

***Interactive comment on “Phytoplankton growth responses to Asian dust additions in the Northwest Pacific Ocean versus the Yellow Sea” by Chao Zhang et al.***

**Chao Zhang et al.**

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**Referee #1**

*General comments:*

*This study describes the results of several experiments in which surface plankton communities from the Yellow Sea and the NW Pacific ocean were amended with atmospheric dust and different nutrients added alone and in various combinations. The responses studied included the numerical abundance of diatoms and dinoflagellates, size-fractionated chl *a* concentration, and nutrient concentration. The main strength of the study is that parallel incubations, in which inorganic nutrients were added in different combinations, allowed the authors to gain insight into the causative mechanisms underlying the phytoplankton responses to dust. However, in some cases (as in the case of *P* availability, see below) the authors seem to over-interpret the available evidence. A limitation of the study is that only standing stocks were examined; no metabolic rate measurements were included, and therefore it is not possible to ascertain the dominant type of nutrient limitation (Blackman versus Liebig). The authors rely heavily on the use of chl *a* as a proxy for phytoplankton biomass. However, variability in *C:Chl a* ratios should be taken into account. The extent to which the simulated process of atmospherical transformation of dust yields materials that are realistic in terms of nutrient content and solubility should be discussed. Finally, some sections of the Discussion are speculative and based on tenuous assumptions. All these limitations should be addressed before publication is recommended in Biogeosciences. Some suggestions as to data presentation and analysis are also given below.*

**Response:**

We would like to thank the referee very much for the valuable comments which enabled us to improve the quality of the manuscript. We will revise the manuscript accordingly to address the comments.

Regarding concern over nutrient limitation status: The standing stocks such as Chl *a* and biomass have been widely used for ascertaining the dominant type of nutrient limitation in the ocean. However, indeed, we acknowledge that

metabolic rate has a better representation than standing stocks in ascertaining nutrient limitation. We will add metabolic rate measurements for future incubation experiments.

Regarding C;Chl *a* ratios, we will illustrate that our results are based on Chl *a* and cell abundance rather than biomass, and also consider the uncertainty in the new manuscript (refer to Q4).

We will provide additional descriptions about the simulating processes in Text S1 (refer to Q1).

Regarding concerns over P, we have made a detailed response to Q5.

### *Specific comments*

*1. Simulation of atmospheric transformation of dust. How do these AM-dust materials compare, in terms of nutrient composition and solubility, with real dust samples collected in situ? This is critical to assess if the results observed are representative of real responses at sea. Table in supp. info. shows that N concentration is increased 4 orders of magnitude relative to N concentration in collected rain. Does this mean that the potential for nutrient supply is grossly overestimated in these artificially treated materials?*

Response:

During a strong Asian dust event, the loading of inorganic nitrogen ( $\text{NO}_3^- + \text{NH}_4^+$ ), soluble P and Fe in atmospheric particles collected in the Yellow Sea was around  $714 \mu\text{mol g}^{-1}$ ,  $4.3 \mu\text{mol g}^{-1}$  and  $550 \mu\text{g g}^{-1}$ , respectively (Shi et al., 2012). The values in AM-dust used in this study are  $577 \mu\text{mol g}^{-1}$ ,  $4.3 \mu\text{mol g}^{-1}$  and  $473 \mu\text{g g}^{-1}$ , respectively (Table 3 in the origin version), which were highly comparable to those observed by Shi et al. (2012). The authors, however, agree that the loadings of these components in atmospheric dust particles could highly vary in different cases. The weakness will be added at the end of the conclusion.

In this study, the aging process of dust followed Guieu's (2010) method and aimed at stimulating the cloud reaction between dust and synthetic evaporating cloud water. The pH around dust in the cloud process (i.e. mix with evaporating cloud water) was found to be as low as  $\sim 1$  during their transport to the Yellow Sea (Meskhidze et al., 2003), whereas the typical pH in rainwater is 5 (Watanabe et al. 2001, Sasakawa and Uematsu, 2002, Wang et al. 2002, Sakihama et al. 2008, Zhang et al. 2011), meaning that a dilution by a factor  $10e^4$ . In consequent, in order to reproduce an evaporating cloud, we have used a concentration 10 000 in our experiments in comparison to the typical concentrations found in rainwater. Considering the typical concentrations of dust in rainwaters was  $10 \text{ mg L}^{-1}$  (Ridame et al., 2002), the dust loading in evaporating cloud water could reach  $100 \text{ g L}^{-1}$ . As a consequence, all of the concentrations in evaporating cloud water were around 10000-fold larger (i.e. 4 orders of magnitude larger) than those in natural rainwater. Table S1 summarized the primary chemical composition of rains in the Eastern Asian regions and the evaporating cloud water used for our simulation.

Shi, J., Gao, H., Zhang, J., Tan, S., Ren, J., Liu, C., Liu, Y., and Yao, X.: Examination of causative link between a spring bloom and dry/wet deposition of Asian dust in the Yellow Sea, China, *Journal of Geophysical Research: Atmospheres*, 117, 2012, doi:10.1029/2012JD017983.

