Author responses to reviews and edits to Biogeosciences manuscript bg-2016-159 "Fate of terrigenous organic matter across the Laptev Sea from the mouth of the Lena River to the deep sea of the Arctic interior"

by Lisa Bröder, Tesi, Salvadó, Semiletov, Dudarev and Gustafsson

We are grateful to the two reviewers for their detailed and insightful comments on our manuscript. We are naturally delighted of the overall supportive assessments. Their constructive reviews and suggestions have contributed to substantially improve the paper during our revisions. All referee comments and our responses, as well as the resulting edits, are detailed below, organized such that first the reviewer comments are given in italic, directly followed by our response and outline of the resulting edit in regular font. References in our response to line numbers refer to the revised manuscript version (with tracked changes).

Reviewer #1

GENERAL COMMENTS:

“Degradation of terrestrial organic matter (TerrOC) along its transport into the ocean is an extensively investigated yet not fully understood aspect of the global carbon cycle. The Siberian Arctic Shelf, with a width > 800 km, is a unique and ideal place to study the transformation of TerrOC upon its entry into the sea. In particular, with climate-induced mobilization of permafrost-locked TerrOC, this area is receiving greater attention than many other shelves on earth. This paper uses a series of terrestrial biomarkers (including lignin phenols, cutin acids, and wax lipids) to study the abundances as well as degradation of TerrOC along a 800 km transect from Lena River mouth across the shelf to the slope and rise. In conjunction with carbon isotope and surface area analyses, the authors display an increasing TerrOC degradation with increasing distance from the coast. The dataset is large and unique, and the writing is clear and organized. I have a few suggestions for the authors to consider.

First, in the Results and Discussion, many comparisons are made to other published Arctic (or other non-Arctic) studies, which is great and necessary. But the text is a bit reiterative and I wonder if there is a better way to display all the information from literature with tables or figures more vividly, which will help the readers to digest.
Second, TerrOC degradation is not unique to permafrost-derived OC: it happens in other shelf environments without permafrost OC input. What I am interested in this study is what is special about the transformation of TerrOC in the Siberian Arctic Shelf in comparison with other parts of the world or other depositional environment. I think the authors have made some very nice comparisons with the Mackenzie Shelf. But I think this may be more emphasized in the conclusions, etc.”

RESPONSE:

We are glad about the positive appraisal and appreciate the clear, concise and constructive comments. We agree with the suggestion of a more illustrative way for the comparisons made with other studies and have now sought to improve on this. Therefore, we have
inserted an additional figure (Fig. 4), where the lignin phenol data of this study are compared to the literature data for several Arctic Shelf seas. For the other proxies (biomarker concentrations and ratios), we added similar figures in the Supplementary Information (Fig. S1 and S2). Here we recalculated the HMW lipid data to match the definition in previous studies (carbon chain lengths of ≥ 20 for n-alkanoic acids and ≥ 21 for n-alkanes) in order to be truly comparable (see also Response to Reviewer#2).
The Laptev Sea and adjacent East Siberian Sea are among the widest continental margins on Earth. The resulting long transport and time spent in oxic sediments exert first order control on the land-derived OC degradation. Our study area thus is a perfect natural laboratory to test hypotheses on the fate of permafrost carbon in terms of carbon-climate feedback. This is a key aspect which hadn’t been sufficiently discussed in the Introduction and in the Conclusions. We have now updated the text accordingly (L75-79 and 634-637).

SPECIFIC POINTS:

1) “L19: Change to "Mobilized permafrost carbon" can be either…”
This has been changed accordingly in the text.

2) “L147: How does combustion affect surface area measurement?”
From the original text it was not clear that the combustion was done to remove organic material in order to obtain the mineral specific surface area. According to Keil and Cowie (1999), this particular method does not alter the surface area systematically compared to the removal of organic matter with sodium pyrophosphate/ hydrogen peroxide as in Mayer (1994). This explanation has been inserted in the text (L154-157).

3) “L165: What about FeOx, which plays a key role in the preservation of TerrOC?”
First of all, the iron quantified here refers to the bulk iron which includes both fractions associated with OM and detrital material (e.g. clay material). In addition, XRF results are reported only as Fe2O3 as samples are combusted (450C, 12h) prior to the analysis to remove the organic fraction. Thus, we decided not to include the iron data obtained from the XRF measurements here, as we cannot determine the fraction of amorphous iron oxide/hydroxides (of high surface area) with this method and therefore do not obtain any information of the association between Fe and organic matter. Even though not in the focus of this study, we do acknowledge the importance of FeOx in the Laptev Sea and have therefore added a reference to the study by Salvadó et al. (2015), where the association between iron and organic matter on the ESAS is studied in more detail (L67-68).

4) “L346-348: How does hydrodynamic sorting affect the SA-normalized abundance of lignin? This is probably an important aspect (if not more important) other than degradation, which may explain the varied decrease rate for lignin vs. wax lipids. Does this bias the SA-normalized abundances?”
This is indeed an important aspect that we have tried to address with the help of the study by Tesi et al. (2016), which had the objective of disentangling exactly these two processes: degradation and hydrodynamic sorting during cross-shelf transport. We have
now elaborated more on the matter and included some rough estimates of how much of the lignin decrease can be attributed to degradation vs sorting (L449-459). According to Tesi et al. (2016) most of the sorting is occurring in close proximity to the coast, i.e. water depths of less than 25 m. Assuming that for the shallowest station up to 75 % of the lignin phenols are associated to the low density/large plant fragment fraction and thus retained close to the shore, we can “correct” for sorting by focusing on the lignin phenols in the fine, mineral associated fraction (reducing the original value for the bulk to 25 %, i.e. 11 instead of 45 µg m⁻²). The cross-shelf loss from about 11 µg m⁻² to 0.43 ± 0.09 µg m⁻² still corresponds to a reduction by 96 ± 1 % that can be attributed to degradation (instead of 99.1 % net loss), which agrees with the values presented in Tesi et al. (2016).

