Potential effects of deep seawater discharge by an Ocean Thermal Energy Conversion plant on the marine microorganisms in oligotrophic waters

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Abstract

Installation of an Ocean Thermal Energy Conversion pilot plant (OTEC) off the Caribbean coast of Martinique is expected to use approximately 100 000 m\textsuperscript{3} h\textsuperscript{-1} of deep seawater for its functioning. This study examined the potential effects of the cold nutrient-rich deep seawater discharge on the phytoplankton community before the installation of the pilot plant. Thermal effect induced by the deep seawater upwelled by the OTEC was described using the Regional Ocean Modeling System. Numerical simulations of deep seawater discharge showed that a 3.0 °C temperature change, considered as a critical threshold for temperature impact, was never reached during an annual cycle on the top 150 m of the water column on two considered sections centered on the OTEC. The thermal effect should be limited, less than 1 km\textsuperscript{2} on the area exhibited a temperature difference of 0.3 °C (absolute value). The impact on phytoplankton of the resulting mixed deep and surface seawater was evaluated by \textit{in situ} microcosm experiments. Two scenario of water mix ratio (2 \% and 10 \% of deep water) were tested at two incubation depths (deep chlorophyll-a maximum: DCM and bottom of the euphotic layer: BEL). The larger impact was obtained at DCM for the highest deep seawater addition (10 \%), with a development of diatoms, whereas 2 \% addition induced only a limited change of the phytoplankton community. This study suggested that the OTEC plant would significantly modify the phytoplankton assemblage only in the case of a discharge affecting the DCM and would be restricted to a local scale.

1. Introduction

Ocean Thermal Energy Conversion (OTEC) uses the solar energy by exploiting the temperature gradient between surface and bottom seawater. In an OTEC plant, the cold deep seawater pumped close to sea bottom is used to condense a working fluid (like ammonia), whereas warm surface waters, pumped close to the surface, serve to evaporate it. The difference of pressure, generated by the evaporation and condensation of the fluid, drives a turbine that produces mechanical energy.
This energy is then converted to electrical energy in a generator. Due to the need of a 20 °C difference between the cold deep and the warm surface waters for the OTEC exploitation, tropical areas are well suited for the installation of OTEC plants.

The Martinique, a tropical island of Lesser Antilles, is ideally suited for OTEC functioning with its narrow continental slope in the Caribbean part of the island, allowing an implementation of the plant close to the coast. The implementation of a 10 MW OTEC pilot plant off the Caribbean coast of Martinique is expected in 2020 as part of the French NEMO project (Akuo Energy, DCNS). This OTEC will pump approximately 100,000 m$^3$.h$^{-1}$ of deep seawater at 1100 m depth. In order to optimize the energy efficiency, the deep seawater should be rejected close to the surface. However, this large discharge could induce important disturbances on the upper ocean ecosystem, and this impact should be estimated.

Environmental assessment of OTEC functioning was studied since the 1980’s (NOAA, 1981; 2010). The deep seawater discharge was described as one of the major drivers impacting the marine environment in OTEC plant. However, only a few studies specifically detailed this critical aspect (Taguchi et al., 1987; Rocheleau et al., 2012). The deep seawater discharge in OTEC plant generates a phenomenon similar to the one naturally occurring in the ocean within upwelling systems. Equatorward winds along the coast in the eastern Atlantic and Pacific linked to atmospheric high-pressure systems force Ekman transport and pumping, relocating coastal surface waters offshore. Thereby, deep water transport towards the surface is generated close to the coast. In these systems, the large amount of macronutrients and trace metals carried to the euphotic zone by the enriched deep seawater supports a large development of the phytoplankton, making upwelling the most productive oceanic regions (Bakun, 1990; Pauly and Christensen, 1995; Chavez and Toggweiler, 1995; Carr and Kearns, 2003). The tropical surface waters off the Caribbean coast of Martinique exhibit low nutrients concentrations and can be significantly enriched by the deep seawater discharge. Whereas phytoplankton assemblages in upwelling systems are usually dominated by large phytoplankton and particularly by diatoms (Bruland et al., 2001; Van Oostende et al., 2015), the phytoplankton community in oligotrophic systems is composed of smaller organisms (Agawin et al., 2000).

Due to these important differences, it is thus of critical interest to investigate the potential effects of the deep seawater discharge of the planned OTEC plant on the phytoplankton community off Martinique.

In this study, the impact of deep seawater discharge on the thermal structure of surface waters was first assessed. Modification of the surface waters stratification should indeed impact the phytoplankton community. It is crucial to provide a depth where the deep seawater could be
A high-resolution oceanic model was used to examine the thermal impact induced by the deep seawater dispersion. Eight configurations of discharge depth were tested, corresponding to the deep chlorophyll-a maximum (DCM), the bottom of the euphotic layer (BEL) and five depths below the BEL. Temperature differences between numerical simulations without and with the deep seawater discharge were compared on the upper 150 m of a vertical section.

The distribution of the ambient phytoplankton community and the biogeochemical properties of the deep and surface seawater mixture that could impact the phytoplankton community were then described. Phytoplankton distribution and assemblage were detailed in order to assess short time and small scales variabilities of phytoplankton assemblage and primary production in the study site.

Finally, in order to simulate the OTEC deep seawater input, enrichment experiments were conducted on the future site of the pilot plant. Enrichment experiments are commonly used in oceanography to assess the effects on phytoplankton community and primary production. For example, large iron (Fe) enrichment experiments were conducted from 1993 to 2005 to estimate the potential of Fe limitation on ocean primary production (De Baar et al., 2005; Boyd et al., 2007). Several experiments also showed that macro- and micro-nutrients enrichments induce changes in the phytoplankton community in upwelling regions (Hutchins et al., 2002) as well as in oligotrophic regions (Kress et al., 2005). Enrichment experiments were usually conducted with mesocosms immersed close to the surface (Escaravage et al., 1996; Duarte et al., 2000) or in laboratory under artificial light and temperature using phytoplankton model species (Brzezinski, 1985). A laboratory experiment intended to evaluate the effects of an OTEC seawater discharge in Hawaiian waters on the natural phytoplankton community was previously conducted (Taguchi et al., 1987) under such artificial conditions, and thus, it could not totally reproduce what occurred in the natural environment. Other deep seawater discharge experiments were realized in situ (Aure et al., 2007; Handå et al., 2014). For example, the use of a moored platform to upwell deep seawater and discharge it close to the surface has shown an increase in primary production in a western Norwegian fjord where the euphotic zone is nutrient-depleted during summer (Aure et al., 2007), as it would be expected with the OTEC discharge. Whereas such a pumping system is well adapted for pumping seawater at 30 m depth for example, it cannot be applied for OTEC experiments where deep seawater must be collected far deeper (1100 m depth) and also discharged more deeply in the water column to reduce the potential effects on the phytoplankton community. These conditions can be obtained by the use of in situ microcosms, in which light and temperature are the same as in the natural surrounding waters, avoiding additional bias, and several conditions (enrichment, incubation...
depth) can be simulated. Therefore, we used the unique device of immerged microcosms we developed (Giraud et al., 2016) for assessing the effects of deep seawater discharge on the phytoplankton community. Two incubation depths (DCM and BEL) with two ratios of enriched seawater (mixtures of surface water with 2 % and 10 % of deep seawater) were tested. These experiments allowed the evaluation of critical mixing rate and discharging depth where effect was maximal.