Guieu, C., Dulac, F., Desboeufs, K., Wagener, T., Pulido-Villena, E., Grisoni, J.-M., Louis, F., Ridame, C., Blain, S., and Brunet, C.: Large clean mesocosms and simulated dust deposition: a new methodology to investigate responses of marine oligotrophic ecosystems to atmospheric inputs, *Biogeosciences*, 7, 2765-2784, doi: 10.5194/bg-7-2765-2010, 2010.

Meskhidze, N., Chameides, W. L., Nenes, A., and Chen, G.: Iron mobilization in mineral dust: Can anthropogenic SO<sub>2</sub> emissions affect ocean productivity?. *Geophysical Research Letters*, 30(21), 2003, doi: 10.1029/2003GL018035.

Watanabe, K., Ishizaka, Y., & Takenaka, C.: Chemical characteristics of cloud water over the Japan Sea and the Northwestern Pacific Ocean near the central part of Japan: airborne measurements. *Atmospheric Environment*, 35(4), 645-655, 2001, doi: 10.1016/S1352-2310(00)00358-7.

Sasakawa, M., & Uematsu, M.: Chemical composition of aerosol, sea fog, and rainwater in the marine boundary layer of the northwestern North Pacific and its marginal seas. *Journal of Geophysical Research: Atmospheres*, 107(D24), 2002, doi: 10.1029/2001JD001004.

Wang, Z., Akimoto, H., & Uno, I.: Neutralization of soil aerosol and its impact on the distribution of acid rain over east Asia: Observations and model results. *Journal of Geophysical Research: Atmospheres*, 107(D19), 2002, doi: 10.1029/2001JD001040.

Sakihama, H., Ishiki, M., & Tokuyama, A.: Chemical characteristics of precipitation in Okinawa Island, Japan. *Atmospheric Environment*, 42(10), 2320-2335, 2008, doi: 10.1016/j.atmosenv.2007.12.026.

Zhang, J., Zhang, G. S., Bi, Y. F., & Liu, S. M.: Nitrogen species in rainwater and aerosols of the Yellow and East China seas: Effects of the East Asian monsoon and anthropogenic emissions and relevance for the NW Pacific Ocean. *Global Biogeochemical Cycles*, 25(3), 2011, doi: 10.1029/2010GB003896.

Ridame, C., & Guieu, C.: Saharan input of phosphate to the oligotrophic water of the open western Mediterranean Sea. *Limnology and Oceanography*, 47(3), 856-869, 2002, doi: 10.4319/lo.2002.47.3.0856.

2. *Section 3.2. This section should present first the changes in nutrient concentration, and then those of chl a concentration. In both cases, the actual increases (absolute values) should be described (e.g. nutrient or chl a concentration increased by xx umol/L or ug/L), rather than just the relative increases (xx-fold). It is important to describe the chl a responses in terms of absolute value of increase, so that they can be compared with the amount of nutrient released from the dust or provided by the nutrient amendments.*

Response:

Agree. We will revise accordingly through the manuscript.

3. *Conversion efficiency index. This index should be described in the Methods section.*

Response:

Agree. It will be moved to the Method.

*4. It is unclear why the index is formulated in this way. Why not use just final minus initial chl<sub>a</sub> concentration, as is done for N? It does not make sense to sum consecutive differences over time in chl<sub>a</sub> concentration between treatments and control. In addition, the index has a potential flaw, because C:Chl<sub>a</sub> ratios are bound to be different in the different sites (due, for instance, to differences in nutrient and/or light availability). So the same response in terms of % increase in biomass (carbon) will yield higher chl<sub>a</sub> concentration (in absolute values), and thus higher conversion efficiency, in waters with low phytoplankton C:Chl<sub>a</sub> values. The limitations of using Chl<sub>a</sub> as a proxy for biomass should be acknowledged and discussed. Finally, when reporting the values of this index in the text, its units should be indicated.*

Response:

In the revision, we will clarify the net conversion efficiency index (NCEI) proposed in this study to be an approximate estimation for the utilization of N for the growth of phytoplankton. Therefore, the capacity to synthesize Chl *a* per unit concentration of nitrogen (N) in different treatments can be compared. We agree that the sum consecutive differences over time in Chl *a* concentration between treatments and control will lead to an overestimation of the real net conversion efficiency because of the accumulation effect. Theoretically, the use of the maximum difference will lead to an underestimation of the real net conversion efficiency because of degradation of Chl *a* in the growth period. The real net conversion efficiency should be between those calculated by the two approaches.

We agree that C:Chl *a* ratios can be different at the different sites. However, most of the calculated indexes at different sites in this study were highly consistent. This implied that C:Chl *a* ratios at these site in this study might narrowly vary. However, it should be cautious for extrapolating the current findings to other sites. This will be clarified in the revision.

The unit of NCEI in the text will be added in the revision.

