Stable isotopes of nitrate reveal different nitrogen processing mechanisms in streams across a land use gradient during wet and dry periods

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Abstract. Understanding the relationship between land use and the dynamics of nitrate (NO₃⁻) is the key to constrain sources of NO₃⁻ export in order to aid effective management of waterways. In this study, isotopic compositions of NO₃⁻ (δ¹⁵N-NO₃⁻ and δ¹⁸O-NO₃⁻) were used to elucidate the effects of land use (agriculture in particular) and rainfall on the major sources and sinks of NO₃⁻ within the Westernport catchment, Victoria, Australia. This study is one of the very few studies carried out in temperate regions with highly stochastic rainfall patterns; enabling a more comprehensive understanding of the applications of NO₃⁻ isotopes in catchment ecosystems with different climatic conditions. Longitudinal samples were collected from five streams with different agriculture land use intensities on five occasions – three during dry periods and two during wet periods. At the catchment scale, we observed significant positive relationships between NO₃⁻ concentrations, δ¹⁵N-NO₃⁻ and percentage agriculture reflecting the dominance of anthropogenic nitrogen inputs within the catchment. Different rainfall conditions appeared to be major controls on the predominance of the sources and transformation processes of NO₃⁻ in our study sites. Artificial fertiliser was the dominant source of NO₃⁻ during the wet periods while nitrified organic matter in sediment and nitrified manure were more apparent sources of NO₃⁻ to the surface water during the dry periods. Denitrification was prevalent during the wet periods while uptake of NO₃⁻ by plants or algae was only observed during the dry periods in two streams. The outcome of this study suggests that effective reduction of NO₃⁻ load to the streams can only be achieved by prioritising management strategies based on different rainfall conditions.

1 Introduction

Anthropogenic sources of NO₃⁻ from catchments can pose substantial risk to the quality of freshwater ecosystems (Vitousek et al. 1997; Galloway et al. 2004; Galloway et al. 2005). Over-enrichment of NO₃⁻ in freshwater systems is a major factor in development of algal blooms which often promote bottom water hypoxia and anoxia. Such anoxia intensifies nutrient recycling and can lead to disruption of ecosystem functioning and ultimately loss of biodiversity (Galloway et al. 2004; Carmago and Alonso 2006). Freshwater streams are often sites for enhanced denitrification (Peterson et al. 2001; Barnes and Raymond...
However, when NO\textsubscript{3} loading from the catchment exceeds the removal and retention capacity of the streams, NO\textsubscript{3} is transported to downstream receiving waters including estuaries and coastal embayments, which are often nitrogen-limited, further compounding the problem of eutrophication.

Understanding the sources, transport and sinks of NO\textsubscript{3} is critical, particularly in planning and setting guidelines for better management of the waterways (Xue et al. 2009). Establishing the link between land use and the biogeochemistry of NO\textsubscript{3} provides fundamental information to help develop NO\textsubscript{3} reduction and watershed restoration strategies (Kaushal et al. 2011). To date, the most promising tool to investigate the sources and sinks of NO\textsubscript{3} are the natural abundance, stable isotopes of NO\textsubscript{3} (expressed as δ\textsuperscript{15}N-NO\textsubscript{3} and δ\textsuperscript{18}O-NO\textsubscript{3} in ‰). Combining the distinctive isotopic signatures that differentiate the various NO\textsubscript{3} sources/end members (e.g. inorganic and organic fertiliser, animal manure, atmospheric deposition, and nitrification) and the predictable fractionation effect when NO\textsubscript{3} undergoes different biological processes (e.g. denitrification and assimilation) have enabled successful application to various environmental settings (Kendall 2007).

In addition to source identification and apportionment, previous studies have utilized the dual isotopic signatures of NO\textsubscript{3} to study the effects of different land uses on the pool of NO\textsubscript{3} in headwater streams (Barnes and Raymond 2010, Sebilo et al. 2003), creeks (Danielescu and MacQuarrie 2013) and large rivers (Voss et al. 2006; Battaglin et al. 2001). Barnes and Raymond (2010) for example found that both δ\textsuperscript{15}N-NO\textsubscript{3} and δ\textsuperscript{18}O-NO\textsubscript{3} varied significantly between urban, agricultural and forested areas in the Connecticut River watershed, USA. Several other investigators (Mueller et al. 2016; Mayer et al. 2002) showed positive relationships between δ\textsuperscript{15}N-NO\textsubscript{3} and the percent of agricultural land in their study area, indicating the applicability of δ\textsuperscript{15}N-NO\textsubscript{3} and δ\textsuperscript{18}O-NO\textsubscript{3} to distinguish NO\textsubscript{3} originating from different land uses. Danielescu and MacQuarrie (2013) and Chang et al. (2002) on the other hand, found no correlations between NO\textsubscript{3} isotopes and land use intensities in the Trout River catchment and the Mississippi River Basin; respectively. These studies attributed the lack of correlation to catchment size (Danielescu and MacQuarrie, 2013) and the homogeneity of land use (Chang et al. 2002). Despite the extensive application of NO\textsubscript{3} isotopes to study the transport of terrestrial NO\textsubscript{3} to the tributaries in the catchment; majority of these studies were carried out in the United States and Western Europe where climatic conditions, for example rainfall patterns are different compared to that in the southern hemisphere. The lack of similar studies in the southern hemisphere (Ohte et al. 2013) impedes a more thorough understanding of NO\textsubscript{3} dynamics within catchment ecosystems.