Reviewer #2

GENERAL COMMENTS:

“Does fill a much needed role as it is one of few papers that looks at the fate of terrigenous organic matter as it is carried out past the continental shelf

• They do compare their data to other shelf studies but some of there comparisons are not valid (i.e. explaining differences in HMW degradation in different studies is due to differences in chain length)

• Seems likely that the amount of time spent during cross-shelf transport is correlated with terrigenous organic matter degradation”

RESPONSE:
We appreciate and are encouraged by the positive assessment, yet have paid attention to the constructive criticism and suggestions. In particular, we have recalculated the results for the HMW n-alkanes and n-alkanoic acids using the same chain-length definition as in the studies we are comparing our values to. These values are then compared in the new figures S1 and S2.

SPECIFIC POINTS:

1) “Figure 1 should include coastal currents and could have an inset of where the Laptev Sea is relative to the rest of the arctic”

Figure 1 has been changed accordingly.

2) “Need to justify in the paper that the terrigenous matter in this study is only coming from the Lena River and not from the two other rivers (shown in Figure 1 and are not labeled) that empty into the Laptev Sea”

The other two rivers in Fig. 1, Olenjok and Anabar (names have now been added to the map), discharge combined only less than 7 % of water and total suspended matter (TSM) of the total for the Laptev Sea (less than 10 % of that of the Lena River) according to Gordeev (2004). The second largest river discharging to the Laptev Sea, Khatanga (not in the map, to the west of Anabar), contributes <12 % of total water and <6 % of TSM (~16 % and ~8 % of that of the Lena River, respectively). We therefore assumed that the largest fraction of riverine-delivered TerrOC should originate from the Lena River (>70 %
of both water and TSM discharge to the Laptev Sea) for both water and TSM. The overwhelming importance of the Lena River as sediment source to the Laptev Sea has now been more emphasized in the text (L101-102).

3) “Need specific references when discussing what TOC/SA ratios are expected for what kind of environment (i.e. river, deep ocean; lines 248-253).”

The corresponding references (Mayer, 1994; Mayer et al., 2002; Aller and Blair, 2006) have been added to the text (L264, L267, L268).

4) “Line 272: Uses a lateral transport time for an active margin instead of one of a passive margin. Suggest using an east coast system from the U.S.”

Unfortunately we could not find any quantitative constrains for lateral transport times of OC across other margins. The numbers presented by Keil et al. (2004) should serve only as a rough estimate of the timescale to test if the explanation of ageing during transport could hold. We state in the text that transport across the wide Laptev Sea is expected to take much longer.

5) “Along the same lines as bullet point number 2 in this section, if you cannot prove the source of this OC is the same, then you can not prove that it is aging”

As stated earlier, the main POC sources for the Laptev Sea are coastal erosion and Lena River, followed by a much smaller contribution from marine sources. Preferential degradation of the modern source (marine) could also shift the 14C signature towards lower (older) values. This alternative explanation, yet not supported by the d13C values, has been added to the text (L293-301). Another piece of evidence for protracted transport is the highly reworked condition of the terrestrial material as shown by all degradation proxies, which are discussed in Section 3.3. We also have unpublished stable hydrogen isotope ratio data on HMW n-alkanes (in other manuscript in advanced stage for submission), which do not show any significant difference between the shelf and slope/rise sediments. These results suggest that there is no significant change of input material or preferential degradation of coastal erosion (yedoma) vs riverine TerrOC between shelf and slope/rise.

6) “Lines 307-310: One sentence that has been made into its own paragraph. Should incorporate this sentence with the following paragraph.”

This has been changed accordingly in the text.

7) “Chose HMW wax lipids based on chain length (lines 376-379)”

In this study, we are only reporting concentrations of HMW n-alkanes and n-alkanoic acids. For these compounds, due to their simple chemical structure, the carbon chain-length determines the molecular weight. The terms “long-chained” and “HMW” may therefore be used interchangeably. We have now moved this explanation and definition (cutoff for HMW n-alkanes 23 carbon atoms, for HMW n-alkanoic acids 24 carbon atoms) to the methods section (L234-236).

8) “Need to include Fig. 3B for reference in the parentheses in lines 376-379”

This has been inserted.

9) “Typo: line 388, should say terrigenous d13C endmember, not marine”

Thank you for pointing this out. It has been changed accordingly.
10) “They don’t mention what lignin phenols they used”
We have now included this information (L369-373).

11) “Every time they reference figure 5 in the paper, they should be referring to figure 4 (example: 5A should be 4A)”
Since we have inserted a new figure 4, now these references are actually correct and have therefore not been changed.

12) “The authors then need to include Fig. 5 in the text of their paper once they made the changes to Fig. 4”
References to this figure (now Fig. 6) have been included in the text (L545 and 582).

13) “Figures 2-5 are also very descriptive. Leave the interpretation of the data to the discussion”
We assume that this comment is directed at the relatively long captions of Fig. 2-5. We have now shortened those substantially and removed any interpretations of the displayed data.

14) “Their S/V and C/V explanation (section 3.2.2) should be taken with a grain of salt, the loss of C can make it look like woody material when it is not”
We have explicitly mentioned this alternative explanation for the observed decrease in C/V ratios in the text (L498-500). For S/V ratios, however, we observe the opposite trend (increasing with increasing water depth) as found for degradation by fungi (decreasing, Hedges et al., 1988).

15) “Acid/aldehyde values for the syringyl phenols off the shelf seem too high (Fig. 5A)”
We have double-checked our results for the acid/aldehyde values and did not find anything unusual (similar response for all, peak shapes look fine, data are above quantification limit). Moreover, our findings seem to be consistent with other studies that reported values in a similar range (e.g. Winterfeld et al., 2015), see also new figure S1.

16) “Lines 513-516: statement does not seem accurate and it also applies to a different shelf system”
One of the three transects studied in Tesi et al. (2016) (called W, located in the Laptev Sea) is actually part of the (longer) transect investigated here. But we had apparently over-simplified their findings for the acid/aldehyde ratios. This has been rectified (L556-562).

17) “Lines 537-538: the chain length should not determine which lipids are HMW”
As stated in the response to point 7, in the case of n-alkanes and n-alkanoic acids the molecular weight directly depends on the chain length. We have therefore used the chain lengths for the definition of HMW.