*5. Section 4.3. This section is speculative and difficult to follow. It is unclear how the ‘increase in bioavailable P concentration following AM-dust addition’ has been identified. The relationship between N:P ratios in supply vs demand is tentative at best, since actual supply N:P ratios were not measured. The paragraph on lines 395-406 starts with an untenable assumption, namely that ‘C\_N:P in AM-dust treatments is equal to that in N treatments’. To the extent that dust additions and N additions create distinct nutrient environments, it is most unlikely that consumption N:P ratios will be the same. In fact, the previous paragraph has argued that consumption N:P ratio is lower in dust treatments than in N treatments. Thus the subsequent calculations and conclusions have no use. This sub-section (l. 395-406) should be deleted. In the subsequent paragraph, the basis for the need for additional P supply is unclear.*

Response:

We have determined the chemical contents of AM-dust in the laboratory. The details can be seen in Methods Section 2.4. The content of bioavailable nutrients from AM-dust has been listed in Table 3. As the added concentration of AM-dust was  $2 \text{ mg L}^{-1}$ , thus, we can get the theoretical amounts of N and P nutrients supplied by AM-dust in the incubation system. In the field incubation experiments, the time interval of adding materials (e.g. AM-dust and various nutrients) into the incubation bottles and nutrient measurement (i.e. measurement on day 0) was around 1-2h. During this period, microbial uptake, scavenging by cell surface and bottle wall are all possible to influence the measurement of added N and P nutrients from AM-dust, which has been proved by the previous incubation experiments (Liu et al., 2013). Therefore, the calculated theoretical amounts of N and P nutrients supplied by AM-dust in the incubation system is close to the actual values relative to those measured directly on day 0.

We will correct the inappropriate expression of ' $C_{N:P}$  in AM-dust treatments is equal to that in N treatments' and change the equation (2) into inequation (2).

The last paragraph in the Discussion Section 4.3 is particularly important because it provides a new insight to analyze the budget of bioavailable P in the AM-dust addition incubation experiments. Unfortunately, the original presentation seemed to be unclear and didn't service the target well. We will rewrite this paragraph to make our thoughts understandable.

Liu, Y., Zhang, T., Shi, J., Gao, H., and Yao, X.: Responses of chlorophyll a to added nutrients, Asian dust, and rainwater in an oligotrophic zone of the Yellow Sea: Implications for promotion and inhibition effects in an incubation experiment, *Journal of Geophysical Research: Biogeosciences*, 118, 1763-1772, doi: 10.1002/2013JG002329, 2013.

#### Minor comments

1. lines 53-54: Rewrite sentence: '*The N:P ratio of dust deposition is much higher than the Redfield ratio (N:P=16)...*'

Response:

Corrected.

2. line 110. Were attenuation filters used? PAR levels should be given.

Response:

We did not use attenuation filters in this study. The surface seawater (2-5 m) was collected and incubated under natural light.

3. line 139. The ultrasonic method should be described briefly. The use of ultrasounds maximises the extraction of

*nutrients but it probably overestimates the amount of nutrients that is actually released in real conditions at sea.*

Response:

We will make a brief description of the ultrasonic method in the text.

In general, more than 30 minute of ultra-sonication treatment will increase the temperature and may enhance the leaching of nutrients and thus overestimate. In our study, the time duration used for ultra-sonication is 30 minute. We used the ice pack to keep the temperature of water bath stable at  $\sim 0^{\circ}\text{C}$ . Please also see Specific Comments No. 3 for Referee #3.

4. line 141. *Re-write sentence, '...and filtrates were stored...'*

Response:

Corrected.

5. line 153: *'...enumeration of...'*

Response:

Corrected.

6. line 171: *delete 'evidently'.*

Response:

Corrected.

7. line 174: *'trophic level' means something else. Replace by appropriate phrase.*

Response:

We will replace 'trophic level' with 'trophic state'.

8. line 185: *Why is P gained during the treatment?*

Response:

The acid process can enhance the solubility of P in the mineral dust.

9. line 189 *Here cite Fig. 3, otherwise the reader does not know where is that increase reported. Tables do not report the nutrient increases observed in the treatments.*

Response:

Corrected.

10. line 194 and elsewhere (including Fig. legends). *The phrase 'successive increase' should be omitted. Sentence should read simply: 'During the incubation...'*

Response:

The duration of the incubation experiment in this study is 9~10 days, but we mainly analyzed the data on the initial 3-5 days, in which the Chl *a* concentration showed a successive increase. Thus, we used the ‘successive increase’ to distinguish the initial 3-5 days from the whole incubation period (9-10 days).

11. line 271. Remove ‘certain amount of’.

Response:

Corrected.

12. line 300. This sentence seems to assume that all N present in the dust becomes bioavailable, because the concentrations referred to are those given in Table 3 (which correspond to concentrations in the dust, not in seawater).

Response:

We will rewrite the sentence. The contents of N and P in the untreated and AM-dust listed in Table 3 were determined in seawater. The details can be seen in the description of ‘ultrasonic bath’.

13. line 448-449. Here the authors are deriving biogeochemical conclusions on the functioning of the biological pump, but their data consider just phytoplankton. Without information on how the metabolic activity of heterotrophs, bacteria in particular, changes in response to nutrient/dust additions, the ultimate effect on the biological pump remains unknown.

Response:

Thank you for your thoughtful suggestion. We will rewrite the sentence and illustrate that the enhanced biological pump was only a possibility.

14. Table 1. Silicate measurements are missing – they would have been helpful to constrain the stoichiometry of diatom blooms in response to nutrient/dust amendments.