Most previous studies investigating the relationship between land use and NO\textsubscript{3} export using δ\textsuperscript{15}N-NO\textsubscript{3} and δ\textsuperscript{18}O-NO\textsubscript{3} have either focused on the seasonal or spatial variations in one stream, or used multiple streams with one site per stream (i.e. Mayer et al. 2002; Yevenes et al. 2016). Far fewer studies have incorporated longitudinal sampling of multiple streams over multiple seasons. Nitrate concentrations and concomitant isotopic signatures can change substantially, not only spatially but temporally. Changes in hydrological and physicochemical (notably temperature) conditions of a river can affect the relative contribution of different sources of NO\textsubscript{3} and the seasonal predominance of a specific source (Kaushal et al. 2011; Panno et al. 2008). Denitrification and assimilation of NO\textsubscript{3} by plants and algae have been reported to be more prominent during the dry periods compared to the wet periods (Riha et al. 2014; Kaushal et al. 2011). As such, if spatial and temporal variations of δ\textsuperscript{15}N-NO\textsubscript{3} and δ\textsuperscript{18}O-NO\textsubscript{3} are not considered thoroughly in a sampling regime, it can lead to
misinterpretation of the origin and fate of NO$_3^-$.
Proper consideration of the temporal variability of NO$_3^-$ isotope signatures
and transformation are particularly pertinent in catchments with highly stochastic rainfall patterns, such as Australia.

In this study, we examine both spatial and temporal variations of NO$_3^-$ concentrations and isotopic compositions
within and between 5 streams in 5 catchments spanning an agricultural land-use gradient, enabling us to evaluate (1) the effects
of agriculture land use on the sources and transformation processes of NO$_3^-$ and (2) the effects of rainfall on the predominance
of the sources and fate of NO$_3^-$ in the catchments.

2 Materials and methods

2.1 Study area

This study was undertaken using 5 major streams (Bass River, Lang Lang River, Bunyip River, Watsons Creek and Toomuc
Creek) draining into Western Port (Fig. 1) which lies approximately 75km south east of Melbourne, Australia. Western Port
is a nitrogen-limited coastal embayment (CSIRO, 1996) recognised as a Ramsar site for migratory birds. The catchments in
the Western Port contain three marine national parks, highlighting its environmental and ecological significance. The
catchments cover an area of 3,721 km$^2$ with land uses ranging from semi-pristine/state forest to high density residential and
intense agricultural activities. The area experiences a temperate climate with average annual rainfall ranging from 750mm
along the coast to 1200mm in the northern highlands. Mean monthly rainfall was about 20mm and 53mm in 2014 and 2015,

The catchment overlies a multi-layered combined aquifer system. The main aquifer consists of Quaternary alluvial
and dune deposit (average thickness of <7m) as well as Baxter, Sherwood and Yallock formations (average thickness between
20 and 175m). These aquifers are generally unconfined with radial groundwater flow direction from the basin edge towards
Western Port bay. The hydrogeology of Western Port can be found in Carillo-Rivera, 1975.

Five longitudinal surveys were carried out between April 2014 and May 2015, two during wet periods (14/4/2014;
15/5/2015 - the total rainfall for 5 days before sampling was between 45 and 65mm) and three during dry periods (8/4/2014;
22/5/2014; 21/3/2015 - the total rainfall for 5 to 10 days before sampling was <5mm). A total of 21 sampling sites, indicated
in Fig. 1 were selected across a gradient of catchment land use intensity. The five streams were selected based on the extent
and distribution of land use types between and within each stream sub-catchment, thus enabling comparisons within and
between the streams.

In this study, catchment intensive agriculture was used as predictor of land use intensity in the catchment. These data
were obtained from the National Environmental Stream Attributes database v1.1 (Stein et al. 2014), Bureau of Rural Sciences’
2005/06 Land Use of Australia V4 maps (www.agriculture.gov.au/abares/aclump) and Victorian Resources Online (VRO). In
the context of this study, the catchment intensive agriculture variable is termed as ‘percentage agriculture’. This term represents
the percentage of the catchment subject to intensive animal production, intensive plant production (horticulture and irrigated
cropping) and grazing of modified pastures. This variable also reflects the integrated diffuse sources of nutrients derived from

intense agriculture including animal manure and inorganic fertilisers. The percentage agriculture for the sampling sites ranged between 2 to 96% with the Bass River (94±2%) > Lang Lang (79±5%) > Watsons (76±4%) > Toomuc (71±16%) > Bunyip (upper Bunyip: 12±9%; lower Bunyip: 54±10%; Fig. 2). For the purpose of this study, Bunyip is divided into two sectors (upper and lower Bunyip) based on the proximity of the sampling sites (Fig. 1) and the percentage of land use. All the sampling sites in the upper Bunyip are situated in areas with >30% forestation. In general, the percentage agriculture decreases with increased distance from the Western Port Bay (WPB) for all the streams except Bass River. There is an increase of about 2% in percentage agriculture for Bass River with increased distance from WPB. Watsons Creek has the largest percentage of market gardens (~91%).

