J.  
815 Tranvik. Selective decay of terrestrial organic carbon during transport from land to sea.  
816 *Global Change Biology* 18: 349–355, 2012.
- 817 Weyhenmeyer, G. A., S. Kosten, M. B. Wallin, L. J. Tranvik, E. Jeppesen, and F. Roland.



- 818 Significant fraction of CO<sub>2</sub> emissions from boreal lakes derived from hydrologic inorganic  
819 carbon inputs. *Nature Geosci.* doi:10.1038/ngeo2582, 2015.
- 820 Weyhenmeyer, G. A., and D. J. Conley. Large differences between carbon and nutrient loss rates  
821 along the land to ocean aquatic continuum-implications for energy:nutrient ratios at  
822 downstream sites. *Limnology and Oceanography* 10: 141–11. doi:10.1002/lno.10589, 2017.
- 823 Wilkinson, G. M., M. L. Pace, and J. J. Cole. Terrestrial dominance of organic matter in north  
824 temperate lakes. *Global Biogeochemical Cycles* 27: 43–51. doi:10.1029/2012GB004453,  
825 2013.
- 826 Williams, C.J., Frost, P.C., Morales-Williams, A.M., Larson, J.H., Richardson, W.B., Chiandret,  
827 A.S. and Xenopoulos, M.A. Human activities cause distinct dissolved organic  
828 matter composition across freshwater ecosystems. *Glob Change Biol*, 22: 613–626.  
829 doi:10.1111/gcb.13094, 2016.
- 830 Williamson, C. E., D. P. Morris, M. L. Pace, and O. G. Olson. Dissolved organic carbon and  
831 nutrients as regulators of lake ecosystems: Resurrection of a more integrated paradigm.  
832 *Limnology and Oceanography* 44: 795–803. doi:10.4319/lno.1999.44.3\_part\_2.0795, 1999.
- 833 Williamson, C. E., J. E. Saros, W. F. Vincent, and J. P. Smol. Lakes and reservoirs as sentinels,  
834 integrators, and regulators of climate change. *Limnology and Oceanography* 54: 2273–2282,  
835 2009.
- 836 Williamson, C.E., J.A. Brentrup, J. Zhang, W.H. Renwick, B.R. Hargreaves, L.B. Knoll, E.P.  
837 Overholt, and K.C. Rose. Lakes as sensors in the landscape: Optical metrics as scalable  
838 sentinel responses to climate change. *Limnology and Oceanography* 59: 840–850, 2014.
- 839 Williamson, C. E., E. P. Overholt, R. M. Pilla, T. H. Leach, J. A. Brentrup, L. B. Knoll, E. M.  
840 Mette, and R. E. Moeller. Ecological consequences of long- term browning in lakes. *Sci.*  
841 *Rep.* 5. doi:10.1038/srep18666, 2015.
- 842 Williamson, C. E., E. P. Overholt, J. A. Brentrup, and others. Sentinel responses to droughts,  
843 wildfires, and floods: effects of UV radiation on lakes and their ecosystem services.  
844 *Frontiers in Ecology and the Environment* 14: 102–109. doi:10.1002/fee.1228, 2016.
- 845 Zwart, J. A., N. Craig, P. T. Kelly, S. D. Sebestyen, C. T. Solomon, B. C. Weidel, and S. E.  
846 Jones. Metabolic and physiochemical responses to a whole-lake experimental increase in  
847 dissolved organic carbon in a north-temperate lake. *Limnology and Oceanography* 61: 723–  
848 734. doi:10.1002/lno.10248, 2015.