Response:

In the original experimental settings, we aimed at exploring the relationship between phytoplankton growth and supplied bioavailable nutrients (N, P, and Fe) by AM-dust additions. Thus, we did not determine silicate (Si) concentrations. For the Yellow Sea, Si is generally not a limiting nutrient for the growth of phytoplankton in spring because of the influence of riverine input (Wang et al., 2003). Si may become a limiting nutrient during the diatom blooms in the open ocean of the northwest Pacific, but it will not influence the result of this study, which was mainly concerned with N, P, and Fe nutrients. We thank the comment and will add the measurement of silicate for future incubation experiments.

Wang, B. D., Wang, X. L., and Zhan, R. Nutrient conditions in the Yellow Sea and the East China Sea. *Estuarine, Coastal and Shelf Science*, 58, 127-136, doi: 10.1016/S0272-7714(03)00067-2, 2003.

15. Table 3. The data labelled 'increased concentrations' are theoretical or expected concentrations, assuming 100% of the nutrients in the dust becomes dissolved. This should be explicitly acknowledged in the Table legend. Is there any evidence to support the tenet that, upon dust deposition onto the ocean's surface, all nutrients become dissolved and bioavailable?

Response:

We will add 'theoretically' in the Table legend.

The  $\text{NO}_3^- + \text{NO}_2^-$  in the AM-dust would dissolve almost thoroughly once exposing to the filtered seawater (Ridame et al., 2014) while the dissolution of  $\text{PO}_4^{3-}$  would continue over multiple days because the dissolution of less labile (but still soluble) P compounds would take some time to dissolve in seawater (Mackey et al., 2012). But phytoplankton in the incubated bottles can absorb all the soluble  $\text{NO}_3^- + \text{NO}_2^-$  and  $\text{PO}_4^{3-}$  from AM-dust if they need.

Ridame, C., Dekaezemacker, J., Guieu, C., Bonnet, S., L'Helguen, S., and Malien, F.: Contrasted Saharan dust events in LNLC environments: impact on nutrient dynamics and primary production, *Biogeosciences*, 11, 4783-4800, doi: 10.5194/bg-11-4783-2014, 2014.

Mackey, K. R., Roberts, K., Lomas, M. W., Saito, M. A., Post, A. F., and Paytan, A.: Enhanced solubility and ecological impact of atmospheric phosphorus deposition upon extended seawater exposure, *Environmental science & technology*, 46, 10438-10446, doi: 10.1021/es3007996, 2012.

16. Figure legends: The phrase 'successive increase' is awkward. Delete in all fig. legends. It should be simply: 'changes in xxxx during the incubation period at each station'.

Response:

Please see 'Minor comments-No.10'.

17. Fig. 2. Y-axis intervals should be regular (e.g., 0.5 or 1.0 ug/L) and consistent in all plots. Minor ticks should be included, to help the reader ascertain the magnitude of responses.

Response:

Corrected.

18. Fig. 3. Symbol for control should be more visible (it is often masked by other symbols).

Response:

Corrected.

19. Fig. 5. Revise species names spelling (e.g. *Skeletonema*). Species names and genera should be written in italics (but not 'spp.').

Response:

Corrected.

20. Figs. 6 and 7. The index values shown here result from the subtraction and division of variables measured independently, each with its own error. Therefore the error bars shown should be computed using the error propagation formulae for addition and division.

Response:

Thanks for the valuable suggestion. We will use the error propagation formulae to calculate the error bars.

21. Fig. 8. These N:P ratios should be defined in the Methods section. Strictly speaking, the N:P supply ratio is not known, since no solubility experiments were conducted.

Response:

We will move the definitions of N:P ratios to the Methods section. The dissolved concentrations of N and P from AM-dust in the seawater have been determined in the laboratory (Table 3). Please also see ‘Specific comments-No.5.

## Referee #2

### General comments

1. Biological productivity of the open ocean regions, especially oligotrophic parts, have attracted the attention of several researchers. Our present understanding of the role of mineral dust in enhancing the primary productivity of the oligotrophic ocean by supplying bio-available nutrients is still in its infancy. The present manuscript aims at advancing such an understanding of enhancement of phytoplankton growth and the phytoplankton community structure by the nutrient supply from the Asian in the subtropical gyre, Kuroshio Extension of Northwest Pacific Ocean and Yellow Sea region. The authors attempt this by incubation experiment onboard R/V Dongfanghong II during spring. For incubation experiment, the mineral dust collected from Gobi Desert region was artificially modified and phytoplankton assemblages from subtropical gyre, Northwest Pacific ocean and Yellow Sea region were used. Each of the five microcosm experiment lasted for 9 to 10 days. Using a net conversion efficiency index, proposed by the authors, of nitrogen conversion to chlorophyll a the authors explore the role of bio-available nutrients from mineral dust in the primary production at the above mentioned three regions.

2. The subject matter of the manuscript addresses an important aspect of phytoplankton growth by “bio-available nutrients” from “treated soil from Gobi desert” which is “expected to” simulate the natural mineral dust. Though there are several concerns that have been listed under specific comments, in my opinion, the results are publishable; but only after adequately addressing the concerns.

Response:

We greatly appreciate the referee for the thoughtful comments and will revise our manuscript accordingly.