2.2 Sample collection and preservation

Water quality parameters (pH, electrical conductivity, turbidity, dissolved oxygen (DO) concentration and water temperature) were measured using a calibrated Horiba U-10 multimeter. Stream samples were collected for the analyses of dissolved inorganic nutrients-DIN (ammonium, NH₄⁺; NO₃ and nitrite, NO₂⁻), dissolved organic carbon (DOC) and NO₃⁻ isotopes (δ¹⁵N-NO₃ and δ¹⁸O-NO₃). These samples were filtered on site using 0.2µm Pall Supor® membrane disc filters. Filtered DOC samples were acidified to pH < 2 with concentrated hydrochloric acid. Samples for δ¹⁸O-H₂O were collected directly from the streams without filtering. Sediment samples were collected from the bottom of the rivers and were kept in zip-lock bags. All samples were stored and transported on ice until they were refrigerated (nutrients samples were frozen) in the laboratory. In addition to stream water and sediment, we also collected samples of artificial/inorganic fertiliser (from the fertiliser distributor in the area) and cow manure (from local farmers).

2.3 DIN and DOC concentration measurements

All chemical analyses were performed within 1-2 weeks of sample collection except for isotope analyses (within 2 months). The concentrations of NO₃⁻, NO₂⁻, and NH₄⁺ were determined spectrophotometrically using a Lachat QuikChem 8000 Flow Injection Analyzer (FIA) following standard procedures (APHA 2005). DOC concentrations were determined using a Shimadzu TOC-5000 Total Organic Carbon analyser. Analysis of standard reference materials indicated the accuracy of the spectrophotometric analyses and the TOC analyser was always within 2% relative error.

2.4 Isotopic analyses

The samples for δ¹⁵N-NO₃ and δ¹⁸O-NO₃ were analysed using the chemical azide method based on the procedure outlined in McIlvin et al. (2005). In brief, NO₃ (NO₃⁻ + NO₂⁻) was quantitatively converted to NO₂⁻ using cadmium reduction and then to N₂O using sodium azide. The initial NO₂⁻ concentrations were insignificant, typically <1% relative to NO₃⁻. Hence, the influence of δ¹⁵N-NO₂⁻ was negligible and the measured δ¹⁵N-N₂O represents the signature of δ¹⁵N-NO₃⁻. The resultant N₂O was then analysed on a Hydra 20-22 continuous flow isotope ratio mass spectrometer (CF-IRMS; Sercon Ltd., UK) interfaced to a cryoprep system (Sercon Ltd., UK). Nitrogen and oxygen isotope ratios are reported in per mil (‰) relative to atmospheric
air (AIR) and Vienna Standard Mean Ocean Water (VSMOW), respectively. The external reproducibility of the isotopic analyses lies within ±0.5‰ for \(\delta^{15}N\) and ±0.3‰ for \(\delta^{18}O\). The international reference materials used were USGS32, USGS 34, USGS 35 and IAEA-NO\(_3^\)-. Lab-internal standards (KNO\(_3^\) and NaNO\(_2^\)) with pre-determined isotopic values were also processed the same way as the samples to check on the efficiency of the analytical method. The \(\delta^{18}O\)-H\(_2\)O values were measured via equilibration with He-CO\(_2\) at 32°C for 24 to 48 hours in a Finnigan MAT Gas Bench and then analysed using CF-IRMS. The \(\delta^{18}O\)-H\(_2\)O values were referenced to internal laboratory standards, which were calibrated using VSMOW and Standard Light Antarctic Precipitation. Measurement of two sets of triplicate samples in every run showed a precision of 0.2‰ for \(\delta^{18}O\)-H\(_2\)O. Sediment samples for the analysis of \(\delta^{15}N\) of total nitrogen were dried at 60°C before being analysed on the 20-22 CF-IRMS coupled to an elemental analyzer (Sercon Ltd. UK).

2.5 Data Analysis

The relationships between percentage agriculture and surface water NO\(_3^-\) concentrations were assessed using linear regression. Percentage agriculture was the predictor variable. NO\(_3^-\) concentration, and \(\delta^{15}N\)-NO\(_3^-\) were response variables. Relationships between \(\delta^{15}N\)-NO\(_3^-\) and NO\(_3^-\) concentration as well as \(\delta^{18}O\)-NO\(_3^-\) and \(\delta^{15}N\)-NO\(_3^-\) were assessed using Pearson’s correlation. The NO\(_3^-\) isotopes response variables were assessed at two spatial scales – individual stream and catchment scale. The catchment scale integrates data from all five studied streams. Any graphical patterns or relationships derived from using these scales represent processes that occur somewhere in the catchment either in the streams or prior to entering the streams with data from the individual stream is likely to represent more localised processes to that particular stream.

3 Results

The streams were oxic throughout the course of our study period with %DO saturation between 70 to 100%. There was no apparent spatial and temporal variation in DO; however, %DO saturation was slightly lower during the dry periods (average of 73±20%) compared to the wet periods (average of 82±12%). Temperature was also relatively consistent with an average of 13±2°C. Ammonium concentration was generally low (<4 µM) except during the wet periods in Bunyip (≈7 µM), Lang Lang (≈21 µM) and Bass (≈29 µM). DOC concentrations were typically 0.8±0.4 mM.

