

Anonymous Referee #1

The authors conducted a field experiment to assess how the use of ethylene biosynthesis inhibitor (AVG) would alleviate the inhibitory effects of ethylene from plants due to drought stress to methane oxidation of soil microbes. Their results showed that adding AVG could increase soil methane oxidation rates compared to control, and thus they concluded that AVG application can increase soil methane oxidation process under moisture stressed conditions. I found this manuscript was straightforward and well written. The topic falls within the scope of the journal. But I think this paper is more suitable for a short communication, as the paper is too short and data presented here was very limited. I suggest the authors can modify it to a short communication.

R: Many thanks for your positive comments. We have asked the journal manager to change the type of this manuscript into ‘Technical Note’, a kind of short communication for this journal.

Some minor points are here.

1. L67-70 Rephrase it. Too long to understand.

R: It has been revised. Lines 72-76

2. L111-116 How many gas sampling was conducted to measure CH₄ oxidation rates in one jar? How about the coefficient for linear regression? It’s better to show gas concentration over time for different treatments in this study.

R: We conducted two gas sampling at the beginning and the end of the incubation. Then soil methane oxidation rates in each jar were calculated from differences in the headspace CH₄ concentration over the incubation time. We cannot provide the changes in CH₄ concentration and we cannot calculate the linear regression of soil CH₄ oxidation rates over time, but we acknowledge that these data will make these results more interesting. Lines 126-128

3. L135 “CH₄ methane oxidation”

R: Thanks. It has been changed into ‘CH₄ oxidation’. Line 152

4. L161-164 Can excessive irrigation directly reduce aerobic methane oxidation as the authors proposed in L148-152?

R: Yes, it might be possible. However, it would be expected that the combination of irrigation and AVG would produce CH₄ oxidation rates either the same, or potentially greater than, those observed for AVG alone. In this study, we did not find this case, so the explanation for the significant interaction remains unknown. A further study are needed to investigate the effects of plant, soil and microbial interactions under drought stress. Lines 183-190

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4 **Technical Note**
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Manipulating interactions between plant stress responses and soil methane oxidation rates

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30 **Abstract**

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32 It has recently been hypothesised that ethylene, released into soil by stressed plants,
33 reduces the oxidation of methane by methanotroph. To test this, a field trial was
34 established in which maize plants were grown with and without soil moisture stress,
35 and the effects of addition aminoethoxyvinylglycine (AVG; an ethylene biosynthesis
36 inhibitor), and biochar (increases soil water holding capacity and reduces plant stress)
37 were determined following the static incubation of soil samples. AVG increased
38 methane oxidation rates by 50% ($P=0.039$), but only in the absence of irrigation. No
39 other treatment effects were observed. This result provides evidence for a positive
40 feedback system between plant stress, ethylene production, and impacts on
41 methanotrophic activity.

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43 **Keywords** methane oxidation; plant stress; ethylene inhibition; methanotroph; positive
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59 **1 Introduction**

60 The atmospheric concentration of methane (CH_4) has almost tripled over the past 150
61 years, making a substantial contribution to climate change (Forster et al., 2007).
62 Aerobic soils provide an important habitat for methanotrophic bacteria, and provide the
63 only significant biological sink for atmospheric CH_4 ($20\text{-}45 \text{ Tg CH}_4 \text{ yr}^{-1}$) (Forster et al.,
64 2007). However, CH_4 uptake by these soil ecosystems can be impacted by
65 environmental stress (Kolb, 2009). A common plant physiological response to
66 ecological stress, such as drought, is the production of ethylene (Morgan and Drew,
67 1997). In soils, however, ethylene maybe inhibitory to methanotrophic activity (Jäckel
68 et al., 2004; Pierek et al., 2006; Zhou et al., 2013), and thereby reduce CH_4 oxidation.
69 This potential interaction needs to be understood, as it may constitute an important
70 positive feedback loop between climate disruption, soil ecosystem disturbance, and
71 reduced CH_4 removal from the atmosphere (Bousquet et al., 2006; Zhou et al., 2013).