Major concerns:

*1. The major concern is that the authors have not succeeded in unambiguously resolving the issue of quantification of bioavailability of nutrients from artificially modified mineral dust, especially phosphorous, for phytoplankton growth, which is the central theme of the manuscript. See for example, lines 334-341. The ambiguity regarding the 'missing N and P'.*

Response:

We are sorry that the word of 'missing' in the manuscript confused the referee. The 'missing N and P' and 'missing parts' in the text meant the N and P might be adsorbed on the bottle walls, suspended particles, and phytoplankton in the solution, while 'missing sources' meant there might be other sources to supply bioavailable P. Thus, we will change 'missing sources' to 'other sources' to express them clearly. Considering the comment and the similar comments from other two referees, we will rewrite the part related to the budget of P in incubation experiments. Please see our revised manuscript.

*2. How would the authors differentiate the phytoplankton growth-response due to N/P/Fe and that due to Mn and Zn (see for example, Saito et al., 2008; Sunda 2012).*

Response:

We used the various nutrient addition experiments to illustrate which nutrients (i.e. N, P, and Fe) limit phytoplankton growth. As we did not conduct Mn or Zn addition incubation experiments and determine the concentration of Mn and Zn in the seawater, it is difficult for us to accurately see whether phytoplankton were under limitation of Mn and/or Zn. However, through making a comparison of the net conversion efficiency index (NCEI) in various nutrient addition treatments, we can roughly determine whether there were other nutrients except N, P, and Fe dissolved from AM-dust to stimulate phytoplankton growth. For instance, the NCEI value in AM-dust treatment was significantly higher than that in N+P+Fe treatments, indicating the N, P, and Fe supplied by AM-dust cannot explain the corresponding phytoplankton growth. As reported in literature, trace metals such as Mn and Zn can affect phytoplankton growth rate (Sunda, 2012), which might lead to higher NCEI. The effect of Mn and Zn on phytoplankton growth is just a hypothesis, which will be clarified in the revision.

Sunda, W.: Feedback interactions between trace metal nutrients and phytoplankton in the ocean, *Frontiers in microbiology*, 3, 204, doi: 10.3389/fmicb.2012.00204, 2012.

*3. The authors do not observe any community shift in the phytoplankton in their study. There is no discussion on this aspect and authors need to address this.*

Response:

We thank the comment and will add some discussions about community shift of phytoplankton in Discussion

## Section 4.2.

*4. Based on the information provided, it is hard to see how closely the artificially modified soil collected from Gobi desert mimics the nature. Some more robust information on the atmospheric (chemical) processing of the mineral dust during the long-range transport from source to the proposed study site during spring is needed along with the upper air wind vectors. What are the chemical constituents of such atmospheric processed mineral dust in presence of anthropogenic aerosol is not clear.*

Response:

During a strong Asian dust event, the loading of inorganic nitrogen ( $\text{NO}_3^- + \text{NH}_4^+$ ), soluble P and Fe in atmospheric particles collected in the Yellow Sea was  $\sim 714 \mu\text{mol g}^{-1}$ ,  $\sim 4.3 \mu\text{mol g}^{-1}$  and  $\sim 550 \mu\text{g g}^{-1}$ , respectively (Shi et al., 2012). The values in AM-dust used in this study are  $577 \mu\text{mol g}^{-1}$ ,  $4.3 \mu\text{mol g}^{-1}$  and  $473 \mu\text{g g}^{-1}$ , respectively (Table 3 in the origin version), which were highly comparable to those observed by Shi et al. (2012). The authors, however, agree that the loadings of these components in atmospheric dust particles could highly vary in different cases. The weakness will be added at the end of the conclusion.

We will add a description of preparing AM-dust in the Text S1:

In this study, the aging process of dust followed Guieu's (2010) method and aimed at stimulating the cloud reaction between dust and synthetic evaporating cloud water. The pH around dust in the cloud process (i.e. mix with evaporating cloud water) was found to be as low as  $\sim 1$  during their transport to the Yellow Sea (Meskhidze et al., 2003), whereas the typical pH in rainwater is 5 (Watanabe et al. 2001, Sasakawa and Uematsu, 2002, Wang et al. 2002, Sakihama et al. 2008, Zhang et al. 2011), meaning that a dilution by a factor  $10^4$ . In consequent, in order to reproduce an evaporating cloud, we have used a concentration 10 000 in our experiments in comparison to the typical concentrations found in rainwater. Considering the typical concentrations of dust in rainwaters was  $10 \text{ mg L}^{-1}$  (Ridame et al., 2002), the dust loading in evaporating cloud water could reach  $100 \text{ g L}^{-1}$ . As a consequence, all of the concentrations in evaporating cloud water were around 10000-fold larger (i.e. 4 orders of magnitude larger) than those in natural rainwater. Table S1 summarized the primary chemical composition of rains in the Eastern Asian regions and the evaporating cloud water used for our simulation.

Shi, J., Gao, H., Zhang, J., Tan, S., Ren, J., Liu, C., Liu, Y., and Yao, X.: Examination of causative link between a spring bloom and dry/wet deposition of Asian dust in the Yellow Sea, China, *Journal of Geophysical Research: Atmospheres*, 117, 2012, doi:10.1029/2012JD017983.

Guieu, C., Dulac, F., Desboeufs, K., Wagener, T., Pulido-Villena, E., Grisoni, J.-M., Louis, F., Ridame, C., Blain, S., and Brunet, C.: Large clean mesocosms and simulated dust deposition: a new methodology to investigate responses of marine oligotrophic ecosystems to atmospheric inputs, *Biogeosciences*, 7, 2765-2784, doi: 10.5194/bg-7-2765-2010, 2010.