2. Materials and methods

2.1. Modelling the thermal effect

The hydrodynamic numerical model ROMS-Regional Ocean Model System (Shchepetkin and McWilliams, 2005; 2009) was used to describe the resulting thermal effect due to OTEC functioning. The model was run in a 2-ways AGRIF configuration allowing to define a parent and child domains around the Martinique Island which are run simultaneously, transferring automatically open boundary conditions. The parent grid ranges from 63° W to 59° W and 13° N to 15.9° N with a resolution of 1/60° (around 1.8 km) while the child domain narrows the parent one and was from 61.74° W to 60.41° W and 14.21° N to 15.11° N with a resolution down to 1/180° (around 600 m). The bottom topography and coastline are interpolated from the GINA database (1/120°, www.gina.alaska.edu/data/gtopo-dem-bathymetry) (Fig. 1).

The model is forced by the monthly Climate Forecast System Reanalysis (NCEP-CFSR) for wind stress, heat and freshwater fluxes. For the open boundary conditions and initial conditions of the parent domain, a monthly climatology computed from the Simple Ocean Data Assimilation (SODA) reanalysis (Carton and Giese, 2008) was used for the dynamical variables (temperature, salinity and velocity fields). The NCEP-CFSR products do no cover the period of our mesocosm experiments (November 2013 and June 2014). The simulations were thus performed over another period when the atmospheric forcing was available. We choose the 3 years period of 1998-2000, using 1998 and 1999 as a spin-up and the last year 2000 to analyze the thermal structure and circulation field. Model outputs were stored as daily averages. The configurations were run without and with a deep seawater discharge mimicking the OTEC functioning. The deep seawater discharge was initiated on January 1st 2000. Eight cases of horizontal discharge settings were simulated at different depths: 1) the DCM (45 m), 2) the BEL (80 m), that were estimated on June 12th 2014, and 3) six depths below the euphotic zone (110 m, 140 m, 170 m, 250 m, 350 m and 500 m). In the OTEC plant, deep water will be pumped at 1100 m where temperature is around 5 °C and salinity 35. Circulation of this water through the plant system will warm it up until 8 °C prior to its release in the upper ocean. We thus
applied at the location of the OTEC plant (61°13'0" W, 14°35'48" N), a cold water discharge (temperature 8 °C, salinity 35) at a flow rate of 28 m³ s⁻¹ and with a northward orientation. The thermal impact of the cold-water source was assessed documenting the differences between simulations without and with the modelled OTEC plant functioning over the full year 2000.

2.2. Field observations and in situ experiments

2.2.1. Sampling and analytical methods
Temperature, salinity, and fluorescence profiles were performed using Seabird SBE19+ probe with in situ Fluorimeter Chelsea AQUAtracka III.

Seawater was collected in the water column in ultra-clean conditions (Giraud et al., 2016) to measure in situ parameters and to prepare the microcosms. Seawater and microcosms were sampled similarly in a land laboratory a few hours after collection.

Nitrate (NO₃⁻), nitrite (NO₂⁻), phosphate (PO₄³⁻) and silicate (Si(OH)₄) concentrations were determined in filtered waters (<0.6 μm PC membrane) stored at -20 °C until analysis using a Bran + Luebbe AAIII auto-analyzer (Aminot and Kérouel, 2007).

Filtered samples (0.2 μm; 300AC-Sartobran™ capsules) for dissolved trace metals determination were collected under pure-N₂ pressure (0.7 atm) in acid cleaned low density polyethylene bottles, acidified with ultrapure HCl (pH < 2) and stored in two plastic bags in dark at ambient temperature. Concentrations of dissolved trace metals (cadmium, Cd; lead, Pb; iron, Fe; zinc, Zn; manganese, Mn; cobalt, Co; nickel, Ni; and copper, Cu) were determined in UV-digested samples by ID-ICP-MS (Milne et al., 2010) after preconcentration on a WAKO resin (Kagaya et al., 2009) using an Element XR ICP-MS. Blanks, limits of detection, accuracy and precision (assessed using reference samples) of the ID-ICP-MS method are reported in Table 1. The values determined by ID-ICP-MS were in excellent agreement with the consensus values, apart for Cd that yielded higher concentration in S-SAFE reference sample than the consensus value (Table 1).

The pH was determined using a pH ultra-electrode (pHC28) mounted on a HQ40d multi pH-meter (HACH) with an accuracy of ± 0.002 pH unit in samples preserved with saturated HgCl₂ in glass bottles hermetically closed with Apiezon grease, sealed with Parafilm® and stored in the dark at ambient temperature.

Three complementary methods were used to analyze the phytoplankton community. Pigment signatures were measured by HPLC (using an Agilent Technologies 1100-series) on polysulfone filters (0.22 μm pore-size) frozen at -20 °C and stored in liquid nitrogen, after internal standard addition (vitamin E acetate) and extraction in a 100 % methanol solution (Hooker et al., 2012). Fifty pigments were identified and associated to phytoplankton groups (Uitz et al., 2010). Identification
and enumeration of pico-phytoplankton were realized by flow-cytometry using a BD-FACSVerse™ (Marie et al., 1999) in samples preserved in cryotube with addition of 0.25 % glutaraldehyde frozen at -20 °C and stored in liquid nitrogen. Four groups of pico-phytoplankton were identified: Prochlorococcus, piceukaryotes (< 10 μm), and 2 groups of Synechococcus discriminated, respectively, by their low and high phycourobilin (PUB) to phycoerythrobilin (PEB) ratios. Taxonomic identification and enumeration of micro-phytoplankton (20 - 200 µm) and a part of nano-phytoplankton (2-20 µm) (Dussart, 1966) were carried out using an inverted microscope (Wild M40) in samples preserved with neutral lugol solution. Utermöhl settling chambers (Hasle, 1988) were used for micro-phytoplankton analyses, and a smaller sedimentation chamber (2.97 mL) for the analyses of nano-phytoplankton. When possible, phyttoplankton was identified to the lowest possible taxonomic level (species, genus or group). Biovolume of each species was also estimated from these microscope analyses (Hillebrand et al., 1999).