72 To test our previous hypothesis that drought-induced *in planta* ethylene production
73 reduces soil CH_4 oxidation rates (Zhou et al., 2013), we manipulated plant stress
74 responses by adding the ethylene biosynthesis inhibitor aminoethoxyvinylglycine ([S]-
75 trans-2-amino-4-(2-aminoethoxy)-3-butenoic acid hydrochloride; hereafter AVG)
76 (Boller et al., 1979). In addition, the study tested the hypothesis that addition of biochar
77 to soils may result in increased water holding capacity, reducing drought stress and
78 thereby acting as a potential tool to maintain CH_4 oxidation (Karhu et al., 2011). This
79 is illustrated conceptually in Fig. 1, in which the application of irrigation (IR) and
80 biochar (BC) are able to maintain rates of CH_4 oxidation by reducing moisture stress
81 and therefore ethylene production, whereas AVG prevents the production of ethylene
82 after the plant experiences stress.

83 **2 Material and methods**

84 **2.1 Study site**

85 The study site was located in the Bjelke-Petersen *Research Station* at Kingaroy
86 (26.53° S , 151.83° E) in the South Burnett Region of Queensland, Australia.
87 Precipitation averages 789 mm per annum with erratic summer droughts frequent in the
88 region. Soil at the field trial site soil is an acidic red ferrosol (pH 5.5) with high cation

89 exchange capacity (Isbell, 1993). The site has a long history of cultivation, supporting
90 peanut and maize rotations with winter fallows.

91 **2.2 Experiment design and management**

92 A full factorial, split plot design field trial was established as follows: two IR
93 treatments (IR and no IR) \times two BC treatments (BC at 9.2 t ha $^{-1}$ and no BC) \times two
94 ethylene suppression treatments (AVG and no AVG). Each treatment had five replicates,
95 producing a total of 40 plots. Due to practical concerns regarding application and
96 maintenance, the IR treatments were established in two discrete areas that were spanned
97 by five blocks. A schematic of the trial site is given in the supplementary information
98 (Fig. S1).

99 The BC treatment was established through application of peanut shell BC to the
100 surface of the planting zone (~450 mm wide strip each row) in early 2013. The BC was
101 incorporated into the soil with a rotary hoe to a depth of 200 mm. The chemical
102 properties of the peanut shell BC are provided in the supplementary information (Table
103 S1).

104 The site was machine planted with maize cultivar Pioneer 32p55 (Dupont Pioneer
105 Australia) at a density of approximately 4 plants m $^{-2}$ in late January, 2014. Compound
106 fertiliser (N:P $_2$ O $_5$:K $_2$ O 11.9:14.1:9.9) at 180 kg ha $^{-1}$, and urea at 100 kg ha $^{-1}$, were
107 applied at sowing. Trickle tapes, installed into plots receiving IR, were used to distribute
108 water equivalent to ~50 mm of rainfall whenever there was a continuous dry spell for
109 two weeks throughout the growing season (late January to late June).

110 To reduce the *in planta* production of ethylene, the commercial plant growth
111 regulator ReTain (containing 15% AVG, Valent Bioscience Cooperation, Walnut Creek,
112 CA, USA) was sprayed onto the crop four times from mid-April to mid-June (the peak
113 maize growth window) at intervals of three weeks. During each event, the treated rows
114 of maize received approximately 750 ml of ReTain solution (prepared at the label rate
115 of 1 g ReTain l $^{-1}$ water) directly to the surface of the plants.

116 **2.3 Sample collection and analysis**

117 In late June 2014, six soil cores from 0 - 100 mm depth were collected from the maize
118 rooting zone of each plot using a 30 mm diameter soil auger. All samples were collected

119 from the two middle rows of maize in each plot, and the six soil cores from within each
120 plot bulked to a single plot sample. After sieving to 2 mm, a 50 g (fresh mass)
121 subsample of each sample was set aside for CH₄ oxidation rate measurements and the
122 remaining material dried at 105°C for 48 hours to determine soil moisture content.