Meskhidze, N., Chameides, W. L., Nenes, A., and Chen, G.: Iron mobilization in mineral dust: Can anthropogenic

SO<sub>2</sub> emissions affect ocean productivity?. *Geophysical Research Letters*, 30(21), 2003, doi: 10.1029/2003GL018035.

Watanabe, K., Ishizaka, Y., & Takenaka, C.: Chemical characteristics of cloud water over the Japan Sea and the Northwestern Pacific Ocean near the central part of Japan: airborne measurements. *Atmospheric Environment*, 35(4), 645-655, 2001, doi: 10.1016/S1352-2310(00)00358-7.

Sasakawa, M., & Uematsu, M.: Chemical composition of aerosol, sea fog, and rainwater in the marine boundary layer of the northwestern North Pacific and its marginal seas. *Journal of Geophysical Research: Atmospheres*, 107(D24), 2002, doi: 10.1029/2001JD001004.

Wang, Z., Akimoto, H., & Uno, I.: Neutralization of soil aerosol and its impact on the distribution of acid rain over east Asia: Observations and model results. *Journal of Geophysical Research: Atmospheres*, 107(D19), 2002, doi: 10.1029/2001JD001040.

Sakihama, H., Ishiki, M., & Tokuyama, A.: Chemical characteristics of precipitation in Okinawa Island, Japan. *Atmospheric Environment*, 42(10), 2320-2335, 2008, doi: 10.1016/j.atmosenv.2007.12.026.

Zhang, J., Zhang, G. S., Bi, Y. F., & Liu, S. M.: Nitrogen species in rainwater and aerosols of the Yellow and East China seas: Effects of the East Asian monsoon and anthropogenic emissions and relevance for the NW Pacific Ocean. *Global Biogeochemical Cycles*, 25(3), 2011, doi: 10.1029/2010GB003896.

Ridame, C., & Guieu, C.: Saharan input of phosphate to the oligotrophic water of the open western Mediterranean Sea. *Limnology and Oceanography*, 47(3), 856-869, 2002, doi: 10.4319/lo.2002.47.3.0856.

*5. It is not clear from the manuscript, whether the ocean atmospheric conditions during the 3-month period (March-May 2014) at each of the sampling locations could be considered as a part of the same season where ocean and atmosphere represents similar conditions. The data from the Table 1 do not support this. For example, the average temperature (is it SST?) at S4 is quite different from the rest of the stations, which was sampled in May 2014. Similar, the MLD also is quite different. The authors need to address these issues.*

Response:

We are sorry that we made a mistake of sea surface temperature (SST) and mixed layer depth (MLD) at S5 and will correct them in the revision.

The seawater at S4 and S5 was sampled in May, which was later than the sampling date at other stations (Table 1). But the SST at both stations were lower than those at other stations. Thus the difference of SST between S4 and other stations was not caused by the different season. The S4 and S5 stations are located in the cold water regions, which will make SST in this region lower than that in the open oceans of the northwest Pacific.

The open ocean in the northwest Pacific is accompanied by complicated hydrological conditions such as frequent warm and cold eddies, which lead to strong water mixing in the upper ocean and may influence the MLD. While in the Yellow Sea, the seawater depth at S4 and S5 were lower than 100 m, which lead to a completely different MLD compared to those in the open oceans of the northwest Pacific.

We will add a description of SST and MLD conditions at the sampling locations in Results Section 3.1.

6. *The authors need to discuss the efficacy of the proposed “net conversion efficiency index” in varying Redfield ratio conditions.*

Response:

Good suggestion! We will add some discussions about the efficacy of the proposed “net conversion efficiency index” in varying Redfield ratio conditions in Discussion Section 4.2.

7. *Most of the results obtained from the incubation experiment, such as co-limitation of nutrients, response of phytoplankton biomass and structure, are largely known as could be seen from the literature cited in the manuscript. For example, co-limitation in the south China Sea and its response to aeolian input (Gao et al., 2012), Fu et al. (2009) on N:P ratios during spring, Fu et al (2009) study on the phytoplankton biomass and structure in South China Sea. Nishibe et al. (2015) work in the Kuroshio Extension during spring. See also under Minor comments (8).*

Response:

We recognize that previous studies have identified the nutrient limitation status in the study area. We are content that our nutrient addition results are consistent with theirs, as this confirms that the conditions we were studying were representative to the regions.

The similar results compared to other studies aimed at illustrating the following ideas:

The main strength of the study is to gain insight into the causative mechanisms underlying the phytoplankton responses to dust through parallel incubations, in which inorganic nutrients were added in different combinations. The added amount of inorganic nutrients (e.g. N:  $2 \mu\text{mol L}^{-1}$ , P:  $0.2 \mu\text{mol L}^{-1}$ , Fe:  $2 \text{ nmol L}^{-1}$ ) was not equal to that dissolved from AM-dust, which leads to incomparable increases of Chl *a* concentrations in AM-dust and various nutrient treatments. Thus, we proposed the conversion efficiency index (NCEI) to quantify the role of N, P, and Fe dissolved from AM-dust played in stimulating phytoplankton growth, based on making a comparison with the results concluded from various nutrient addition treatments. Finally, we used the correlation of  $S_{N:P}$  and  $C_{N:P}$  to highlight increased bioavailability of P in AM-dust addition experiments.