2.2.2. In situ microcosm experiments

The potential impact of deep seawater discharge on the phytoplankton community was simulated by in situ microcosm incubations of various deep and surface seawater mixing (Giraud et al., 2016). The experiments were conducted from 12th (D0) to 19th (D7) of June 2014. The deep and surface seawaters were collected at the site of the future OTEC pilot plant (61°11'52“ W-14°37'57” N; Fig. 1). Microcosms bottles were incubated on two stainless steel structures set at the depths of deep chlorophyll-a maximum (DCM) and at the bottom of the euphotic layer (BEL) on a mooring chain located, for practical reasons, closer to the coast (61°10’9“ W-14°39’8“ N, seafloor at 220 m depth) during 6 days (Giraud et al., 2016). Seawater was collected at D0 at the depths of DCM (45 m depth) and BEL (80 m depth) identified on the future OTEC site from the fluorescence profile, and close to the bottom (1100 m depth corresponding to the pumping depth of the future OTEC plant) in ultra-clean conditions. Deep seawater was mixed in three proportions (0 % as a control hereafter referred to as “Control”, 2 % as a low input called “2 % of deep seawater”, and 10 % as a large input called “10 % of deep seawater”) with DCM and BEL waters. Each resulting mixture was distributed in 2.3 L polycarbonate bottles filled up to overflow level, of which four replicates per mixing condition per depth were immersed at their respective sampling-depth for 6 days; duplicates per mixing condition per depth were kept in dark at 25 °C for a few hours until sampling for later characterization of phytoplankton assemblage and biogeochemical properties at D0 (called “Surrounding waters D0”); and duplicates per mixing condition per depth were used to estimate carbon and NO3- uptakes at D0 (called “Surrounding waters D0”) as described below.
Same sampling and mixtures were realized at day 6 (D6, June 18th) just to evaluate the temporal evolution in the natural environment, resting on duplicate bottles per mixing condition per depth for phytoplankton and biogeochemical characterizations at D6 (called “Surrounding waters D6”) and using other duplicates to estimate carbon and NO$_3^-$ uptakes at D6 (called “Surrounding waters D6”). After the 6 days incubation, all the incubated microcosm bottles on the mooring (called “Microcosm D6”) were brought on board. A quarter of each four replicates per condition was put in a new 2.3 L clean bottle and used to estimate carbon and NO$_3^-$ uptakes after 6 days of incubation (called “Microcosm D6”). The remaining microcosm contents were kept for sampling and analysis.

2.2.3. Carbon and nitrate uptakes

Carbon (primary production) and NO$_3^-$ uptake rates were estimated in the same sample using the dual $^{13}$C/$^{15}$N isotopic label technique (Slawyk et al., 1977). Immediately after sampling, $^{13}$C tracer (NaH$^{13}$CO$_3$, 99 atom%, Eurisotop, 0.25 mmol$^{13}$C mL$^{-1}$) and $^{15}$N tracer (Na$^{15}$NO$_3$, 99 atom%, Eurisotop, 1 µmol$^{15}$N mL$^{-1}$) were added to seawater mixtures at 10:1 v/v ratio. The initial enrichment was 10 atom% excess of $^{13}$C for the bicarbonate pool and 16-95 atom% excess of $^{15}$N for the NO$_3^-$ pool depending on the ambient NO$_3^-$ concentration. The $^{13}$C/$^{15}$N amended bottles were incubated for 24 h on the mooring line at the DCM and BEL depths, after which 1 L samples were filtered onto precombusted (450 °C, 4 h) glass fiber filters (Whatman). Filters were stored at -20 °C and oven dried (60 °C, 24 h) prior to analysis. Concentrations of carbon (POC), nitrogen (PON) as well as $^{13}$C and $^{15}$N enrichments in particulate matter were measured with a mass spectrometer (Delta plus, ThermoFisher Scientific) coupled to a C/N analyzer (Flash EA, ThermoFisher Scientific). Standard deviations were 0.009 µM and 0.004 µM for POC and PON, and 0.0002 atom% and 0.0001 atom% for $^{13}$C- and $^{15}$N-enrichments, respectively.

The absolute uptake rates ($\rho$, in µmol L$^{-1}$ h$^{-1}$) were calculated for nitrogen (Dugdale and Wilkerson, 1986) and carbon (Fernández et al., 2005) using the particulate organic concentrations measured after 24 h of incubation. These rates were converted into biomass specific uptake rates ($V$, in µmol µmol POC or PON$^{-1}$ h$^{-1}$) by dividing $\rho$ by POC or PON. The addition of $^{15}$N tracer would cause a substantial increase in dissolved inorganic nitrogen concentrations especially in the surface waters and, in turn, an overestimation of uptake rates (Dugdale and Wilkerson, 1986; Harrison et al., 1996). The NO$_3^-$ uptake rates were corrected for this perturbation (Dugdale and Wilkerson, 1986) using a half-saturation constant of 0.05 µmol L$^{-1}$ characteristic for nitrogen-poor oceanic waters (Harrison et al., 1996) and the measured NO$_3^-$ concentration. Overestimation was low (< 5 %) in samples with an addition of deep seawater but it was of about 50 % in samples without deep seawater addition. The uptake rates measured in these samples represented therefore estimations rather than actual values.
2.2.4. Statistical analyses

Kruskal-Wallis test was applied on the set of pigments concentrations, pico-phytoplankton abundances and macronutrients concentrations. If significant differences (p < 0.05) were found, Mann-Whitney test was run to identify the samples significantly different. Statistical analyses were performed using Statgraphics Centurion XVI software.

3. Results

3.1. Modeling of the deep seawater discharge

3.1.1. Model evaluation

We compared modeled daily profiles (temperature, salinity) of June and November 2000 with in situ CTD data at OTEC station we recorded in June 2014 and November 2013 (Fig. 2 a-b).

In June, the modeled and observed vertical profiles of temperature were quite in agreement with a well mimicked thermocline depth. However, a warm bias of ~1.5 °C was simulated by the model in the top 50 m. Between 300 and 500 m depth, a cold bias of ~1.5 °C depth was also observed. The modeled and experimental salinity profiles presented a similar pattern. However, the salinity was largely overestimated by the model in the top-120 m, especially in the upper 60 m (by ~2 units), as compared to field observations. Between 120 m and 150 m, the model slightly underestimated the salinity.

In November, the thermocline and halocline were well reproduced with modeled vertical profiles of temperature and salinity, in good agreement with observations. However, temperature was slightly overestimated by the model, with warm bias of ~0.8 °C. At deeper depths, the modeled and observed temperatures were in excellent agreement. Salinity was underestimated by the model within the top 50 m by ~1 unit, and between 70 and 200 m depths by at maximum 0.3 units. Below 200 m depth, modeled and observed salinities exhibited similar profiles.

ADCP measurements (horizontal velocity and direction of currents) were made by our DCNS partner at the study site for a feasibility study, but in June 2011 (between 40 m and 800 m depths).

ADCP data were compared to model outputs for June 2000 (Fig. 3). Current directions were quite similar between model outputs and ADCP data with a mean direction toward the South/South-East.

The horizontal velocity norm was also quite close between both data sets with larger velocity close to the surface at ~50 m depth. Larger difference appeared in subsurface in ADCP data but similar trends were observed and values were relatively close.

Modeled physical properties (temperature, salinity, currents) were therefore quite similar to those directly observed at the study site. The small differences observed between model and field data are...
likely due to inter-annual variability since years examined were indeed different for the model simulation (2000) and the field data (2011, 2013 and 2014).

### 3.1.2. Impact of the deep seawater discharge on the thermal structure in surface

In order to assess the deep seawater discharge impact on the thermal structure of the upper 150 m of the water column, the dispersion of temperature differences ($\Delta T$ in °C) obtained without and with the deep seawater discharge in the model outputs was examined on two vertical sections. A section of 124 km for the large domain (corresponding to the child domain) and another section of 10 km for the near-OTEC domain (defined from 61.24° W to 61.17° W and 14.60° N to 14.67° N) were defined, both centered on the OTEC site and parallel to the coast (Fig. 1). Presently, there are no environmental standards defining threshold levels for temperature difference that will be induced by an OTEC deep seawater discharge. So, the study relied on the World Bank Group prescriptions for liquefied natural gas facilities which set at 3 °C the temperature difference limit at the edges of the zone where initial mixing and dilution take place (IFC, 2007). We thus considered for each discharge depth the cooling and warming outputs from the model, which exhibit a $|\Delta T| \geq 3$ °C. Areas (in % of the considered domain) impacted by these cooling and warming effects were added (absolute values) in order to compare the potential impact of each discharge depth configuration. None of the discharge depth configurations could produce a modification of the thermal structure of the top 150 m of the water column, higher than or equal to the considered temperature threshold ($|\Delta T| \geq 3$ °C), for both domains sections. Then, a lower temperature difference of 0.3 °C (absolute value) was considered. This temperature difference represented a low threshold as compared to the World Bank Group prescriptions (IFC, 2007) that instead represent a high threshold. The areas exhibiting a $|\Delta T| \geq 0.3$ °C in the top 150 m (Table 2) were extremely small (< 1 km²) and were not significantly different in both sections and at the different discharge depths, on an annual average and in June (our experimental period).