The spatial and temporal variations of NO\(_3^-\) concentration, \(\delta^{15}N\) and \(\delta^{18}O\) across the sites are shown in Fig. 3. NO\(_3^-\) concentrations ranged between 7 µM and 790 µM with averages of 21±15 µM, 50±130 µM, 64±43 µM, 71±43 µM and 190±280 µM for Toomuc, Bunyip, Bass, Lang Lang and Watsons, respectively. The lowest NO\(_3^-\) concentration was observed in the lower Bunyip (4 µM) while the highest NO\(_3^-\) concentration was observed in Watsons Creek (790 µM) at the most downstream site. Nitrate concentrations were generally higher during the wet periods compared to the dry periods in all streams (Fig. 3). During the wet periods, NO\(_3^-\) concentrations typically followed an increasing trend heading downstream except for the Bass River which exhibited the opposite NO\(_3^-\) trend with lower concentrations at downstream sites. During the dry periods, only the Bunyip and Bass Rivers showed apparent longitudinal patterns in NO\(_3^-\) concentrations; with decreasing concentrations...
moving downstream in both. Sites with high percentage agriculture generally also exhibited high NO$_3^-$ concentrations (Fig. 4), particularly during the wet periods.

Overall, isotopic signatures of the riverine NO$_3^-$ spanned a wide range (+4 to +33‰). Approximately 62% of the obtained δ$^{15}$N-NO$_3^-$ values fell below +10‰. More enriched δ$^{15}$N-NO$_3^-$ values (> +10‰) were typically observed during the dry periods and were coincident with a high percentage agriculture (Fig. 4). Among all sites, δ$^{15}$N-NO$_3^-$ values in the Bunyip were relatively depleted (+4 to +12‰), with the lower range found at upper Bunyip (+4 to +8‰). There was no discernible pattern spatially or temporally in δ$^{18}$O-NO$_3^-$, except that higher values were found in Lang Lang and Bass during the wet periods with +4 to +6‰ and +5 to +9‰, respectively compared to the dry periods (<+4‰). For other sampling sites, δ$^{18}$O-NO$_3^-$ ranged between +2 to +13‰. The isotope compositions of sediment, water, artificial fertiliser and cow manure/organic fertiliser are presented in Table 1. The δ$^{15}$N-TN of three potential sources – artificial fertiliser, organic fertiliser and soil organic matter ranged from -0.5 to +0.7‰, +6 to +13‰ and +4 to +5‰, respectively.

4 Discussion

4.1 Potential sources of NO$_3^-$

There are three major potential sources of NO$_3^-$ in the catchments – artificial fertiliser, cow manure/organic fertiliser and soil organic matter (SOM) – see Table 1 for the δ$^{15}$N-TN values. Mineralisation and subsequent nitrification of these end members usually produce NO$_3^-$ which retains (within a few per mil) the N isotope of the δ$^{15}$N-TN (Kendall 2008). The δ$^{18}$O of NO$_3^-$ generated by nitrification of these sources relies on the oxygen isotope of water (δ$^{18}$O-H$_2$O) and dissolved oxygen (δ$^{18}$O-O$_2$) and can be estimated (Eq. 1), Kendall (2007):

\[
\delta^{18}O-NO_3^- = \frac{2}{3} \delta^{18}O-H_2O + \frac{1}{3} \delta^{18}O-O_2
\]  

(1)

Based on this equation, the δ$^{18}$O of NO$_3^-$ produced from nitrification of the potential sources was predicted to be ~ 4.3‰. This value was calculated by using the average δ$^{18}$O-H$_2$O (-5.3‰, assuming nitrification occurred within the stream) and +23.5‰ for δ$^{18}$O-O$_2$. The δ$^{18}$O-O$_2$ value was adopted to reflect that the stream dissolved oxygen concentration was in equilibrium with the atmosphere (Kroopnick and Craig. 1972); which was frequently the case in our study sites. Given that in-river evaporation could have affected the measured δ$^{18}$O-H$_2$O, it was possible that δ$^{18}$O of the nitrified NO$_3^-$ was more depleted than -4.4‰ particularly during the wet period. Based on Casciotti et al. (2002), δ$^{18}$O of nitrified NO$_3^-$ could comprised only 1/6 of the δ$^{18}$O-O$_2$ resulting δ$^{18}$O-NO$_3^-$ of ~ +0.3‰. As such, we considered the δ$^{18}$O of nitrified NO$_3^-$ to range from +0.3 to +4.3‰.

The δ$^{15}$N-TN of cow manure (+6 to +13‰) was most variable compared to other end members. This variation reflects the extent of volatilisation, a highly fractionating process. Volatilisation can cause a fractionation effect of up to 25‰ in the residual NH$_4^+$ (Hubner 1986). As such, the lower value of +6‰ indicates a relatively fresh manure sample and is assumed to represent the initial δ$^{15}$N of the cow manure before undergoing any extensive fractionation.
Atmospheric deposition did not appear to be an important source of NO$_3^-$ in this study based on the relatively depleted $\delta^{18}$O-NO$_3^-$ values (ranged from +2 to +8‰ during the wet periods; +1.5 to +13‰ during the dry periods) of the riverine samples. The $\delta^{18}$O-NO$_3^-$ of atmospheric deposition were reported to range from +60 to +95‰ in the literature (Kendall 2007; Elliott et al. 2007; Pardo et al. 2004). Similarly, groundwater was not considered as an important source of NO$_3^-$ to the streams based on the low NO$_3^-$ concentrations (~0.7 to 7.0µM) reported in previous studies (Water Information System Online; http://data.water.vic.gov.au/monitoring.htm).