Minor concerns:

8. *Lines 310-311: Authors need to at least briefly state what are those “Complex hydrographic conditions”.*

Response:

We will add a brief description of complex hydrographic conditions in the revised manuscript.

9. Lines 339-342: *This is purely speculative and needs further substantiation.*

Response:

We will rewrite this sentence to express it clearly.

In the field incubation experiments, the time interval of adding materials (e.g. AM-dust and various nutrients) into the incubation bottles and nutrient measurement (i.e. measurement on day 0) was around 1-2h. During this period, microbial uptake, scavenging by cell surface and bottle wall are all possible to influence the measurement of added N and P nutrients from AM-dust, which lead to the concentrations determined on day 0 were lower than the theoretical adding amounts. When the concentrations of  $\text{NO}_3^- + \text{NO}_2^-$  and  $\text{PO}_4^{3-}$  decreased in the seawater, the  $\text{NO}_3^- + \text{NO}_2^-$  and  $\text{PO}_4^{3-}$  absorbed by cell surface and bottle wall had the potential to be released into the solution again for reaching equilibrium.

10. Lines 307-308: *So what is new/different from the work of Nishibe et al. (2015).*

Response:

Please see Major concerns No.7.

11. Line 143: *Expand SPSS*

Response:

Corrected.

12. Table 3 : *2ns foot note (b) is missing in the Table*

Response:

Corrected.

13. *Also explain "E-3, E-4"*

Response:

We will add an explanation of 'E-3' and 'E-4' in Table 3.

## Referee #3

### *General comments*

*This paper present a set of microcosm experiments performed on-board using sea water collected from two distinct oceanic regime: 1) Oligotrophic waters (Northwest Pacific Ocean) and 2) nutrient rich waters (Yellow Sea), to*

*understand the impact of atmospheric dust (or processed dust) and other nutrients (N, P, Fe e.t.c) on the phytoplankton productivity in terms of increase in chlorophyll a (Chl a) and abundance of phytoplankton in various size fractions (e.g. micro, nano, pico e.t.c). The artificially modified atmospheric dust (AM-dust) is prepared using surface soil collected from Gobi Desert. The set of experiments lasted for 9-10 days and clearly indicate an overall increase in Chl a concentrations due to addition of various combination of nutrients including AM-dust. Authors have proposed a “new” net conversion efficiency index (NCEI) to better understand the impact of specific nutrients (N, P, Fe, and AM-dust) on primary productivity at the sampled locations. The presented case study makes an important contribution towards improving our understanding on impact of aeolian deposition (external source of nutrients) on productivity and ocean biogeochemistry. The results obtained from set of experiments are discussed well, manuscript is easy to read (except few sections, see in specific comments) and should be of great interest to the Biogeoscience community. So, I recommend this paper for publication in Biogeosciences, but after addressing some of the concerns detailed in specific comments.*

**Response:**

We greatly appreciate the referee for the constructive comments and will revise our manuscript accordingly.

*Specific comments:*

*1 Section 2.1: What is the size distribution of soil samples used for preparing AM-dust? This information is important because if, majority of collected soil particle are in coarse fraction (e.g. more than 30 microns), most of them gets deposited at the source region and hardly get transported to the Pacific. So, the soil used for AMdust preparation is not at all a representative undergoing long-range transport and depositing on surface waters. The fine fraction (less than 5 microns) or typically clay fraction of the soil is a more representative dust which can be artificially processed to mimic the processed aeolian dust.*

**Response:**

Recently, the transport routes of Asian dust move the northward. We had practical difficulty to collect the sufficient amount of ambient dust samples for incubation experiments. Alternatively, we used AM-dust for experiments as those reported by Guieu and Ridame, etc (Guieu et al., 2010; Ridame et al., 2014). In this study, we did not determine the size distribution of soil samples, but only the fraction less than 20  $\mu\text{m}$  was used for preparing AM-dust in this study. Fe and P composition in  $\text{PM}_{10}$  and  $\text{PM}_{20}$  generated from the same soils were reported to be quite similar (Shi et al., 2011; Nenes et al., 2011). We agree that finer dust, e.g., less than 5 microns, should be more representative of those in the aeolian dust transported to the sea. However, it is also practically difficult to gain the sufficient amount of the fine dust for modification. Thus, it is a practical compromise by using artificially modified  $\text{PM}_{20}$ . The information will be added in the revision.

Guieu, C., Dulac, F., Desboeufs, K., Wagener, T., Pulido-Villena, E., Grisoni, J.-M., Louis, F., Ridame, C., Blain, S., and Brunet, C.: Large clean mesocosms and simulated dust deposition: a new methodology to investigate

responses of marine oligotrophic ecosystems to atmospheric inputs, *Biogeosciences*, 7, 2765-2784, doi: 10.5194/bg-7-2765-2010, 2010.

Ridame, C., Dekaezemacker, J., Guieu, C., Bonnet, S., L'Helguen, S., and Malien, F.: Contrasted Saharan dust events in LNLC environments: impact on nutrient dynamics and primary production, *Biogeosciences*, 11, 4783-4800, doi: 10.5194/bg-11-4783-2014, 2014.