### 3.2. Biogeochemical properties and phytoplankton community

#### 3.2.1. Expected biogeochemical properties of the resulting mixed waters

The pH was very similar at the DCM and BEL at the OTEC site on D6 (8.24 and 8.25, respectively), whereas deep seawater-pH showed lower value (7.81). The addition of 2 % and 10 % deep seawater to surface waters could thus induce a pH-decrease of respectively, 0.01 and 0.07 unit. Hence, the effect on pH could be rather limited compared to the 0.1 pH decrease (from 8.2 to 8.1) between the pre-industrial time and the 1990’s [39].
and PO$_4^{3-}$ concentrations (Table 3) were below the detection limit (< 0.02 µM) at the DCM (55 m) and BEL (80 m) at the OTEC site on observational D4 (June 16$^{th}$ 2014), whereas Si(OH)$_4$ concentrations were above detection limit (> 0.08 µM), particularly at the DCM (2.4 µM). NO$_2^-$ concentrations showed the highest values at the BEL whereas they were negligible at the DCM (<0.02 µM). In deep seawater, as commonly observed, NO$_3^-$, PO$_4^{3-}$ and Si(OH)$_4$ concentrations were largely higher compared to the surface (Table 3). The 2 % and 10 % deep water additions represented a large input for NO$_3^-$ (from <0.02 µM to 0.54 and 2.71 µM, respectively). If the 10 % ratio also induced a large input of PO$_4^{3-}$ (from <0.02 to 0.19 µM), the input of 2 % deep water was more limited (0.04 µM). The effect of 2 % or 10 % deep seawater addition was more limited for Si(OH)$_4$ relatively to NO$_3^-$ and PO$_4^{3-}$ input, yet it accounted for 50-63 % increase for 10 % deep seawater addition (Table 3). Finally, because deep and DCM waters were NO$_2^-$ depleted, the deep seawater input did not modify the NO$_2^-$ concentration at the DCM. At the BEL, NO$_2^-$ concentration was higher and the 10 % addition slightly diluted NO$_2^-$ at this depth.

Mn showed maximum concentrations in the surface layer on D4 at the OTEC site (Table 4) decreasing with depth as observed close to the Lesser Antilles in the Atlantic Ocean (Mawji et al., 2015), but the measured surface concentrations were particularly high, especially at the DCM. Fe that commonly dispatches hybrid distribution combining a nutrient-type profile in surface waters and a scavenged-type distribution in deep waters (Bruland, 2003) also exhibited high surface values, particularly at the DCM (Table 4). Cd, Zn, Co, Ni, and Cu dispatched nutrient-type profiles, whereas Pb exhibited scavenged-type profile (Nozaki, 1997; Gruber, 2008), but like for dissolved Fe and Mn, their concentrations in the upper waters were particularly high (Table 4). For all trace metals at both depths, the 2 % deep seawater addition will not induce significant changes in their surface concentrations (Table 4). The 10 % deep seawater addition could increase Cd, Ni and Zn concentrations in surface waters (Table 4), whereas it would not constitute an input of Pb, Cu, Co, and Fe, and it can even dilute Mn (Table 4).

The surface waters can thus be enriched in macronutrients (NO$_3^-$, PO$_4^{3-}$) when submitted to a deep seawater discharge (particularly with 10 % deep seawater addition) in proportion depending on the depth. The same scheme can be applied in some of the dissolved trace metals (Cd, Ni, Zn) when a large ratio of deep seawater (10 %) is discharged.

### 3.2.2. Phytoplankton community in the natural environment

A set of seven accessory pigments identified as biomarkers of specific taxa (Uitz et al., 2010; Table 5) were analyzed at OTEC station at D0, D4 and D6 in surrounding surface waters (Fig. 4), as well as population abundance and their biovolume using light microscopy (Fig. 5).
The total chlorophyll a (TChl a defined as the sum of chlorophyll a and divinyl chlorophyll a), a proxy of the phytoplankton biomass, was higher at DCM than at BEL, as usually observed, by about two-folds. The fucoxanthin (biomarker of diatoms) concentrations were similar at the DCM and BEL on D0 (Fig. 4), like the total abundance of diatoms (Fig. 5). Fucoxanthin concentration increased by D4 and then by D6 at the DCM, corresponding to increases of cumulated diatoms biovolume on D4 (Fig. 5) and of diatoms abundance on D6 (Fig. 5). Peridinin, a biomarker of dinoflagellates, was detected at the DCM unlike at the BEL, with relatively high abundance and biovolume of dinoflagellates (Fig. 5). The 19’-hexanoyloxyfucoxanthin (biomarker of haptophytes) concentration (Fig. 4) and the prymnesiophytes (haptophyte) abundance and biovolume (Fig. 5) showed higher values at the DCM than at the BEL only at D4.

At the DCM, dinoflagellates largely dominated the nano- and micro-phytoplankton assemblage with the largest abundance and biovolume. Whereas prymnesiophytes showed the second highest abundance, its biovolume was very low, on the contrary to diatoms that dispatched lower abundance but higher biovolume (Fig. 5). At the BEL, dinoflagellates, prymnesiophytes and diatoms showed similar abundance, dinoflagellates and the diatoms occupied the major part of the total biovolume. Three groups of dinoflagellates were observed by light microscopy but they could not be identified at species level. However, their small size and the lack of colored starch (using lugol) in the cytoplasm suggested they were mixotrophic or heterotrophic population. Furthermore, the low concentrations of peridinin in samples supported this assumption.

At both depths, light microscopy analyses suggested that the large cyanobacteria, mainly Trichodesmium sp., were low in abundance and biovolume. Flow cytometry identification and count indicated that the small cyanobacteria Prochlorococcus dominated the pico-phytoplankton assemblage, but they showed a significant decrease from D0 to D6 (Fig. 6). A significant portion of Synechococcus was also observed while picoeukaryotes were poorly represented. Both Prochlorococcus and Synechococcus showed higher abundance at the DCM than at the BEL (by 65 % and 86 %, respectively), in line with the pigments analyses of zeaxanthin (biomarker of cyanobacteria) and total chlorophyll b concentrations (prochlorophytes).

3.2.3. Primary production and nitrate uptake in the natural environment
The phytoplankton distribution and assemblage can partly drive the intensity of primary production, so the specific uptake rate of carbon (V_C; Fig. 7) and NO3- (V_NO3) were estimated at D0 and D6.
V<sub>C</sub> in surrounding surface waters was relatively low at D0 (Fig. 7) indicating low primary production in these poor-nutrients waters. Yet, V<sub>C</sub> was approximately four-times higher at the DCM (2.10<sup>-3</sup> h<sup>-1</sup>) than at the BEL (5.10<sup>-4</sup> h<sup>-1</sup>) at D0, but drastically decreasing on D6 at the DCM (to ~6.10<sup>-4</sup> h<sup>-1</sup>). V<sub>N<sub>NO3</sub></sub> were also very low at D0 (1.10<sup>-3</sup> h<sup>-1</sup> at DCM, 4.10<sup>-4</sup> h<sup>-1</sup> at BEL) and drastically decreased at D6, below the detection limit (data not shown).