123 Soil CH₄ oxidation rates were determined using the laboratory incubation. Briefly,
124 about 20 g soil subsamples were incubated in 1-litre glass jars at ambient atmospheric
125 CH₄ concentration (assumed to be 1.9 ppm) for one week in the dark at 25°C.
126 Headspace gas samples (approximately 30 ml) were collected through a rubber septum
127 in the jar lid at the beginning and the end of the incubation, and concentrations of CH₄
128 determined using GC-FID (GC-2010 Plus, Shimadzu, Japan). The CH₄ oxidation rates
129 in each jar were calculated from differences in the headspace CH₄ concentration over
130 the incubation time (Zhou et al., 2008), and adjusted to soil dry weight. Standards were
131 measured once every 10 samples; the coefficient of variation in CH₄ oxidation rate was
132 less than 5% and control jars had ambient CH₄ concentrations.

133 **2.4 Statistical analysis**

134 Statistical analysis was carried out in R-3.2.3 (Zhou et al., 2017) using a multi-factor
135 ANOVA model incorporating an error structure accounting for the split-plot design
136 associated with the non-random assignment of the IR treatment. The multi-comparison
137 analysis methods provided in the “easyanova 4.0” R package was used to test for
138 treatment interactions.

139 **3 Results**

140 Over the course of the field trial, five dry spells occurred. Irrigation to the IR-plots
141 resulted in delivery of 250 mm more water to this treatment than the controls. This
142 resulted in significantly increased soil moisture ($P<0.001$) in IR soils (18.9%) compared
143 with the non-irrigated soils (15.4%) at sampling time. Neither the AVG or BC
144 treatments had any effect ($P>0.05$) on soil moisture.

145 No significant main effects were observed, but a significant interaction between
146 irrigation and AVG application was detected (Table 1). Exploration of this interaction
147 with multi-comparison analysis determined that CH₄ oxidation rates were increased by
148 50% following AVG application ($P=0.039$), but only in the absence of IR (Fig. 2). The

149 addition of biochar had no effect on CH₄ oxidation rates either as a main or interactive
150 effect.

151 **4 Discussion**

152 The increase in CH₄ oxidation with the AVG treatment either alone or in combination
153 with the BC treatment aligns with past studies assessing the effect of increased ethylene
154 concentrations on soil CH₄ oxidation rates (Jäckel et al., 2004; Xu et al., 2008). This
155 response also supports the hypothesis that *in planta* ethylene production in response to
156 stress decreases the capacity of soil to support methanotrophic activity (Zhou et al.,
157 2013).

158 The lack of effect of BC on CH₄ oxidation is at odds with the results of previous
159 work (e.g. Karhu et al., 2011; Kim et al., 2017). However, BC added in this study had
160 no influence on soil moisture content, and this is proposed to be a key mechanism for
161 BC to support CH₄ oxidation in drought conditions (Karhu et al., 2011). The reason
162 why BC addition did not result in increased soil moisture in this case is unclear, but
163 may be related to differences in BC chemistry or amounts applied as these vary from
164 study to study (e.g. Kim et al., 2017).

165 The significant interaction between the AVG and IR treatments is more difficult to
166 reconcile. The IR treatment was intended to significantly increase soil moisture content
167 compared to the no IR treatment, reducing water stress and likely *in planta* ethylene
168 production. It was noted that increased soil moisture content can directly influence
169 methanotrophic activity, as water-driven increases in microbial activity can enhance
170 methanotroph, whereas water content that exceed field capacity can rapidly decrease
171 CH₄ oxidation rates by reducing gas mobility through soil pores (Le Mer and Roger,
172 2001). Given the initial soil water content and scale of the increase with the IR treatment,
173 direct stimulation of CH₄ oxidation was considered the most likely outcome when
174 considering plant-independent effects. Consequently, it was anticipated that any effect
175 of AVG on CH₄ oxidation (putatively via reductions in ethylene production) would only
176 manifest without IR, as the IR treatment would make the AVG treatment redundant.
177 However, CH₄ oxidation rates in plots treated with the either IR or IR and AVG in
178 combination were not significantly greater than untreated control plots. It is possible

179 that the moisture addition associated with the IR treatment was insufficient to
180 substantially alleviate plant drought stress, driving an increase in ethylene production,
181 which could then account for the numerical difference between the AVG and IR
182 treatments (Fig. 2). The water addition may have also been insufficient to meaningfully
183 directly stimulate methanotroph activity. However, it would be expected that the
184 combination of IR and AVG would produce CH₄ oxidation rates either the same, or
185 potentially greater than, those observed for AVG alone. This was not the case, and the
186 explanation for the significant interaction remains unknown. As discussed above, it is
187 possible for increased soil moisture content to inhibit CH₄ oxidation via decreased
188 porosity and gas diffusion (see Zhou et al., 2014), but given the IR treatment alone did
189 not reduce CH₄ oxidation rates relative to the control, this is not a feasible explanation
190 in this case.