4.2 General characteristics of NO$_3^-$ in the streams

Agricultural land use (i.e. market gardens and cattle rearing) influenced NO$_3^-$ concentrations in our study sites. This contention is supported by the clear increasing trend of NO$_3^-$ concentration with increasing percentage agriculture during both dry and wet periods although the relationships between these variables was not strong (Fig. 4). As shown in Fig. 4, high NO$_3^-$ concentrations were particularly observed during the wet periods at sites with more than 70% agricultural land use while during the dry periods, NO$_3^-$ concentrations were consistently lower (NO$_3^-$ < 36 µM). These observations lead to the inference that during the wet periods, in-stream NO$_3^-$ comprised mainly of terrestrially derived NO$_3^-$ entered the streams through surface runoff, while during the dry periods, when surface runoff was negligible, there was minimal introduction of terrestrial NO$_3^-$ into the streams. The availability of in-stream NO$_3^-$ during the dry periods was solely controlled by the NO$_3^-$ transformation processes (i.e. volatilisation, nitrification, denitrification and assimilation) in the streams. Hence, we postulate that the initial source of NO$_3^-$ was the same in all streams during different rainfall events and that artificial fertiliser was the primary initial source of NO$_3^-$.

Artificial fertilisers were more commonly used by the local farmers in the area compared to organic fertilisers (personal communication with the local fertiliser distributors).

The $\delta^{15}$N-NO$_3^-$ signatures of the streams in the Western Port catchment (+4 to +33‰) were generally more variable than found in similar studies (Table 1). In our study, there was a significant positive relationship between $\delta^{15}$N-NO$_3^-$ and percentage agriculture (Fig. 4) indicating that enriched $\delta^{15}$N-NO$_3^-$ in the stream originated from agricultural activities. In fact, the most enriched $\delta^{15}$N-NO$_3^-$ values (>30‰) were observed at the most downstream site of Watsons Creek which has the largest percentage of market gardens (although the total agricultural area is not the highest amongst all the studied sites). Other researchers (e.g. Mayer et al. 2002 and Voss et al. 2006) have also documented similar trends of enriched $\delta^{15}$N-NO$_3^-$ with increasing percentage agriculture. These previous studies, however showed comparatively narrower and more depleted range of $\delta^{15}$N-NO$_3^-$ (Table 2). Given that none of the predicted sources of NO$_3^-$ in the area exhibited an initial $\delta^{15}$N-NO$_3^-$ of more than +6‰, the isotopically-enriched NO$_3^-$ values observed in this study were consequences of a series of transformations processes. This substantiates the earlier speculation on transformations based on the NO$_3^-$ concentration data alone. Hence, we propose the following factors to explain the heavy isotopes observed in our study:

(1) During the wet periods when residence time of the water column was low, there was limited in-stream processing of NO$_3^-$.

The heavy $\delta^{15}$N-NO$_3^-$ values reflect the occurrence of mineralisation, nitrification and subsequent preferential
denitrification of the isotopically lighter NO$_3^-$ source/s in either the waterlogged soil or in the soil zone underneath the market gardens before transport to the streams through surface runoff.

(2) During the dry periods when residence time of the water column was high, in-stream processing of NO$_3^-$ was more apparent than during the wet periods. In addition to mineralisation and nitrification, volatilisation and assimilation by plant and algae was highly likely to occur in the stream further fractionating the isotopic signature of NO$_3^-$.

These processes are conceptualised in Fig. 5 and are corroborated in the following discussion using two graphical methods: the Keeling plot and the isotope biplot. In an agricultural watershed, the co-existence of multiple sources and transformation processes can potentially complicate the use of NO$_3^-$ isotopes as tracers of its origin. Keeling plots ($\delta^{15}$N-NO$_3^-$ versus $1/[\text{NO}_3^-]$) are generally very useful to distinguish between mixing and fractionation (i.e. assimilation and bacterial denitrification) processes (Kendall et al. 1998). The latter typically results in progressively increasing $\delta^{15}$N-NO$_3^-$ values as NO$_3^-$ concentrations decrease and yields a curved Keeling plot. Meanwhile, mixing of NO$_3^-$ from two or more sources can result in concomitant increase of both $\delta^{15}$N-NO$_3^-$ and NO$_3^-$ concentrations and results in a straight line on the Keeling plot (Kendall et al. 1998). A biplot ($\delta^{18}$O-NO$_3^-$ versus $\delta^{15}$N-NO$_3^-$) on the other hand, is an effective method to differentiate between assimilation and denitrification. Nitrate assimilation creates a 1:1 slope on a biplot while the simultaneous increase of $\delta^{18}$O-NO$_3^-$ and $\delta^{15}$N-NO$_3^-$ in a 2:1 pattern indicates the presence of denitrification (Fry 2006).