3.3. Impacts on the phytoplankton community of the deep seawater discharge

3.3.1. Changes in the phytoplankton assemblage

At the DCM, TChl a was similar in all treatments (p < 0.05) after 6 days of incubation in microcosms (Fig. 8). Only fucoxanthin and 19'-butanoyloxyfucoxanthin showed significant (p < 0.05) higher concentrations in 10 % enrichments as compared to controls, indicating higher abundance and/or biovolume of diatoms and haptophytes. The other diagnostic pigments did not show any significant difference between enriched microcosms and controls. Picoeukaryotes and *Synechococcus* abundances did not show significant variations between the treatments (Fig. 9a). Reversely, *Prochlorococcus* population showed higher (p < 0.05) abundance both in 2 % and 10 % enriched microcosms as compared to controls (Fig. 9a).

At the BEL, after the 6 days incubation period, pigments concentrations were below the detection limit indicating very low abundance of phytoplankton. Pico-phytoplankton did not show significant variations between the treatments and the controls (Fig. 9b). Pico-phytoplankton were clearly much less abundant at the BEL (< 1000 cells mL<sup>-1</sup>) than at DCM (Fig. 9b), 20-times even lower than that observed in surrounding waters at this depth on D6. For comparison, total abundance at the DCM was ~5-times lower in incubated microcosms on D6 compared to surrounding surface waters.

3.3.2. Changes in the primary production and nitrate uptake

Deep water inputs (2 % and 10 %) to surrounding waters collected at the DCM on D0 led to an increase of V<sub>C</sub> within 24 h compared to the controls (by 43 % and 48 %, respectively; Fig. 7); but they had no effect on D6 despite very low value in natural waters at this depth (6.10<sup>4</sup> h<sup>-1</sup>). The 6 days incubated microcosms showed very low V<sub>C</sub> in all treatments (Fig. 7). At the BEL, V<sub>C</sub> were quite similar on D0 and D6 and after 6 days of incubation, without significant differences between the treatments (Fig. 7). V<sub>N<sub>NO3</sub></sub> measured in microcosms after a 6-days *in situ* incubation were below the detection limit (data not shown).
4. Discussion

4.1. Natural variabilities in the oligotrophic area

4.1.1. Modeling of the deep seawater discharge

Salinity field data showed large seasonal variations, with low values in June 2014 (34.6 on the top 50 m) and much higher values in November 2013 (35.5 on the top 50 m). The model run for year 2000 did not fully reproduce these variations. Indeed, salinity was overestimated by the model in June whereas it was underestimated in November. The observations we made at the OTEC station showed that the low salinity observed in June was associated with high Si(OH)$_4$ concentrations. High Si(OH)$_4$ levels in fresher seawater have been already reported in surface waters in the Caribbean Sea and they were attributed to Amazon and Orinoco fresh rivers inputs (Steven and Brooks, 1972; Moore et al., 1986; Muller-Karger et al., 1995; Hu et al., 2004). Fresh surface waters enriched in Si(OH)$_4$ (Moore et al., 1986; Edmond et al., 1981) can be transported from the Amazon and Orinoco rivers towards the Caribbean Sea by the North Brazil Current and the Guiana Current (Muller-Karger et al., 1988, 1995; Osborne et al., 2014, 2015). It is likely that the rivers discharges and thus its inputs in the Caribbean Sea were quite different between 2000 (modeled year) and 2014 (in situ observations), thus explaining the discrepancy between modeled and observed salinities. Meso- and submeso-scale features resulting from the rivers flows could also induce short-term variability in the area and then could explain the observed differences.

4.1.2. Biogeochemistry and phytoplankton community structure

The very low PO$_4^{3-}$ and NO$_3^-$ concentrations recorded in the oligotrophic surrounding surface waters were likely favorable to the development of small phytoplankton, especially to the cyanobacteria as shown with the significant occurrence of Prochlorococcus in these waters, which are typical of poor nutrient waters (Partensky et al., 1999). In line with the very low $V_{NO3}$ measured here, it has been shown that $V_{NO3}$ by Prochlorococcus represents indeed only 5-10 % of its nitrogen uptake whereas reduced nitrogen substrates (NO$_2^-$, ammonium, and urea) uptake accounts for more than 90-95 % (Casey et al., 2007). By contrast, the development of larger phytoplankton taxa (particularly diatoms), which have higher NO$_3^-$ and PO$_4^{3-}$ requirements for their growth, were probably limited by these elements. Actually, NO$_3^-$ and PO$_4^{3-}$ concentrations in surrounding waters at the DCM were both lower than the detection limit (< 0.02 µM at D4) which is much lower than the average values of half-saturation constants for diatoms (1.6 ± 1.9 µM for NO$_3^-$ and 0.24 ± 0.29 µM for PO$_4^{3-}$; Sarthou et al., 2005). For Si(OH)$_4$, surrounding surface concentrations at DCM (2.39 µM) were in the range of diatoms half-saturation constants (3.9 ± 5.0 µM; Sarthou et al., 2005), hence the
diatoms development was probably not limited by Si(OH)$_4$. Furthermore, diatoms showed low abundance in spite of relatively high Si(OH)$_4$ and dissolved trace metals (in particular Fe) concentrations in surface waters. The potential of Fe limitation on phytoplankton community has been reported previously in upwelling systems, with an apparent half-saturation constant for diatoms growth of 0.26 nM Fe in the Peru Upwelling system (Hutchins et al., 2002). This constant is far lower than the concentration of Fe measured in surrounding waters at DCM (1.08 ± 0.03 µM at D4), suggesting that diatoms were probably not limited by Fe. This further supports growth limitation of diatoms by NO$_3$ and/or PO$_4$.$^3$-

Advection of waters from Amazon and Orinoco rivers can explain the relatively high Si(OH)$_4$ observed in the Caribbean Sea. However, little information is available on the input of trace metals by these waters into the Caribbean Sea. Amazon river can be a source of dissolved Fe, Cu, Ni, Pb and Co for the western-subtropical North Atlantic (Tovar-Sanchez and Sañudo-Wilhelmy, 2011), but this input can decrease rapidly away from its source like it has been shown for Co in the Western Atlantic (Dulaquais et al., 2014). Those inputs into the Caribbean Sea will have to be further examined, especially for Fe, Cd, Ni, Zn, Mn whose relatively high concentrations were detected in the Si(OH)$_4$-enriched surface waters of this study. Additionally, other inputs of trace metals such as atmospheric deposition can also increase surface concentrations, and those inputs can be substantial (Shelley et al., 2012).
4.2. Impact of deep seawater discharge

4.2.1. Temperature effects
The numerical simulation showed that the area impacted in the top-150 m by a temperature difference larger than or equal to 0.3 °C (absolute value) was lower than 1 km$^2$ (~2-3 % of the considered domain) and was insensitive to the injection depth or to the size of the tested domain (Table 2). This suggests that temperature difference might rather be linked to internal variability of the system. Since the effect of the discharge appears undetectable within 2-3 % variation of the model, it can be deduced that in a worst-case scenario, only 3 % of the small domain (300 m along the section, down to the 150 m depth) would be impacted by a temperature difference larger than or equal to 0.3 °C (absolute value). The impact of a 0.3 °C temperature variation on the growth of diatoms, notably on Pseudo-nitzschia pseudodelicatissima species that were observed in our study area, is limited to a change in the growth rate of 0.03 d$^{-1}$ [61]. For Synechococcus, a 0.3 °C variation of the temperature would also have a limited impact on the growth, with a variation of only 0.02 d$^{-1}$ (Boyd et al., 2013), like for Emiliania huxleyi (coccolithophyceae) for which the induced variation of maximum growth rate will be lower than 0.01 d$^{-1}$ (Fielding, 2013). The thermal effect on the phytoplankton assemblage could thus be considered negligible.