Nenes, A., Krom, M. D., Mihalopoulos, N., Cappellen, P., Shi, Z., Bougiatioti, A., Zampas, P., and Herut, B.: Atmospheric acidification of mineral aerosols: a source of bioavailable phosphorus for the oceans, *Atmospheric Chemistry and Physics*, 11(13): 6265-6272, doi: 10.5194/acp-11-6265-2011, 2011.

Shi, Z., Bonneville, S., Krom, M. D., Carslaw, K.S., Jickells, T. D., Baker, A.R., and Benning L.G.: Iron dissolution kinetics of mineral dust at low pH during simulated atmospheric processing, *Atmospheric Chemistry and Physics*, 11(3): 995-1007, doi: 10.5194/acp-11-995-2011, 2011.

*2. Section 2.2: Line 110-112: Why Day 1 was not sampled?*

Response:

In our previous incubation experiments, there was no distinct difference of Chl *a* concentrations on day 1 and day 0 in all cases (Liu et al., 2013). Thus, we did not take a sample on day 1 in this study. We thank the comment and will change back our sampling protocol for future incubation experiments. We agree that the data on day 1 may be valuable. This will be clarified in the revision.

Liu, Y., Zhang, T., Shi, J., Gao, H., and Yao, X.: Responses of chlorophyll *a* to added nutrients, Asian dust, and rainwater in an oligotrophic zone of the Yellow Sea: Implications for promotion and inhibition effects in an incubation experiment, *Journal of Geophysical Research: Biogeosciences*, 118, 1763-1772, doi: 10.1002/2013JG002329, 2013.

*3 Section 2.4: The ultrasonic bath treatment may overestimate the nutrient concentration. What was the time duration used for ultra-sonication? More than 30 minute of treatment will increase the temperature and may enhance the leaching of nutrients and thus overestimate. Usually, the treatment is done for aerosol samples collected on filter substrate to loosen the particles from matrix.*

Response:

The time duration used for ultra-sonication is 30 minute. We used the ice pack to keep the temperature of water bath stable at ~0°C.

*4. Section 4.1: Line 328-329: This is a speculative statement and of course, it need further investigation. Except here, the role of trace metals (as nutrient or toxicant) is nowhere discussed. The concentration of suite of trace metals (in AM-dust) are very low except Fe and Mn. This once again indicate that, AM-dust is not the best representative for processed dust.*

Response:

The sentence will be revised as “Micro-nutrients, e.g., Mn, (Coale, 1991; Jakuba et al., 2008; Saito et al., 2008; Sunda, 2012) from the AM-dust may have contributed to the phytoplankton growth in the incubations at S2. This potential synergistic effect is worthy of further investigation.”

We will also revise the part related to the role of trace metals (as nutrient or toxicant) other than Fe and Mn. As showed in our Table 3, most of trace metals are indeed negligible.

The aging process of dust in this study focused on the reaction between dust and inorganic acids ( $H_2SO_4$  and  $HNO_3$ , details can be seen in Text S1). The distinguishing characteristic of AM-dust relative to untreated dust mainly reflects in the increased contents of soluble N, P, and trace metals. Trace metals Fe and Mn were mainly originated from mineral aerosols, thus we can observe noticeably enhanced solubility of Fe and Mn in the AM-dust relative to untreated dust. The contents of other soluble trace metals such as Cu, Pb, and Zn in the dust were mainly affected by anthropogenic factor such as automobile exhaust and coal combustion, which were not reflected well in our study. To this point, we will add an illustration at the end of the conclusion.

*5. Section 4.2: It is very difficult to follow the proposed conversion efficiency index (NCEI). It may be a good tool to specifically understand the role of nutrients on nitrogen consumption or productivity, but need to be elaborated more. It is not clear, why summation of differences of treatment and control for consecutive days are used?*

*The proposed conversion efficiency index (NCEI).*

Response:

In the revision, we will clarify the net conversion efficiency index (NCEI) proposed in this study to be an approximate estimation for the utilization of N for the growth of phytoplankton. Therefore, the capacity to synthesize Chl *a* per unit concentration of nitrogen (N) in different treatments can be compared. We agree that the sum consecutive differences over time in Chl *a* concentration between treatments and control will lead to an overestimation of the real net conversion efficiency because of the accumulation effect. Theoretically, the use of the maximum difference will lead to an underestimation of the real net conversion efficiency because of degradation of Chl *a* in the growth period. The real net conversion efficiency should be between those calculated by the two approaches.

We will add a more elaborated description of NCEI in Discussion Section 4.2.

*6. Section 4.3: Line 408-419: This paragraph is mostly speculative and difficult to follow, although authors have concluded the importance of DOP determination in seawater.*

Response: The part is particularly important because it provides a new insight to analyze the budget of bioavailable P in the AM-dust addition incubation experiments. Unfortunately, the original presentation seemed to be unclear and didn't service the target well. We will rewrite this paragraph to make our thoughts understandable.

Minor comments:

*7. Line 181: should be Table 1.*

Response:

Corrected.

*8. The legends used in Fig. 2, 3 and 4 for nutrients (other than AM-dust) are in same colors and very hard to make out. Most of them are superimposed. It will be useful for reader if different coloured legends with connecting lines can be used.*

Response:

Corrected.