### 4.3 Key controlling processes of nitrate during the wet periods

Careful examination of the Keeling plots for individual streams (Fig. 6) revealed that during the wet periods, NO$_3^-$ concentrations were significantly and linearly correlated with $1/[\text{NO}_3^-]$ in all the streams. These relationships strongly suggest mixing between two sources (with distinctive isotopic signatures) rather than in-stream processing of NO$_3^-$ was the dominant process regulating the isotopic composition of the residual NO$_3^-$ in the streams during the wet periods. Further evidence to suggest mixing rather than in-stream processing comes from the lack of relationships between $\delta^{18}$O-NO$_3^-$ and [NO$_3^-$] as well as between $\delta^{18}$O-NO$_3^-$ and $\delta^{15}$N-NO$_3^-$ for the individual streams (Fig. 3). If denitrification was dominant, both $\delta^{15}$N-NO$_3^-$ and $\delta^{18}$O-NO$_3^-$ values are expected to increase at low NO$_3^-$ concentration and there would be systematic increase of both N and O isotopes of NO$_3^-$ (Fry 2008). In addition, high DO in the water column ruled out the possibility of pelagic denitrification.

The different trends in the Keeling plots (Fig. 6) for individual streams indicate that the isotopic signature of the dominant NO$_3^-$ source varied temporally and spatially across the catchments. Negative trends on the Keeling plots for Bunyip, Lang Lang and Toomuc (Fig. 6) clearly show that the dominant NO$_3^-$ source was isotopically enriched (above +10‰ for Bunyip and Toomuc and +14‰ for Lang Lang) while the positive trends on the Keeling plots for Bass and Watsons show that the dominant NO$_3^-$ source was more isotopically depleted (less than +8‰ for Bass and less than +9‰ for Watsons). Nevertheless, the isotopic signatures of the dominant source; indicated by the y-intercepts of the Keeling plots were a lot more enriched than the initial $\delta^{15}$N-NO$_3^-$ of all three pre-identified NO$_3^-$ end members. Interestingly, these $\delta^{15}$N-NO$_3^-$ values increased with percentage agriculture except for Bass (Fig. 7). The fact that there was a clear fractionation pattern (~2:1) when integrating the isotope values of all the streams (catchment scale) suggests that denitrification was still prevalent during the
wet periods (Fig. 8a) but this process was more likely to occur prior to NO$_3^-$ entering the streams via surface runoff. We explain these observations on the basis that increased rainfall created a ‘hot moment’ in the soil whereby organic matter mineralisation and nitrification were stimulated followed by denitrification within the waterlogged soil. Waterlogging can result in root anoxia and increased denitrification; leading to significant isotopic enrichment of the residual NO$_3^-$ (Chien et al. 1977, Billy et al. 2010) which was then washed into the streams. The extent of this process (mineralisation – nitrification – denitrification) was greatest at Bass and Watsons; sites with the highest agricultural activity (Fig. 8a). Based on Fig. 8a, the isotope enrichments of the riverine NO$_3^-$ followed the denitrification trend of the artificial fertiliser and the NO$_3^-$ isotopes were distributed in between the denitrification ranges of both artificial fertiliser and SOM suggesting the important contribution of these two sources during the wet periods.

4.4 Key controlling processes of nitrate during the dry periods

Unlike the wet periods, only NO$_3^-$ in the Bass River showed an apparent relationship with $\delta^{15}$N-NO$_3^-$ (Fig. 6) during the dry periods. There was no obvious relationships between $\delta^{15}$N-NO$_3^-$ and 1/[NO$_3^-$] for all other systems during the dry periods limiting the interpretation available from the Keeling plots. This also suggests that mixing between two end members alone is inadequate to explain the variability of $\delta^{15}$N-NO$_3^-$ during the dry periods. In general, during the dry periods, none of the samples show a noticeable pattern of denitrification on a biplot of $\delta^{18}$O vs. $\delta^{15}$N (Fig. 8b). The isotopic composition of the riverine NO$_3^-$ appeared to be clustered into three groups (A, B and C in Fig 8b):

1. NO$_3^-$ in group A showed consistent $\delta^{18}$O but variable $\delta^{15}$N. This is demonstrated by the Lang Lang and Bass; coincident with the highest percentage of agriculture. The consistent $\delta^{18}$O ($\delta^{18}$O of ~2.5‰) shows the importance of nitrification ($\delta^{18}$O of ~0.3 to 4.3‰) and at the same time ruled out the occurrence of denitrification and assimilation. In the absence of the removal processes, the heavy and variable $\delta^{15}$N-NO$_3^-$ values (+6‰ to +20‰) imply that animal manure was an apparent source of NO$_3^-$ during the dry periods for Lang Lang and Bass. This is because volatilization of $^{14}$N ammonia from the animal manure over time can lead to enrichment of $^{15}$N in the residual NH$_4^+$ to > +20‰ (Batman and Kelly 2007) which can subsequently nitrified to produce isotopically-enriched NO$_3^-$ without affecting its $\delta^{18}$O-NO$_3^-$. Tight coupling between mineralisation and nitrification results in NO$_3^-$ retaining the isotopic signature of the residual NH$_4^+$ (Deutsch et al. 2009) in the manure. Hence, it is not surprising that $\delta^{15}$N-NO$_3^-$ > +13‰ in the group A dataset is indicative of nitrified ‘aged’ animal manure. Because of the huge variability in the fractionation effect of ammonia volatilisation, it is difficult to affix an average $\delta^{15}$N value to represent the isotopic signature of this end member. As such, apportioning the relative contribution of nitrified manure versus other sources (nitrified organic matter in the sediment and inorganic fertiliser) is not possible.