4.2.2. Impact on the phytoplankton community
Microcosms enrichment of DCM waters with 10 % of deep seawater led after 6 days to a significant increase (p < 0.05) of fucoxanthin (diatoms) and 19'-butanoyloxyfucoxanthin (haptophytes) by 71 % and 77 %, respectively, as compared to the controls. If the 2 % enrichment also showed similar trends, the differences of diagnostic pigments concentrations were not significant. NO$_3^-$ and PO$_4^{3-}$ concentrations induced by 10 % deep-water input on D0 (2.57 ± 0.13 µM and 0.14 ± 0.2 µM, respectively; Giraud et al., 2016) were close to NO$_3^-$ and PO$_4^{3-}$ half-saturation constants of diatoms (1.6 ± 1.9 µM and 0.24 ± 0.29 µM, respectively; Sarthou et al., 2005). The 10 % enrichment could thus support a development of diatoms. On the contrary, NO$_3^-$ and PO$_4^{3-}$ enrichments induced by 2 % addition of deep-water were too low (0.57 ± 0.02 µM and 0.04 ± 0.00 µM, respectively; Giraud et al., 2016) compared to these half-saturation constants to support the diatoms development. Therefore, the diagnostic pigments suggested a significant response proportionally to the amount of added deep seawater. Prochlorococcus were also more abundant (p < 0.05) in 2 % and 10 % treatments as compared to the controls. This lack of further Prochlorococcus population increase in 10 % treatments could be
attributed to a higher grazing pressure by haptophytes and/or to NO$_3^-$ and PO$_4^{3-}$ too rich conditions (Giraud et al., 2016).

Phytoplankton assemblage widely evolved in surrounding waters, from a predominance of pico-phytoplankton (*Prochlorococcus*) on D0 towards a higher abundance of micro-phytoplankton (diatoms) on D6. In order to assess if the impact on the phytoplankton assemblage due to 10% deep seawater addition (with a shift towards the diatoms) was in the range of the natural variation observed in the surrounding surface waters, 10% deep seawater microcosms phytoplankton assemblage was compared to the natural phytoplankton assemblage.

Whereas microcosm controls showed a lower *Prochlorococcus* abundance (Fig. 9a) than surrounding surface waters on D6 ($p < 0.05$), the 10% microcosms additionally showed, higher fucoxanthin (diatoms) and 19′-butanoyloxyfucoxanthin (haptophytes) by about 142% and 317% (Fig. 8), respectively, as compared to natural waters at D6. Furthermore, 10% enrichments showed a fucoxanthin increase over the 6 days period by 3-times higher than in surrounding waters, whereas controls only showed an increase by 1.5-times higher than in surrounding waters. Therefore, it can be concluded that the 10% deep seawater enrichment induced higher variations of the phytoplankton assemblage than those observed from D0 to D6 in surrounding surface waters.

$V_c$ were higher ($p < 0.05$) both in 2% and 10% enrichments on D0 as compared to controls, suggesting a positive response of phytoplankton to the deep seawater addition. Conversely, there was no carbon-uptake rate difference ($p < 0.05$) between controls and enriched waters (with 2% and 10% of deep seawater) at D6 with the 6 days incubated microcosms, suggesting that the observed community modifications did not change the primary production. Indeed, the phytoplankton community was quite similar in surrounding surface waters on D6 and in 6 days-incubated microcosm controls. Thus, only the initial phytoplankton assemblage and initial primary production in surrounding surface waters would influence the response of the phytoplankton community and its production.

At the BEL, after 6 days of incubation, deep seawater addition experiments clearly showed lower effects on the phytoplankton community than at the DCM. Indeed, whereas significant differences ($p < 0.05$) between 10% enrichments and controls were observed in diagnostic pigments concentrations at the DCM, pigments concentrations were too low at the BEL to be quantified. It can be suggested that the lower population and lower carbon uptake could be related to the lowest light availability.

Overall, the phytoplankton response was proportional to the amount of added deep seawater. If the phytoplankton assemblage significantly varied over time in the environment, the 10% deep seawater enrichment showed larger variations (for diatoms and haptophytes) than those observed in
the natural environment. The DCM should be more impacted than the BEL by the deep seawater discharge even with a large deep seawater input. On the other hand, the impact on the primary production largely depended on the initial phytoplankton assemblage, which was quite variable over time. The modification of the phytoplankton community to a deep seawater input could also be depending on the initial phytoplankton community. For that, the microcosm experiments did not allow drawing a scenario over the long term of the potential modifications of the primary production and the phytoplankton community associated to the deep seawater discharge by an OTEC.

Light microscopy analyses showed a large abundance of dinoflagellates at the DCM (between 9,240 and 20,400 cells mL⁻¹ on D6 and D4; Fig. 5 a) which could be mixotrophic or heterotrophic and thus probably exert a grazing pressure on the phytoplankton, particularly on the pico-phytoplankton (Liu et al., 2002). However, in this study, the zooplankton larger than 200 µm and its potential control on the phytoplankton community were not considered and should be examined in future studies.

5. Conclusion

Two complementary approaches were applied to study the potential effects of the deep seawater discharge of the planned OTEC plant on the phytoplankton community off Martinique. Because the distribution and the development of phytoplankton are directly linked to the surface stratification, it is important to assess the thermal effect of deep seawater by an OTEC plant.

Modelling of the deep seawater discharge showed that the thermal structure of the top 150 m of the water column on large and near-OTEC sections should be very slightly impacted for the lowest considered temperature differences |ΔT| ≥ 0.3 °C. If World Bank Group prescriptions of not exceeding a higher temperature difference of 3 °C are followed, the environmental perturbations potentially caused by the operation of the OTEC should be considered negligible. The area where the 150 m-depth waters are impacted by the lowest considered temperature differences |ΔT| ≥ 0.3 °C would not exceed 1 km² in a worst-case scenario.

The phytoplankton community and its production could be impacted by a large deep seawater input. Whereas pico-phytoplankton currently largely dominates the phytoplankton assemblage, a ratio of 10 % of deep seawater in DCM waters could induce a shift toward the diatoms and micro-phytoplankton. The ratio of 2 % of deep seawater in DCM waters only showed significant higher Prochlorococcus abundance than controls, but the assemblage and the primary production were not modified by this lower input. The stimulation of Prochlorococcus could be due to one or some of the following causes: NO₃⁻ and/or PO₄³⁻ supply, trace metal supply, lowered pH (higher availability of dissolved inorganic carbon).
Although significant, these results would have to be extended to larger temporal scale, and the phytoplankton interactions with higher trophic levels (such as zooplankton) must be studied. Because no environment standards on the deep seawater discharge effects are available yet, a rigorous monitoring of the phytoplankton community, biogeochemical parameters distribution and of the water column stratification must be established as soon as the OTEC is implemented and during its continuous functioning.