We have also discovered during the review process, that the water depths of two stations had been listed slightly wrong (TB-46 and SW-6: 6 instead of 11 m and 92 instead of 89 m, respectively) and have updated all figures and tables accordingly. All other changes are minor.
Fate of terrigenous organic matter across the Laptev Sea from the mouth of the Lena River to the deep sea of the Arctic interior

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Abstract

Ongoing global warming in high latitudes may cause an increasing supply of permafrost-derived organic carbon through both river discharge and coastal erosion to the Arctic shelves. Mobilized permafrost carbon can be either buried in sediments, transported to the deep sea or degraded to CO$_2$ and outgassed, potentially constituting a positive feedback to climate change.

This study aims to assess the fate of terrestrial organic carbon (TerrOC) in the Arctic marine environment by exploring how it changes in concentration, composition and degradation status across the wide Laptev Sea shelf. We analyzed a suite of terrestrial biomarkers as well as source-diagnostic bulk carbon isotopes ($\delta^{13}$C, $\Delta^{14}$C) in surface sediments from a Laptev Sea transect spanning more than 800 km from the Lena River mouth (~10 m water depth) across the shelf to the slope and rise (2000-3000 m water depth). These data provide a broad view on different TerrOC pools and their behavior during cross-shelf transport. The concentrations of lignin phenols, cutin acids and high-molecular weight (HMW) wax lipids (tracers of vascular plants) decrease by 89-99 % along the transect. Molecular-based degradation proxies for TerrOC (e.g., the carbon preference index of HMW lipids, the HMW acids/alkanes ratio and the acid/aldehyde ratio of lignin phenols) display a trend to more degraded TerrOC with increasing distance from the coast. We infer that the degree of degradation of permafrost-derived TerrOC is a function of the time spent under oxic conditions during protracted cross-shelf transport. Future work should therefore seek to constrain cross-shelf transport times in order to compute a TerrOC degradation rate and thereby help to quantify potential carbon-climate feedbacks.
1 Introduction

Amplified global warming in high latitudes has raised growing concern about potential positive carbon-climate feedbacks. Arctic soils store about half of the global soil organic carbon (Tarnocai et al., 2009), with 60% of this in perennially frozen grounds (Hugelius et al., 2014). With ongoing climate change these vast carbon reservoirs become increasingly vulnerable. Mobilization and transport of old terrigenous organic carbon (TerrOC) into the Arctic Ocean is expected to intensify (Gustafsson et al., 2011) through enhancing river discharge (McClelland et al., 2008) with augmenting sediment loads (Gordeev, 2006; Syvitski, 2002) and accelerating coastal erosion (Günther et al., 2013). This material can be buried in the sediments of the Arctic shelves, transported across the margin towards deeper basins or degraded and re-introduced into the modern carbon cycle as CO₂, thereby not only providing a potential positive feedback to global warming, but also causing severe ocean acidification (Semiletov et al., 2016). The fate of permafrost-released TerrOC in the marine environment is thus crucial for future climate projections, yet insufficiently understood (Vonk and Gustafsson, 2013).

The East Siberian Arctic Shelf (ESAS) is with a width of > 800 km the world’s largest continental shelf. It receives TerrOC both from the erosion of the East Siberian shoreline, largely consisting of organic-rich, late-Pleistocene ice-complex deposits (Yedoma), and via the Great Russian Arctic Rivers, which drain extensive areas of continuous and discontinuous permafrost. The Laptev Sea is a representative for the TerrOC dominated Siberian shelf seas, since its main organic carbon input originates from substantial coastal erosion (as observed in the Buor-Khaya Bay, Sánchez-García et al., 2011; Semiletov et al., 2011; Vonk et al., 2012) and the Lena River, the main fluvial sediment source for the entire ESAS (Holmes et al., 2002).

Previous studies have focused on near-shore areas and the inner shelf (e.g. Bröder et al., 2016; Charkin et al., 2011; Feng et al., 2015; Karlsson et al., 2011; Salvadó et al., 2015; Sánchez-García et al., 2011; Semiletov et al., 2005, 2012, 2013; Tesi et al., 2014; Vonk et al., 2010, 2012, 2014). They
reported large fractions of old TerrOC in particulate organic carbon (POC) and surface sediments close to the coast, using different approaches such as applying carbon-isotope-based source apportionment (e.g. Gustafsson et al., 2011; Semiletov et al., 2005; Vonk et al., 2010, 2012, 2014; Salvadó et al., 2015, for the iron-associated OC fraction in the sediment) and by analyzing terrigenous biomarkers in both surface sediments (e.g. Feng et al., 2013; Stein and Macdonald, 2004; Tesi et al., 2014, 2016) and POC in the water column (e.g. Charkin et al., 2011; Karlsson et al., 2011; Winterfeld et al., 2015a). This is the first study that encompasses sampling stations along the entire transect from the Lena River mouth, across the wide Laptev Sea shelf, to the continental slope and rise. Our major objective was to gain new insights regarding the behavior of different TerrOC pools, in particular investigating potential degradation of permafrost-released material along the land-shelf-basin continuum. The Laptev Sea and adjacent East Siberian Sea are among the widest continental margins on Earth (Jakobsson et al., 2004). The resulting long cross-shelf transport and thereby time spent in oxic sediments might exert first order control on TerrOC degradation (e.g. Keil et al., 2004). Our study area is thus well suited to test hypotheses on the fate of permafrost carbon in terms of carbon-climate feedback. We have therefore characterized TerrOC in surface sediments along the Laptev Sea transect on both bulk and molecular level, exploiting source-diagnostic bulk carbon isotopes ($\delta^{13}$C, $\Delta^{14}$C) as well as an extensive biomarker suite (lignin phenols and cutin acids obtained by alkaline CuO oxidation and high-molecular-weight solvent-extractable lipids, such as $n$-alkanes and $n$-alkanoic acids).
2 Material and Methods

2.1 Study area

The Laptev Sea is the shallowest of the Arctic shelf seas with an average depth of 48 m (Jakobsson et al., 2004). It spans over 498,000 km² with a volume of 24,000 km³ and is located between the Kara Sea and Severnaya Zemlya in the West and the East Siberian Sea and the New Siberian Islands in the East. The main sources of particulate organic carbon (POC) for the Laptev Sea are terrigenous, both from coastal erosion and river runoff (Sánchez-García et al., 2011; Stein and Macdonald, 2004). Marine primary production is limited to on average two ice-free months per year and therefore generally low. Nutrient-poor waters on the Siberian shelves resulting from a strong stratification further impede phytoplankton growth (Sakshaug, 2004).