2. NO$_3^-$ in group B has variable $\delta^{15}$N and $\delta^{18}$O values as shown by Bunyip and Toomuc which are attributable to isotopic fractionation during plant and/or algae uptake of NO$_3^-$ This is substantiated by the increase of $\delta^{18}$O and $\delta^{15}$N in a ~1:1 pattern (Fig. 9). Close convergence of these linear relationships onto the theoretical assimilation trend
of the nitrified SOM as well as the deviation of the more enriched NO$_3^-$ isotopes towards the assimilation trend of
the nitrified manure reiterate the importance of assimilation and the contributions of both nitrified manure and SOM
as dominant sources of the riverine NO$_3^-$ during the dry periods.

3) NO$_3^-$ in group C comprised the most enriched $\delta^{15}$N and $\delta^{18}$O in the entire dataset (Fig. 8). These isotope values were
observed in Watsons Creek which has the highest percentage of market gardens. These samples were collected when
the creek was not flowing, hence the enriched $\delta^{15}$N and $\delta^{18}$O values could be indications of repeated cycles of
recycling processes (i.e. volatilisation, nitrification, denitrification and assimilation) in the same pool which
enriched the N isotope but had slight effects on the O isotope of NO$_3^-$. Although the isotope values during the dry periods appeared to be more likely controlled by manure and SOM, the contribution
from artificial fertiliser cannot be excluded. As mentioned in the preceding text, most of the fertiliser-derived NO$_3^-$ was
denitrified in the catchment during the wet periods creating an artefact of heavy NO$_3^-$ isotopes in the streams. This NO$_3^-$ could exhibit a similar enriched isotopic composition as the volatilised manure. Overlapping of these isotopic values made it difficult to distinguish between the two sources – a disadvantage of using NO$_3^-$ isotopes in a system where multiple sources and transformation processes coexist. One piece of compelling evidence to show that contribution of artificial fertiliser was also prominent during the dry periods is the deviation of the group B data towards the theoretical assimilation ranges of artificial fertiliser (Fig. 8b). However, calculating the relative contribution of the fertiliser end member is not possible.

5 Conclusions

This study highlights the effect of rainfall conditions on the predominance of sources and transformation processes of NO$_3^-$ on
both individual stream and catchment scale. The significant positive relationships between percentage agriculture and NO$_3^-$
concentrations as well as $\delta^{15}$N-NO$_3^-$ showed that enriched NO$_3^-$ concentrations and $\delta^{15}$N-NO$_3^-$ values resulted from agricultural
activities. The dual isotopic compositions of NO$_3^-$ revealed that both mixing of diffuse sources and biogeochemical attenuation controlled the fate of NO$_3^-$ in the streams of the Western Port catchments. During the wet periods, inorganic fertiliser appeared to be the primary source of NO$_3^-$ to the streams while volatilised animal manure and SOM were the dominant sources of NO$_3^-$ during the dry periods. Denitrification in the catchment appeared to be the more active removal process during the wet periods
in contrast to a greater importance of in-stream assimilation during the dry periods. However, these removal processes were insufficient to remove the agricultural-derived NO$_3^-$ inferring that the streams were unreactive conduits of NO$_3^-$ which might pose a potential NO$_3^-$ enrichment threat to downstream ecosystems. To the best of our knowledge, this is the first study in
Australia and also one of the very few targeted studies in the southern hemisphere investigating the origin and sink of NO$_3^-$ on
a catchment scale using both $\delta^{15}$N and $\delta^{18}$O of NO$_3^-$. The application of NO$_3^-$ isotopes in a region with highly variable and
unpredictable rainfall patterns such as the Western Port catchments although challenging; is imperative particularly in setting
guidelines for sustainable land use management actions.
References


Table 1: The isotopic compositions of potential sources of NO$_3^-$ in the catchment

<table>
<thead>
<tr>
<th>Sample</th>
<th>$\delta^{15}$N-TN (‰)</th>
<th>$\delta^{18}$O-H$_2$O (‰)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Artificial/inorganic fertiliser</td>
<td>-0.5 to +0.7</td>
<td>-</td>
</tr>
<tr>
<td>Cow manure/organic fertiliser</td>
<td>+6 to +13</td>
<td>-</td>
</tr>
<tr>
<td>Sediment (SOM)</td>
<td>+4 to +5</td>
<td>-</td>
</tr>
<tr>
<td>Stream water</td>
<td>-</td>
<td>-5.6 to -4.9</td>
</tr>
</tbody>
</table>
Table 2: Comparison of NO$_3^-$ concentrations and isotopes across different systems reported in the literature