191 The lack of data explicitly describing ethylene release into soil in response to the
192 treatments is a limitation to this trial. However, the quantification of ethylene in soil is
193 not trivial, particularly when conducted over time (i.e. continuous) and was outside the
194 resources available for this study. However, given the findings of this study, and
195 considering treatments were field-based, further investigations of the interactions
196 between AVG, plant stress, and CH₄ oxidation should be conducted. In these studies,
197 consideration should be given to collection and integration of ethylene data, particularly
198 given that this data may help shed light on the nature of any interactions between
199 treatments.

200 Overall, the findings of this study indicate that application of an ethylene
201 biosynthesis inhibitor to plant tissue can cause a measurable increase in the capability
202 of soil to oxidise CH₄ under moisture stressed conditions. This supports the hypothesis
203 that the stress-induced production of ethylene by plants can disrupt the activity of
204 methanotrophs, as well as identifying a potential management pathway to help retain,
205 or even enhance, the methanotrophic capability of soils in productive systems. Given
206 the global importance of a positive feedback between environmental stress, plant
207 ethylene production, and lowered microbial CH₄ oxidation activity, further work in this
208 area is needed. In addition, methods to moderate impacts on the methanotrophic

209 community, such as use of alternative forms or rates of biochar application, require
210 investigation to enable provision of important ecosystem services.

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222 **References**

- 223 Argueso C.T., Hansen M. and Kieber J.J.: Regulation of ethylene biosynthesis, *J Plant Growth
224 Regul.*, 26, 92-105, 2007.
- 225 Boller T., Herner R.C. and Kende H.: Assay for and enzymatic formation of an ethylene precursor,
226 1-aminocyclopropane-1-carboxylic acid, *Planta* 145, 293-303, 1979.
- 227 Bousquet P., Ciais P., Miller J.B., Dlugokencky E.J., Hauglustaine D.A., Prigent C., van der Werf
228 G.R., Peylin P., Brunke E.G., Carouge C., Langenfelds R.L., Lathière J., Papa F., Ramonet
229 M., Schmidt M., Steele L.P., Tyler S.C. and White J.: Contribution of anthropogenic and
230 natural sources to atmospheric methane variability, *Nature*, 443, 439-443, 2006.
- 231 Forster P., Ramaswamy V., Artaxo P., Berntsen T., Betts R., Fahey D.W., Haywood J., Lean J., Lowe
232 D.C., Myhre G., Nganga J., Prinn R., Raga G., Schulz M. and van Dorland R.:Changes in
233 atmospheric constituents and in radiative forcing. In: Solomon S et al. (eds) *Climate Change
234 2007: The Physical Science Basis*. Cambridge University Press, UK, pp 130–234, 2007.
- 235 Ho A., Reim A., Kim S.Y., Meima-Franke M., Termorshuizen A., de Boer W., van der Putten W.H.
236 and Bodelier P.L.E.: Unexpected stimulation of soil methane uptake as emergent property
237 of agricultural soils following bio-based residue application, *Global Change Biol.*, 21, 3864-
238 3879, 2015.