The destabilization of Pleistocene Ice-Complex Deposits along the coastline is a main sediment source for the Laptev Sea (Rachold et al., 2000). The total POC input from coastal erosion to Laptev and East Siberian Sea is estimated to be between 4.0 Tg yr⁻¹ (Semiletov, 1999; Stein and Fahl, 2000) and 22 ± 8 Tg yr⁻¹ (including net subsea permafrost-carbon erosion, Vonk et al., 2012).

The Lena River is estimated to provide 20.7 Tg of sediment per year (Holmes et al., 2002), i.e. > 70 % of the total riverine input to the Laptev Sea (Gordeev, 2006) (Holmes et al., 2002) with an average water discharge of 588 km³ yr⁻¹ (Holmes et al., 2012). It drains a watershed of ~2.46 x 10⁶ km² (Holmes et al., 2012), of which 77 % is underlain by continuous permafrost (Amon et al., 2012). Water discharge peaks in June, during the spring flood, when about 75 % of total organic carbon is delivered (Rachold et al., 2004). Total POC discharge by the Lena River can be up to 0.38 Tg yr⁻¹ (Semiletov et al., 2011).

Sediment transport pathways are largely influenced by the prevailing atmospheric conditions: During cyclonic summers (i.e. positive phase of the Arctic Oscillation), northerly winds predominate, strengthening the Siberian Coastal Current, which transports Lena River water masses along the coast towards the East Siberian Sea; whereas during anticyclonic summers (i.e. negative phase of the Arctic Oscillation and mainly southerly winds) the Lena...
River plume is exported onto the mid-shelf and towards the deep part of the Arctic Ocean (Charkin et al., 2011; Dmitrenko et al., 2008; Guay et al., 2001; Wegner et al., 2013; Weingartner et al., 1999). Sediment transport in the Laptev Sea is strongly seasonal. The main transport of Lena River water with high concentrations of suspended particulate matter (SPM) towards the mid-shelf takes place shortly after river-ice breakup (Wegner et al., 2005). During the ice-free summer, SPM circulates between inner and mid-shelf with very little material escaping over the shelf break to the deeper parts of the Arctic Ocean. Significant sediment export is suggested to happen during freeze-up through SPM that is incorporated in sea ice and then transported across the continental margin (Dethleff, 2005; Eicken et al., 1997) or through the formation of dense bottom water resulting from brine ejection (Dethleff, 2010; Ivanov and Golovin, 2007). Hardly any sediment transport occurs beneath the ice cover.

Holocene-scale linear sedimentation rates for the Laptev Sea vary between 0.12 and 0.59 mm yr\(^{-1}\) according to \(^{14}\)C dating of marine bivalves (Stein and Fahl, 2004, and citations therein), whereas centennial-scale \(^{210}\)Pb-derived rates for the more recent Laptev Sea can be up to 1.3 mm yr\(^{-1}\) (Vonk et al., 2012). These rates do not seem to be correlated with water depth on the shelf, but values for the continental slope and rise tend to be on the lower end (0.12-0.38 mm yr\(^{-1}\)) (Stein and Fahl, 2004, and citations therein).

2.2 Sampling

Sediment sampling locations span from close to the Lena River mouth and in the Buor-Khaya Bay, across the shelf, to the continental slope and rise, covering a transect of > 800 km with water depths increasing by more than two orders of magnitude. Samples SW-1, SW-2, SW-3, SW-4, SW-6, SW-14, SW-23 and SW-24 were collected during the SWERUS-C3 expedition on IB ODEN during summer 2014 using an Oktopus multicorer (8 Plexiglas tubes, 10 cm diameter). All other samples were collected during the International Siberian Shelf Study (ISSS-08) expedition onboard the RV Yacob Smirnitskyi during summer 2008. The YS-4, YS-6, YS-13 and YS-14 samples were taken with a GEMAX gravity corer (2 Plexiglas
tubes, 9 cm diameter); YS-9 and TB-46 were collected with a Van Veen grab sampler. For the grab samples only surface sediments (uppermost cm) were subsampled and used in this study. Sediment cores were cut into 1 cm slices within 24 hours after sampling. To account for lower sediment accumulation rates on the rise, for SW-1, SW-2, SW-3 and SW-4 a higher resolution of 0.5 cm for the top 10 cm was chosen. The depositional age for all samples is thus between ~8 and ~70 years (depending on which sedimentation rates are employed). All samples were kept frozen throughout the expedition and freeze-dried upon arrival to Stockholm University laboratories. See Semiletov and Gustafsson (2009) for more information on the ISSS-08 expedition. For exact sampling locations see Table 1.

2.3 Surface area

All surface area analyses have been performed on a Micromeritics Gemini VII Surface Area and Porosity analyzer. Freeze-dried subsamples of ~0.7 g were furnace-dried at 400 °C for 12 h and gently cooled down to room temperature to remove all organic material. Keil and Cowie (1999) have shown that this method yields statistically similar results to the method using removal with sodium pyrophosphate/ hydrogen peroxide (Mayer, 1994). They-The samples were then desalted by repeated mixing with 50 ml of MilliQ water and centrifugation (20 min at 8000 rpm), followed by further freeze-drying. Directly prior to analysis they were degassed in a Micromeritics FlowPrep 060 Sample Degas System for 2 h at 200 °C under a constant nitrogen flow. Each analysis was initiated by measuring the free space in the vial. The specific surface areas were derived from 6 pressure-point measurements (relative pressure p/p0 = 0.05-0.3, equilibration time 5 s) with nitrogen as adsorbing gas (Brunauer et al., 1938). The instrumental error-precision was 0.1-0.3 m² g⁻¹, which corresponds to a relative error-uncertainty of about 1 %. The performance of the instrument was monitored with the surface area reference material Carbon Black (21.0 ± 0.75 m² g⁻¹) provided by Micromeritics.