<table>
<thead>
<tr>
<th>Study area</th>
<th>Percentage agriculture (%)</th>
<th>[NO$_3^-$] (µM)</th>
<th>$\delta^{15}$N-NO$_3^-$ (%)</th>
<th>$\delta^{18}$O-NO$_3^-$ (%)</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mississippi River Basin, USA</td>
<td>0 to 100</td>
<td>3.6 to 1290</td>
<td>-1.4 to +12.3</td>
<td>+3.1 to +43.3</td>
<td>Chang et al. 2002</td>
</tr>
<tr>
<td>Connecticut River Watershed, USA</td>
<td>0.8 to 52</td>
<td>0 to 360</td>
<td>*0 to +14.5</td>
<td>*-2 to +14</td>
<td>Barnes et al. 2010</td>
</tr>
<tr>
<td>New York, USA</td>
<td>0 to 72</td>
<td>*5 to 640</td>
<td>*0 to +9</td>
<td>*-8 to +40</td>
<td>Burns et al. 2009</td>
</tr>
<tr>
<td>Mid-Atlantic and New England states of the USA</td>
<td>2 to 38</td>
<td>7.9 to 184</td>
<td>+3.6 to +8.4</td>
<td>+11.7 to +18.5</td>
<td>Mayer et al. 2002</td>
</tr>
<tr>
<td>Baltic Sea catchment</td>
<td>1 to 81</td>
<td>3 to 216</td>
<td>-1.5 to +14</td>
<td>+10 to +25</td>
<td>Voss et al 2006</td>
</tr>
<tr>
<td>Trout River catchment, Atlantic Canada</td>
<td>~39.7</td>
<td>32 to 170</td>
<td>+2.13 to +6.35</td>
<td>+1.51 to +7.07</td>
<td>Danielescu and MacQuarrie 2013</td>
</tr>
<tr>
<td>Skuterud catchment, Norway</td>
<td>0 to 100</td>
<td>21 to 1850</td>
<td>+3 to +18</td>
<td>+10 to +24</td>
<td>Kaste et al. 2006</td>
</tr>
<tr>
<td>Mordre catchment, Norway</td>
<td>74 to 100</td>
<td>120 to 2320</td>
<td>+8 to +15</td>
<td>+5 to +20</td>
<td>Kaste et al. 2006</td>
</tr>
<tr>
<td>Pearl river drainage basin</td>
<td>~86</td>
<td>41 to 110</td>
<td>+1.9 to +17.6</td>
<td>+5.6 to +17.3</td>
<td>Chen et al. 2009</td>
</tr>
<tr>
<td>Westernport catchment, Australia</td>
<td>2 to 96</td>
<td>4 to 790</td>
<td>+5.7 to +33</td>
<td>+1.4 to +12.7</td>
<td>This study</td>
</tr>
</tbody>
</table>

*Values estimated from presented figures, might not accurately represent the actual data
Figure 1: Map of Western Port Bay (WPB) in southern Victoria, Australia and major rivers discharging into WPB. Closed circles represent sampling sites where surface water samples were obtained.
Figure 2: The percent agriculture for each of the sampling sites.
Figure 3: Spatial and temporal variations of nitrate concentrations and isotopes values. Closed circles represent data obtained during the wet periods. Open circles represent data obtained during the dry periods.
Figure 4: Relationship between (a) NO$_3^-$ concentration; (b) $\delta^{15}$N-NO$_3^-$ and the percentage of agricultural land use.
Figure 5: Conceptual diagram illustrating the sources and processes of NO$_3^-$ during the wet and dry periods in the Western Port catchment. The values of enrichment factor ($\varepsilon$) were obtained from the literature to indicate the relative contribution of the transformation processes to the isotopic compositions of the residual NO$_3^-$. 
Figure 6: Relationship between $\delta^{15}$N-NO$_3$ and $1/[\text{NO}_3^-]$ for individual streams during the wet and dry periods.
Figure 7: Relationship between $\delta^{15}$N-NO$_3$- and percentage agriculture during the wet periods. Data for Bass-dry period was also presented because only the Keeling plot for Bass-dry period indicates mixing between different sources. The shaded area represents the $\delta^{15}$N-TN of the potential end members.
Figure 8: Biplot of δ¹⁵N-NO₃⁻ versus δ¹⁸O-NO₃⁻ for (a) wet and (b) dry periods. Blue shaded area represents possible isotopic compositions of denitrified NO₃⁻ originated from SOM (δ¹⁵N: +4.5‰). Grey shaded area represents the possible isotopic composition of denitrified NO₃⁻ originated from inorganic fertiliser (δ¹⁵N-NO₃⁻: +0.1‰). The δ¹⁸O-NO₃⁻ used were +0.3‰ and +4.4‰ representing the minimum and maximum estimates of δ¹⁸O of nitrified NO₃⁻, respectively. The shaded area were plotted based on the theoretical 2:1 denitrification relationship between δ¹⁵N-NO₃⁻ and δ¹⁸O-NO₃⁻ (Kendall 2007).
Figure 9: Biplot of $\delta^{15}$N-NO$_3$ versus $\delta^{18}$O-NO$_3$ for Bunyip and Toomuc (group B data in Fig. 8b). Shaded areas represent theoretical assimilation trends for cow manure, SOM and inorganic fertiliser. Solid lines represent the assimilation trends for Bunyip (both lower and upper Bunyip) and Toomuc.