Acknowledgements

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Table 1- Comparison of analyses of SAFe (Sampling and Analysis of iron) S and D2 reference samples (http://www.geotraces.org/science/intercalibration) between ID-ICPMS values (this study) and the consensus values. Our mean reagent blanks (based on all blank determinations) for dissolved Cd, Pb, Fe, Ni, Cu, Zn, Mn and Co, and detection limits of ID-ICPMS estimated as three times the standard deviation of the mean reagent blanks are also shown.

<table>
<thead>
<tr>
<th>SAFe D2</th>
<th></th>
<th></th>
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<th></th>
<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>This study</td>
<td>948.83 ± 65.95</td>
<td>28.86 ± 4.44</td>
<td>0.898 ± 0.098</td>
<td>8.60 ± 0.36</td>
<td>2.15 ± 0.16</td>
<td>7.29 ± 0.27</td>
<td>0.40 ± 0.05</td>
</tr>
<tr>
<td>Consensus values</td>
<td>986.00 ± 23.00</td>
<td>27.70 ± 1.50</td>
<td>0.933 ± 0.023</td>
<td>8.63 ± 0.25</td>
<td>2.28 ± 0.15</td>
<td>7.43 ± 0.25</td>
<td>0.35 ± 0.05</td>
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<tr>
<td>n=</td>
<td>20</td>
<td>20</td>
<td>18</td>
<td>19</td>
<td>22</td>
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<td>23</td>
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<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>This study</td>
<td>7.24 ± 1.57</td>
<td>48.42 ± 6.08</td>
<td>0.087 ± 0.025</td>
<td>2.56 ± 0.55</td>
<td>0.52 ± 0.06</td>
<td>0.07 ± 0.01</td>
<td>0.07 ± 0.01</td>
</tr>
<tr>
<td>Consensus values</td>
<td>1.10 ± 0.30</td>
<td>48.00 ± 2.20</td>
<td>0.093 ± 0.008</td>
<td>2.28 ± 0.09</td>
<td>0.52 ± 0.05</td>
<td>0.07 ± 0.01</td>
<td>0.79 ± 0.06</td>
</tr>
<tr>
<td>n=</td>
<td>25</td>
<td>25</td>
<td>27</td>
<td>27</td>
<td>15</td>
<td>15</td>
<td>10</td>
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<th>Detection Limit</th>
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<th></th>
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<tbody>
<tr>
<td>Blanks</td>
<td>0.716</td>
<td>1.809</td>
<td>0.061</td>
<td>0.040</td>
<td>0.019</td>
<td>0.129</td>
<td>0.003</td>
<td>0.32</td>
</tr>
</tbody>
</table>

Table 2- Area (km²) impacted in the top-150 m by a temperature difference |ΔT| ≥ 0.3 °C on two vertical sections centered on the OTEC, considering eight depths of deep seawater discharge (45, 80, 110, 140, 170, 250, 350, 500 m), average and root mean square for the year 2000 (from the monthly data) and for June 2000.

<table>
<thead>
<tr>
<th>Depth of deep water discharge</th>
<th>Mean Year 2000</th>
<th>June 2000</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Large domain</td>
<td>Near-OTEC domain</td>
</tr>
<tr>
<td>45 m</td>
<td>0.4 ± 0.4</td>
<td>0.0 ± 0.1</td>
</tr>
<tr>
<td>80 m</td>
<td>0.6 ± 0.7</td>
<td>0.1 ± 0.1</td>
</tr>
<tr>
<td>110 m</td>
<td>0.6 ± 0.5</td>
<td>0.0 ± 0.1</td>
</tr>
<tr>
<td>140 m</td>
<td>0.4 ± 0.5</td>
<td>0.1 ± 0.1</td>
</tr>
<tr>
<td>170 m</td>
<td>0.5 ± 0.8</td>
<td>0.0 ± 0.1</td>
</tr>
<tr>
<td>250 m</td>
<td>0.5 ± 0.7</td>
<td>0.1 ± 0.1</td>
</tr>
<tr>
<td>350 m</td>
<td>0.5 ± 0.5</td>
<td>0.1 ± 0.1</td>
</tr>
<tr>
<td>500 m</td>
<td>0.5 ± 0.5</td>
<td>0.1 ± 0.1</td>
</tr>
</tbody>
</table>
Table 3 - Nitrate, silicate, phosphate and nitrite concentrations on June 16\textsuperscript{th} 2014 (D4) at the deep chlorophyll maximum (DCM), at the bottom of the euphotic layer (BEL), and at the deep seawater pumping depth. Concentrations were measured at the OTEC site (0 % addition of deep waters) and calculated for 2 % and 10 % deep seawater additions.

<table>
<thead>
<tr>
<th>Depth (m)</th>
<th>Deep seawater ratio</th>
<th>([\text{NO}_3^-]) (µM)</th>
<th>([\text{Si(OH)}_4^-]) (µM)</th>
<th>([\text{PO}_4^{3-}]) (µM)</th>
<th>([\text{NO}_2^-]) (µM)</th>
</tr>
</thead>
<tbody>
<tr>
<td>DCM</td>
<td>0 %</td>
<td>&lt; 0.02</td>
<td>2.39</td>
<td>&lt; 0.02</td>
<td>0.02</td>
</tr>
<tr>
<td></td>
<td>2 %</td>
<td>0.54</td>
<td>2.88</td>
<td>0.04</td>
<td>0.02</td>
</tr>
<tr>
<td></td>
<td>10 %</td>
<td>2.71</td>
<td>4.82</td>
<td>0.19</td>
<td>0.02</td>
</tr>
<tr>
<td>BEL</td>
<td>0 %</td>
<td>&lt; 0.02</td>
<td>1.46</td>
<td>&lt; 0.02</td>
<td>0.32</td>
</tr>
<tr>
<td></td>
<td>2 %</td>
<td>0.54</td>
<td>1.96</td>
<td>0.04</td>
<td>0.32</td>
</tr>
<tr>
<td></td>
<td>10 %</td>
<td>2.71</td>
<td>3.98</td>
<td>0.19</td>
<td>0.29</td>
</tr>
<tr>
<td>1100</td>
<td>100 %</td>
<td>27.11</td>
<td>26.69</td>
<td>1.87</td>
<td>&lt; 0.02</td>
</tr>
</tbody>
</table>

Table 4 - Concentrations of dissolved trace metals (in nM): Mn, Fe, Cd, Zn, Co, Ni, Cu, Pb measured on June 16\textsuperscript{th} 2014 (D4) at the OTEC site at the DCM, BEL and 1100 m (0 % addition of deep waters), and their calculated concentrations in the mixtures with 2 % and 10 % addition of deep water.