- 239 Isbell R.F.: A classification system for Australian soils (third approximation) CSIRO Australia
240 Division of Soils Technical Report 2/1993, CSIRO Division of Soils and National Landcare
241 Program project, Townsville, 1993.
- 242 Jäckel U., Schnell S. and Conrad R.: Microbial ethylene production and inhibition of
243 methanotrophic activity in a deciduous forest soil, *Soil Biol. Biochem.*, 36, 835-840, 2004.
- 244 Karhu K., Mattila T., Bergström I. and Regina K.: Biochar addition to agricultural soil increased
245 CH₄ uptake and water holding capacity - Results from a short-term pilot field study, *Agric.*
246 *Ecosyst. Environ.*, 140, 309-313, 2011.
- 247 Kim J., Yoo G., Kim D., Ding W. and Kang H.: Combined application of biochar and slow-release
248 fertilizer reduces methane emission but enhances rice yield by different mechanisms, *Appl.*
249 *Soil Ecol.*, 117-118, 57-62, 2017.
- 250 Kolb S.: The quest for atmospheric methane oxidizers in forest soils, *Environ. Microbiol. Rep.*, 1,
251 336–346, 2009.
- 252 Le Mer J. and Roger P.: Production, oxidation, emission and consumption of methane by soils: A
253 review, *Eur. J. Soil Biol.*, 37, 25-50, 2001.
- 254 Morgan P.W. and Drew M.C.: Ethylene and plant responses to stress, *Physiologia Plantarum*, 100,
255 620–630, 1997.
- 256 Pierik R., Tholen D., Poorter H., Visser E.J.W., Voesenek L.A.C.J.: The Janus face of ethylene:
257 growth inhibition and stimulation, *Trends Plant Sci.*, 11, 176–183, 2006.
- 258 Xu X., Yuan B. and Wei J.: Vertical distribution and interaction of ethylene and methane in
259 temperate volcanic forest soils, *Geoderma*, 145, 231-237, 2008.
- 260 Zhou X.Q., Wang Y.F., Huang X.Z., Tian J.Q. and Hao Y.B.: Effect of grazing intensities on the
261 activity and community structure of methane-oxidizing bacteria of grassland soil in Inner
262 Mongolia, *Nutr. Cycl. Agroecosys*, 80, 145-152, 2008.
- 263 Zhou X.Q., Smaill S.J. and Clinton P.W.: Methane oxidation needs less stressed plants, *Trends Plant*
264 *Sci.*, 18, 657-659, 2013.
- 265 Zhou, X., Dong, H., Chen, C., Smaill, S.J., Clinton, P.W.: Ethylene rather than dissolved organic
266 carbon controls methane uptake in upland soils, *Global Change Biol* 20, 2379-2380, 2014.
- 267 Zhou X.Q., Guo Z.Y., Chen C.R. and Jia Z.J.: Soil microbial community structure and diversity are
268 largely influenced by soil pH and nutrient quality in 78-year-old tree plantations,

269 Biogeosciences, 14, 2101-2111, 2017.

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280 **Table 1.** Analysis of treatment effects on methane oxidation rates, accounting for the
 281 split-plot design of the trial.

Block	Df	Sum Sq	Mean Sq	F Value	Pr(>F)
Residuals	4	25.27	6.318		
Block:IR	Df	Sum Sq	Mean Sq	F Value	Pr(>F)
IR	1	0.057	0.057	0.01	0.925
Residuals	4	22.785	5.696		
Block:IR:BC	Df	Sum Sq	Mean Sq	F Value	Pr(>F)
BC	1	0.946	0.946	0.245	0.634
IR:BC	1	0.815	0.815	0.211	0.658
Residuals	8	30.924	3.865		
Block:IR:BC:AVG	Df	Sum Sq	Mean Sq	F Value	Pr(>F)
AVG	1	4.77	4.768	1.796	0.199
IR:AVG	1	17.38	17.384	6.549	0.021
BC:AVG	1	6.19	6.186	2.33	0.146
IR:BC:AVG	1	0.42	0.422	0.159	0.695
Residuals	16	42.47	2.654		