2.4 X-Ray Fluorescence
The mineral composition of ~1 g freeze-dried, homogenized subsamples was also characterized with a wavelength dispersive sequential Philips PW2400 X-ray Fluorescence (XRF) spectrometer. Prior to the analysis, sediment samples were combusted for 12h at 450 °C to remove the organic fraction. The XRF was operated under vacuum conditions on samples prepared as glass beads using lithium tetraborate and melted with a fluxer Claisse Fluxy (~1150°C) (Mercone et al., 2001). The relative error was less than 0.6 % for major elements and less than 3 % for trace elements. In this study only SiO$_2$, Al$_2$O$_3$ and CaO are reported.

2.5 Bulk elemental and carbon isotope analysis

Total organic carbon (TOC) concentrations and δ$^{13}$C isotopic composition of total organic carbon (TOC) were determined at the Stable Isotope Laboratory, Department of Geological Sciences, Stockholm University. Homogenized subsamples of ~10 mg were repeatedly acidified (HCl, 1.5 M, Ag capsules) to remove carbonates (Nieuwenhuize et al., 1994). TOC concentrations and δ$^{13}$C isotopic composition were measured simultaneously with a Carlo Erba NC2500 elemental analyzer connected via a split interface to a Finnigan MAT Delta V mass spectrometer. TOC concentrations were blank corrected and the relative error was < 1 %. Stable isotope data are reported relative to VPDB using the δ$^{13}$C notation. Radiocarbon analyses of acidified samples were conducted at the US National Ocean Sciences Accelerator Mass Spectrometry (NOSAMS) Facility of the Woods Hole Oceanographic Institution, USA, according to their standard routines (Pearson et al., 1998). The relative error of the measurements was < 0.5 %. Radiocarbon data are reported using the Δ$^{14}$C notation following Stuiver and Polach (1977).

2.6 Biomarkers

2.6.1 CuO-oxidation products

Microwave-assisted alkaline CuO oxidation was performed according to the method established by Goñi and Montgomery (2000). Homogenized subsamples of 100-400 mg of
sediment (corresponding to 2-5 mg OC) were combined with 300 mg of copper(II) oxide and 50 mg of ferrous ammonium sulfate and oxidized under oxygen-free conditions (degassed NaOH, 8 wt %) at 150 °C for 90 min using an UltraWAVE Milestone 215 Microwave oven. After oxidation, known amounts of trans-cinnamic acid and ethyl vanillin were added as recovery standards. Samples were acidified to pH 1 with HCl (12 M) and repeatedly extracted with ethyl acetate. Anhydrous Na₂SO₄ was added to remove remaining water. The solvent was evaporated and extracts re-dissolved in pyridine. For quantification, subsamples were derivatized with bis-trimethylsilyl-trifluoroacetamide (BSTFA) + 1 % trimethylchlorosilane (TMCS) and analyzed on a gas-chromatograph mass spectrometer in full scan mode (GC-MS, Agilent) using a DB5-MS capillary column (60 m x 250 µm, 0.25 µm stationary phase thickness, Agilent J&W) with a temperature profile of initially 60 °C followed by a ramp of 5 °C min⁻¹ until reaching and holding 300 °C for 5 min. The quantification of lignin phenols, benzoic acids, and p-hydroxybenzenes was achieved by comparison to the response factors (key ions) of commercially available standards. For cutin-derived products, fatty acids and dicarboxylic acids the response factor of trans-cinnamic acid was used as in Goñi et al. (1998).

2.6.2 Solvent-extractable Lipids
Wax lipids were extracted by means of accelerated solvent extraction (Dionex ASE 300) using dichloromethane:methanol (9:1) according to the method described by Wiesenberg et al. (2004). Pre-rinsed stainless-steel vessels were loaded with ~3 g of freeze-dried sediment, filled up with pre-combusted glass beads and pre-combusted glass fiber filters at both ends. Two extraction cycles were performed per sample applying a static pressure of 1500 psi and a temperature of 80 °C for 5 min after a heating phase of 5 min. The flush volume was 50 % of the 34 ml cell size with a purging time of 100 s. Extracts were further cleaned purified (addition of activated Cu for sulfur and anhydrous Na₂SO₄ for water removal) and then separated into a neutral and an acid fraction using BondElut cartridges (bonded phase NH₂, Varian), eluting with dichloromethane:isopropanol
(2:1) for the neutral and methyl tert-butyl ether with 4 % acetic acid for the acid fraction according to the method described by van Dongen et al. (2008a). The neutral fraction was further separated into a polar and a non-polar fraction with an Al₂O₃ column. For each of the three compound classes n-alkanes (neutral non-polar fraction), n-alkanols (neutral polar fraction) and n-alkanoic acids (acid fraction) ~10 mg of one internal standard, d₅₀-tetracosane, 2-hexadecanol and d₃₉-eicosanoic acid respectively, were added to the sediment samples prior to extraction. All fractions were then analyzed on a GC–MS (Agilent) using the same column and temperature program as for the CuO products. The polar and acid fractions were derivatized with BSTFA + 1 % TMCS prior to analysis. Quantification was performed using a 5-point calibration curve with commercially available standards. Here, we only report data for high-molecular weight (HMW) n-alkanes and n-alkanolic acids, where HMW refers to carbon chain-lengths of ≥ 23 for n-alkanes and ≥ 24 for n-alkanolic acids.
3 Results and Discussion

The fate of permafrost-released terrigenous organic carbon (TerrOC) across the Laptev Sea shelf is controlled by competing processes. Degradation and sorting, as well as replacement of TerrOC by autochthonous marine organic matter all co-occur to varying degrees during cross-shelf transport. To disentangle their effects on the fate of permafrost-released TerrOC we first report changes in bulk sediment and OC properties and then focus on differences on the molecular level.