<table>
<thead>
<tr>
<th>Depth (m)</th>
<th>Deep seawater ratio</th>
<th>Mn (nM)</th>
<th>Fe (nM)</th>
<th>Cd (nM)</th>
<th>Zn (nM)</th>
<th>Co (nM)</th>
<th>Ni (nM)</th>
<th>Cu (nM)</th>
<th>Pb (nM)</th>
</tr>
</thead>
<tbody>
<tr>
<td>DCM</td>
<td>0 %</td>
<td>2.97 ± 0.17</td>
<td>1.08 ± 0.03</td>
<td>0.03 ± 0.01</td>
<td>1.54 ± 0.04</td>
<td>0.05 ± 0.00</td>
<td>2.22 ± 0.10</td>
<td>1.70 ± 0.18</td>
<td>0.03 ± 0.00</td>
</tr>
<tr>
<td></td>
<td>2 %</td>
<td>2.92</td>
<td>1.08</td>
<td>0.04</td>
<td>1.56</td>
<td>0.05</td>
<td>2.29</td>
<td>1.70</td>
<td>0.03</td>
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<tr>
<td></td>
<td>10 %</td>
<td>2.71</td>
<td>1.09</td>
<td>0.07</td>
<td>1.63</td>
<td>0.05</td>
<td>2.60</td>
<td>1.71</td>
<td>0.03</td>
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<tr>
<td>BEL</td>
<td>0 %</td>
<td>1.65 ± 0.04</td>
<td>0.68 ± 0.03</td>
<td>0.03 ± 0.00</td>
<td>0.65 ± 0.03</td>
<td>0.03 ± 0.00</td>
<td>2.26 ± 0.17</td>
<td>1.14 ± 0.10</td>
<td>0.03 ± 0.00</td>
</tr>
<tr>
<td></td>
<td>2 %</td>
<td>1.63</td>
<td>0.69</td>
<td>0.04</td>
<td>0.68</td>
<td>0.03</td>
<td>2.34</td>
<td>1.15</td>
<td>0.03</td>
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<tr>
<td></td>
<td>10 %</td>
<td>1.52</td>
<td>0.73</td>
<td>0.08</td>
<td>0.82</td>
<td>0.03</td>
<td>2.64</td>
<td>1.21</td>
<td>0.03</td>
</tr>
<tr>
<td>1100</td>
<td>100 %</td>
<td>0.34 ± 0.02</td>
<td>1.22 ± 0.05</td>
<td>0.45 ± 0.01</td>
<td>2.39 ± 0.07</td>
<td>0.06 ± 0.00</td>
<td>6.00 ± 0.13</td>
<td>1.80 ± 0.08</td>
<td>0.02 ± 0.00</td>
</tr>
</tbody>
</table>

Table 5 - Definition of the diagnostic pigments used as phytoplankton biomarkers (taxonomic significance) and associated phytoplankton size class (Uitz et al., 2010).

<table>
<thead>
<tr>
<th>Diagnostic Pigments</th>
<th>Abbreviations</th>
<th>Taxonomic Significance</th>
<th>Phytoplankton Size Class</th>
</tr>
</thead>
<tbody>
<tr>
<td>Fucxanthin</td>
<td>Fuco</td>
<td>Diatoms</td>
<td>microplankton</td>
</tr>
<tr>
<td>Peridinin</td>
<td>Peri</td>
<td>Dinoflagellates</td>
<td>microplankton</td>
</tr>
<tr>
<td>19'-hexanoyloxyfucxanthin</td>
<td>Hex-fuco</td>
<td>Haptophytes</td>
<td>nanoplankton</td>
</tr>
<tr>
<td>19'-butanoyloxyfucxanthin</td>
<td>But-fuco</td>
<td>Pelagophytes and Haptophytes</td>
<td>nanoplankton</td>
</tr>
<tr>
<td>Alloxanthin</td>
<td>Allo</td>
<td>Cryptophytes</td>
<td>nanoplankton</td>
</tr>
<tr>
<td>chlorophyll b + divinyl chlorophyll b</td>
<td>TChlb</td>
<td>Cyanobacteria, Prochlorophytes</td>
<td>picoplankton</td>
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<tr>
<td>Zeaxanthin</td>
<td>Zea</td>
<td>Chlorophytes, Prochlorophytes</td>
<td>picoplankton</td>
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Figure captions

Figure 1- Bathymetry of the parent and child (grey rectangle) domains interpolated from the GINA data base with a zoom on the near domain (black rectangle); the oblique white and black lines represent the large and small sections, respectively, used for numerical simulations.

Figure 2- Comparison of temperature and salinity between model outputs and field data at the OTEC station (a) on June 16th 2000 and 2014, respectively and (b) on November 28th 2000 and 2013, respectively.

Figure 3- Comparison of mean current direction and horizontal velocity norm between model outputs from June 2000 and ADCP data from June 2011.

Figure 4- Pigment concentrations (from HPLC analysis) at the OTEC site at the DCM (a) and at the BEL (b), on June 12th (D0), 16th (D4), 18th (D6) 2014 (bars represent the standard deviation).

Figure 5- Abundance and biovolume of micro- and part of nano-phytoplankton at the OTEC site on June 12th (D0), 16th (D4), 18th (D6) 2014, at the DCM (a and c, respectively) and at the BEL (b and d, respectively) (bars represent the standard deviation).

Figure 6- Abundance of pico-phytoplankton at the DCM (a) and at the BEL (b), on June 12th (D0), 16th (D4), 18th (D6) 2014 (bars represent the standard deviation).

Figure 7- Specific carbon uptake rate (h⁻¹) at the DCM (a) and BEL (b) depths, on June 12th (D0), and 18th (D6), and in 6 days incubated microcosms (D6), for the three mixing conditions (0 %, 2 % and 10 % of deep seawater additions) (for surrounding waters bars represent the standard deviation for 3 replicates).

Figure 8- Diagnostic pigment concentrations in surrounding surface waters on D0 and D6, and in controls and deep water-enriched (2 % and 10 %) microcosms after 6 days of incubation at the DCM (bars represent the standard deviation). Similar letters (a, b or c) attributed to 2 or more treatments indicate no significant differences (p < 0.05) between these treatments.

Figure 9- Abundance of picophytoplankton in surrounding surface waters on day 0 and 6, and in controls and deep water-enriched (2 % and 10 %) microcosms after 6 days of incubation at 45 m depth (a) and 80 m depth (b) (bars represent the standard deviation). Similar letters (a, b or c) attributed to 2 or more treatments indicate no significant differences (p < 0.05) between these treatments.
Figure 1

Figure 2

a) JUNE 16

- Model-Salinity
- OTEC-Salinity
- Model-Temperature
- OTEC-Temperature

b) NOVEMBER 18

- Model-Salinity
- OTEC-Salinity
- Model-Temperature
- OTEC-Temperature
Figure 5

**a)**

- Surrounding waters D0
- Surrounding waters D4
- Surrounding waters D6

**b)**

- Surrounding waters D0
- Surrounding waters D4
- Surrounding waters D6

**c)**

- Surrounding waters D0
- Surrounding waters D4
- Surrounding waters D6

**d)**

- Surrounding waters D0
- Surrounding waters D4
- Surrounding waters D6
Figure 6

Figure 7
Figure 8: Diagnostic pigment concentrations in surrounding surface waters on days 0 and 6, and in controls and deep water-enriched (2% and 10%) microcosms after 6 days of incubation at the DCM (bars represent the standard deviation). Similar letters (a, b or c) attributed to 2 or more treatments indicate no significant differences (p-value < 0.05) between these treatments.

Figure 9: Cell counts (cells mL⁻¹) of various species.