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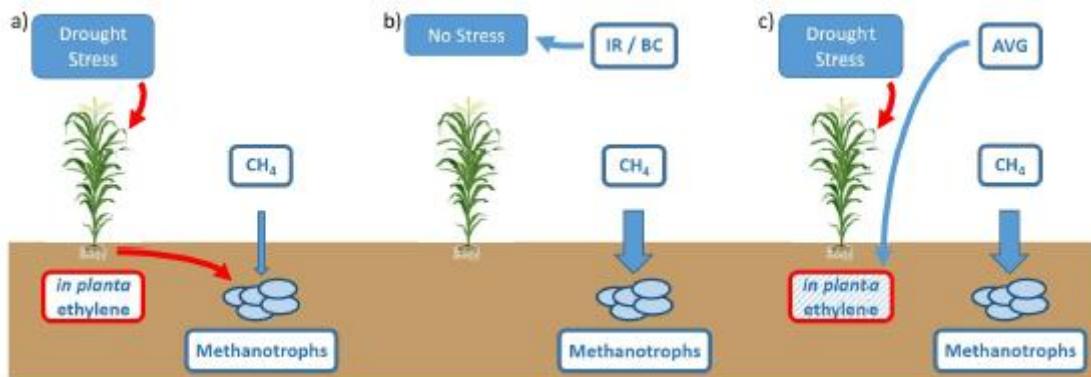
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Fig. 1. Conceptual outline of the proposed relationships between soil CH_4 oxidation rates and aboveground plant biomass with regard to the anticipated effects of the treatments applied in this study. a) Under environmental stress, *in planta* ethylene production is stimulated, resulting in ethylene exudation into the soil atmosphere and the inhibition of soil CH_4 oxidation by methanotrophs. b) The application of irrigation (IR) increases soil moisture while the application of biochar (BC) increases soil moisture holding capacity, both acting to reduce plant stress and prevent ethylene exudation into the soil atmosphere. c) The application of AVG disrupts ethylene production, and limiting or preventing the inhibition of CH_4 oxidation by the stressed plant.

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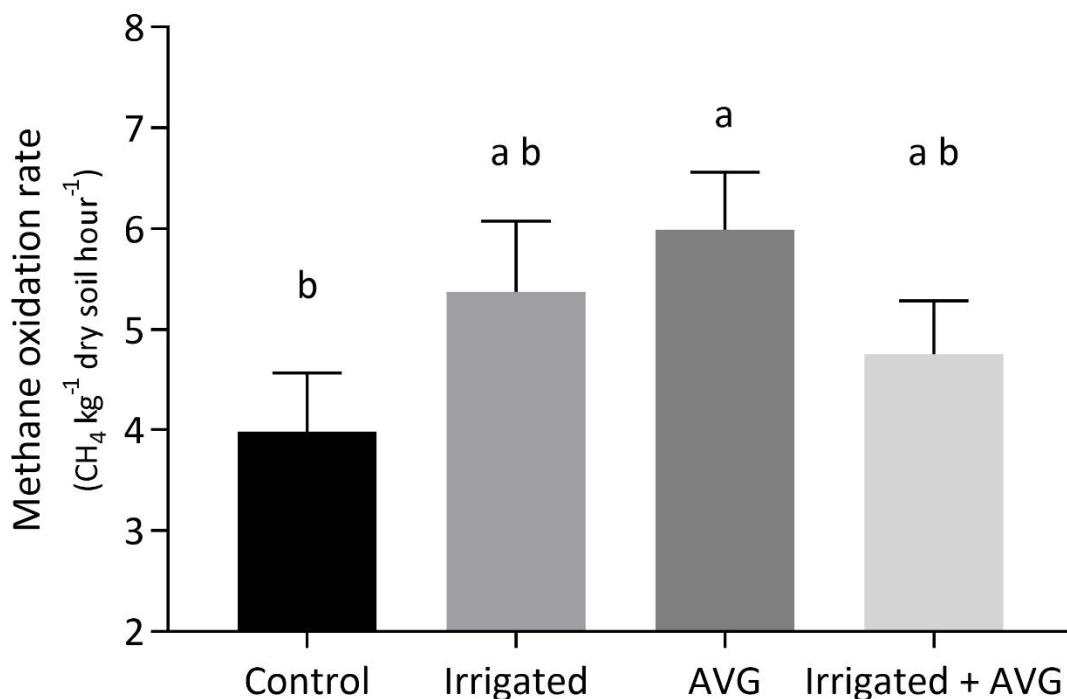
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320 **Fig. 2.** Response of soil CH_4 oxidation rates to treatment with irrigation and AVG under
321 maize plants. Letter groupings indicate significant differences at $P<0.05$; error bars are
322 standard error of the mean. The biochar treatment did not influence results, so the data
323 presented are the means of both biochar and no biochar treatments.

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