3.1 Characterization of the transect on a bulk level

Bulk total organic carbon (TOC) concentrations decreased across the shelf with highest values (~2 %) at shallow water depths and lowest values on the shelf edge (~0.8 %); at high water depths (> 2000 m) concentrations were slightly higher (~1 %). TOC values and the general pattern were in accordance with previous data from the Laptev Sea (Semiletov et al., 2005; Shakhova et al., 2015; Stein and Fahl, 2004; Vonk et al., 2012) and within the same range of those measured for the North American Arctic margin (Goni et al., 2013).

Normalizing TOC concentrations to the mineral-specific surface area (SA) helps to understand the influence of physical sorting and preferential deposition on the observed TOC trends since SA is correlated to the sediment grain size to a first order approximation. To test if the mineral surface area is altered by the input of autochthonous organisms with siliceous or carbonaceous skeleton (e.g. silicoflagellates/diatoms or foraminifera/shells respectively), the mineral composition of the sediments was examined by X-ray fluorescence analysis. There were no apparent trends with water depth for either SiO$_2$/Al$_2$O$_3$ or CaO/Al$_2$O$_3$; therefore, marine production is not expected to have a measureable effect and SA can thus be regarded as a conservative parameter. This was also confirmed by low biogenic silica concentrations for the Laptev Sea reported earlier (< 1.4 %, Mammone, 1998).

The relationship between TOC and SA has been widely studied on continental margins (e.g. Blair and Aller, 2012; Keil et al., 1994; Mayer, 1994). The TOC/SA ratios of typical river suspended sediments range between 0.4 and 1 mg m$^{-2}$ (Mayer, 1994). TOC/SA ratios > 1
mg m\(^{-2}\) have been found in areas with high TOC supply (e.g. river outlets) and where the deposited organic matter had spent little time under oxic conditions (short oxygen exposure time, OET) (Mayer et al., 2002). Ratios < 0.4 mg m\(^{-2}\) generally correspond to sediments from deeper parts of the ocean and long OETs (e.g. Aller and Blair, 2006). Accordingly, the TOC/SA values along the Laptev Sea transect displayed a strong decrease from 2.2 and 1.7 mg m\(^{-2}\) close to the Lena River delta (water depths of 11 and 7 m, respectively) to about 0.3 mg m\(^{-2}\) at water depths greater than 2000 m (Fig. 2A), proposing extensive TOC loss during cross-shelf transport.

Bulk TOC isotopes have been broadly used to distinguish between organic matter sources. Radiocarbon isotopes (\(^{14}\)C) convey information about the age of organic material, with younger OC having higher \(^{14}\)C values. Marine organic matter produced primarily from CO\(_2\) is expected to have modern \(^{14}\)C signatures, whereas permafrost-derived TerrOC has aged both on land and during transport and has thus more depleted \(^{14}\)C values. The \(^{14}\)C values for our Laptev Sea transect were generally low (< -280 ‰, Fig. 2B), suggesting a significant input of pre-aged TerrOC (as in Vonk et al., 2012). Bulk TOC showed less depleted \(^{14}\)C signatures with increasing distance from land on the shelf (from about -500 ‰ to about -340 ‰ on the outer shelf, Fig. 2B), reflecting a dilution of older TerrOC with younger marine material. On the slope and rise, however, \(^{14}\)C values decreased again to about -410 ‰. This difference may be a result of ageing during lateral transport and/or after deposition due to lower accumulation rates on slope and rise. The range between -340 ‰ and -410 ‰ corresponds to a \(^{14}\)C age difference of about 900 years; however, the depositional age differences between shelf and slope samples were estimated to be less than 80 years (see Section 2.2). Ageing after burial alone does therefore not explain the difference in \(^{14}\)C. Keil et al. (2004) estimated a lateral transport time of 1800 years across the Washington margin (158 km) from \(^{14}\)C data of bulk OC in surface sediments. For the > 200 km distance between mid-shelf and rise a bulk ageing of 900 years does therefore not seem unreasonable. It has to be taken into account, however, that mainly the TerrOC fraction of the bulk OC is subject to such protracted lateral transport. Transport times would thus have
to be significantly higher in order to explain this age difference for the entire bulk OC. **One** indication supporting this hypothesis of protracted lateral transport of TerrOC is the degradation status of TerrOC at the deep stations. All molecular degradation proxies point towards highly reworked material (see Section 3.3), suggesting that only the most refractory TerrOC fraction is found at great water depths off the continental margin. Alternatively, the lower Δ¹⁴C values at high water depths may be the consequence of more effective degradation of marine organic matter throughout the water column, resulting in a comparatively lower input of young autochthonous material. However, this latter scenario is not supported by the stable carbon isotopic signature.

For stable carbon isotopes (¹³C), terrigenous sources are generally more depleted than marine organic matter (Fry and Sherr, 1984). In this study, values for δ¹³C of TOC ranged between -26.5 ‰ and -22.3 ‰. The trend towards more enriched TOC with increasing distance from the coast (Fig. 2B) can be explained by a growing proportion of marine organic matter. However, the δ¹³C signature of the marine source appeared to be heavier than typical marine planktonic material in that region (-26.7 ± 1.2 ‰, Panova et al., 2015; -24 ± 3 ‰, Vonk et al., 2012, and citations therein). One possible explanation for this discrepancy is an underestimated influence of ice algae that were reported to have highly enriched δ¹³C values between -15 to -18 ‰ (Schubert and Calvert, 2001). Significant seafloor deposition of ice algal biomass has been observed previously for the Arctic basins (Boetius et al., 2013).

Another option would be a more refractory, isotopically-enriched marine endmember (-21.2 ‰) as suggested by Magen et al. (2010). They argue that lighter isotopes are preferentially consumed by bacteria, which in turn enriches the remaining marine organic matter. Following their reasoning, the more enriched values observed for this transect may be interpreted as an increasing proportion of refractory marine organic matter.

Winterfeld et al. (2015b) analyzed surface water particulate organic carbon (POC) in the Lena River delta and found a mean δ¹³C of -29.6 ± 1.5 ‰. Karlsson et al. (2011) reported similarly depleted δ¹³C values for POC from the Buor-Khaya Bay (-29.0 ± 2.0 ‰), while their mean value for sedimentary OC for the same stations was significantly more enriched (-25.9
and agreed well with our data for the shallow stations (-26.2 ± 0.3 ‰, stations YS-13, YS-14 and TB-46). Lena River POC δ¹³C values from high-discharge periods agree well with the more enriched values we found for the shallow stations (Rachold and Hubberten, 1998). Stein and Fahl (2004), Semiletov et al. (2011, 2012) and Vonk et al. (2012) presented similar δ¹³C ranges and trends for sediments from parts of the Laptev Sea as is reported in the current study for the entire width of the Laptev Sea shelf. For the Arctic Amerasian Continental shelf, Naidu et al. (2000) reported contrasts in absolute δ¹³C values comparing surface sediment samples from different regions, but all commonly displayed an increasing trend for δ¹³C values across the shelf, suggesting a growing fraction of marine organic matter with increasing distance from the coast.

Combining TOC/SA ratios with stable isotope signatures (δ¹³C) may serve to disentangle two different processes, which occur synchronously during cross-shelf transport (as in Keil et al., 1997a): 1.) The net loss of TerrOC and 2.) the replacement of TerrOC with autochthonous marine OC. Net loss of TerrOC, caused by either degradation or hydrodynamic sorting during transport, has been quantified previously using TOC/SA ratios (e.g., Aller and Blair, 2006; Keil et al., 1997a). The carrying-capacity of inorganic particles for OC is assumed to be a function of the SA (Mayer, 1994); a decrease in TOC/SA values can therefore be regarded as TOC net loss. Replacement of TerrOC with autochthonous marine OC does not change this ratio. But since marine OC is known to be isotopically enriched in δ¹³C over TerrOC, this process is recorded by an increasing isotopic signature. Along the Laptev Sea transect, both processes seemed to play an important role (Fig. 2C). High TOC/SA values close to the Lena River decreased sharply **outbound in the nearshore regime**, pointing to extensive net loss, while the increase in δ¹³C values was minor in this area. Once TOC/SA ratios were < 0.8 mg m⁻² (water depths > 20 m), the isotopic changes and thus the replacement of TerrOC with marine OC became increasingly important. Similar trends were observed for the Amazon River delta (Keil et al., 1997b).
However, the TOC/SA trend in the shallower sediments is likely driven by both degradation of OC bound to the mineral matrix during cross-shelf transport and sorting of vascular plant fragments that are retained in the inner-shelf. A recent study (Tesi et al., 2016) has shown that ~50 % of the total OC pool in the inner Laptev shelf surface sediments exists in the form of large vascular plant fragments. They are trapped close to the coast due to their size and resulting settling (Stoke’s law), while the OC bound to the fine mineral matrix is more buoyant and transported offshore towards deeper waters.

3.2 Molecular indicators of organic matter sources

3.2.1 Biomarker distributions

The abundances of different source-diagnostic molecular proxies have been extensively investigated to elucidate complex carbon-cycling mechanisms. In this study, a biomarker suite of CuO oxidation products and solvent-extractable lipids was analyzed in order to gain more insights on TerrOC sources and degradation status along the Laptev Sea transect. All biomarker concentrations were normalized to the sediment-specific surface area (SA) instead of OC content to avoid the signals being overshadowed by other carbon pools. As shown by the lack of water-depth-related changes in the mineral composition (Section 3.1), mineral-matrix dilution by biogenic material is negligible.

Lignin-derived phenols have been widely used to trace TerrOC in the marine environment (e.g. Ertel and Hedges, 1984; Goñi and Hedges, 1995; Hedges and Mann, 1979). The lignin macro-molecule is only synthesized in vascular plants (and certain seaweed species that are not existing in the study area) to render stability to the cell walls. Lignin-derived phenols are typically grouped by phenol type (V: vanillyl phenols, i.e. vanillin, acetovanillone, and vanillic acid; S: syringyl phenols, i.e. syringaldehyde, aceto syringone, and syringic acid; C: cinnamyl phenols, i.e. p-coumaric and ferulic acids). Total lignin refers to the sum of the three groups. Across the shelf, lignin loadings decreased substantially with increasing distance from the coast/water depth (45 µg m⁻² close to the coast, 0.43 ± 0.09 µg m⁻² for the deep stations; loss of 99.1 ± 0.2 %, Fig. 3A).
Cutin-derived hydroxy fatty acids are another compound class obtained from CuO oxidation, which have been used in parallel with lignin phenols (e.g. Goñi et al., 2000; Prahl et al., 1994). They are mainly associated with the soft tissues of vascular plants such as leaves and needles. Cutin acid loadings displayed a similar trend as lignin phenols (11 µg m⁻² close to the coast, 0.061 ± 0.010 µg m⁻² for the deep stations; loss of 99.4 ± 0.1 %, Fig. 3A).

Similar values and sharp declines with increasing distance from the coast for lignin and cutin have been observed for the whole East Siberian Arctic Shelf (ESAS) (Tesi et al., 2014) (Fig. 4 for comparison of lignin phenol concentrations with literature values for different Arctic margins). A recent study (Winterfeld et al., 2015a) for the Buor-Khaya Bay (5.8-17 m water depth) reported lignin phenol concentrations on the same order of magnitude, up to 40 % higher for the shallowest samples, and decreasing with increasing depth. For the Beaufort Sea shelf, Goñi et al. (2000) found a less drastic decline in lignin phenols and cutin acids going from 5 m water depth to 210 m, which likely reflected both lower concentrations in the shallow waters (factor of ~2), and a narrower and steeper shelf. Lignin phenols were also higher at greater water depths than on the ESAS. This may reflect the differences in bathymetry: since the Beaufort Sea shelf is not as wide as, but steeper than, the ESAS, lateral transport is possibly faster, leaving less time for organic matter to be degraded along the way. A comparison between different shelf-slope systems across the North American Arctic margin (Goni et al., 2013) revealed very low lignin and cutin concentrations for the Canadian Archipelago, Lancaster Sound and Davis Strait, whereas both concentrations and trends with water depth for the Beaufort Sea, Chuckchi Sea and Bering Sea were similar to the results from this study. An exception to these patterns was Barrow Canyon, where at water depths of > 2000 m lignin and cutin concentrations were as high as the ones observed close to the Lena River delta, pointing to efficient rapid TerrOC transfer with comparably short oxygen exposure times through this active canyon (Goni et al., 2013) (Fig. 4 and Fig. S1).

Solvent extractable high-molecular weight (HMW) n-alkanes and n-alkanoic acids make up the major part of epicuticular leaf waxes (Eglinton and Hamilton, 1967) and have been
broadly employed as TerrOC biomarkers (for the Arctic Ocean e.g. van Dongen et al., 2008; Yunker et al., 1995, 2005). HMW wax lipids in this study also presented a decreasing trend with increasing water depth/distance from the coast, but to a lesser extent than lignin phenols or cutin acids (HMW n-alkanes, carbon chain lengths of 23-34: 1.1 µg m⁻² close to the coast, 0.12 ± 0.02 µg m⁻² for the deep stations; HMW n-alkanoic acids, chain lengths 24-30: 12 µg m⁻² close to the coast, 0.42 ± 0.29 µg m⁻² for the deep stations; loss of 89 ± 2 % and 96 ± 3 %, respectively, Fig. 3B).

Previous studies in the same area reported similar lipid biomarkers concentrations, which confirm the magnitude of the decreasing trends with increasing water depth (Karlsson et al., 2011; Vonk et al., 2010) (Fig. S1). HMW n-alkane concentrations in the Beaufort and the Chuckchi Sea (Belicka et al., 2004; Yunker et al., 1993) are in accordance with the ones measured on the ESAS, but the shallowest sample on the Beaufort Shelf is ~2 times lower than the shallow ESAS samples (Fig. S1). This might imply that sediments transported by the Mackenzie River to the Beaufort Shelf have lower TerrOC concentrations than Lena River transported sediments. For the Mackenzie Shelf, Goñi et al. (2000) used lignin phenols and cutin acids to estimate a marine terrigenous δ¹³C endmember and therewith derived a terrigenous contribution of almost 80 % for the shallowest sediments, while rough estimates from C/N and δ¹³C data suggested that TerrOC made up only 30-50 % of the organic carbon (Macdonald et al., 2004). For the Lena Delta, source apportionment calculations using δ¹³C and Δ¹⁴C data attributed up to 83 % of the organic carbon to terrigenous sources (Vonk et al., 2012).

All TerrOC biomarker loadings displayed a strong decrease across the shelf, but their relative losses differ substantially between compound classes (Fig. 3C). These findings agree with previous results for the ESAS (Tesi et al., 2014), where similar differences between biomarkers were reported. A somewhat larger decrease was observed for lignin than for cutin, in contrast to this study. The different extents of biomarker losses for the different compound classes may either be attributed to preferential degradation of lignin phenols and cutin acids, implying that they are more labile than HMW n-alkanes and n-alkanoic acids, or
sorting during transport, suggesting that they are associated with a sediment fraction that is hydraulically more retained and carried less efficiently to the outer shelf/slope. A recent study (Tesi et al., 2016) aimed to disentangle these two processes by analyzing different fractions of bulk surface sediments from three transects (yet with only three stations each) across the ESAS. The fractions were separated according to density (1.8 g cm$^{-3}$ cutoff), size (>63 µm, 38-63 µm, < 38 µm) and settling velocity (1 m d$^{-1}$ cutoff). The highest lignin phenol abundance was found in low-density plant fragments (26-55 mg g$^{-1}$ OC). These large particles have a higher settling velocity (Stokes’ law) and are therefore hydraulically retained close to the coast. Cutin acids and plant wax lipids were mainly associated with the mineral high-density fine (< 38 µm, > 1 m d$^{-1}$) and ultrafine (< 38 µm, < 1 m d$^{-1}$) mineral fractions. Within the fine and ultrafine fractions, which made up about 95% of the organic carbon on the outer shelf, they found drastic losses of all biomarkers with increasing distance from the coast, which they attributed to degradation during the protracted cross-shelf transport. Relative decreases appeared to depend on the number of functional groups of the compound class: 98 ± 1% for lignin phenols, 97 ± 1% for cutin acids, 96 ± 1% for HMW n-alkanoic acids and 89 ± 4% for HMW n-alkanes. According to that study, the steep cross-shelf gradients observed here for lignin phenols can be attributed to both hydrodynamic sorting close to the coast and degradation during transport. From the data in the current study alone, the two processes occurring in parallel - degradation and sorting during cross-shelf transport - cannot be disentangled. However, using the data from (Tesi et al., 2016), we can make a rough correction for the sorting part to derive an estimate of the net extent of degradation. For the shallowest station in their study (same as here, TB-46), about 75% of the lignin phenols were associated with the low density fraction that was retained close to the coast. If we thus assume only 25%, i.e. 11 of the 45 µg m$^{-2}$ to be associated with the fine fraction that is actually transported across the shelf, we obtain a reduction by 96 ± 1% that can be attributed to degradation (compared to 99.1% net loss). These results agree with the values presented in (Tesi et al., 2016). For the other compounds analyzed 55-74% are...
Degradation after burial is assumed to play only a minor role in this study. Differences in sedimentation ages are expected to be small (Section 2.1) and a study on centennial-scale sediment cores from the East Siberian Sea (Bröder et al., 2016) detected no significant TerrOC degradation (as recorded by biomarker loss) with increasing sediment depth. Also in that study, lignin phenol and cutin acid loadings were on average 20 times higher on the inner than on the outer shelf, whereas for HMW n-alkanoic acids and n-alkanes the difference between inner and outer shelf was only a factor of ~3-5. Contrasts between the stations were found to be larger than down-core changes. This may be due to the fact that the cores in that study only encompassed about one century of sedimentation ages, while the protracted cross-shelf transport possibly likely requires much longer timescales.

3.2.2 Lignin Phenol sources

Relative distributions of different lignin phenol classes reveal more information on TerrOC sources since they are specific to different plant types. Syringyl phenols are not produced by gymnosperm (non-flowering) plants; elevated syringyl to vanillyl ratios (i.e. S/V > 1, Hedges and Parker, 1976) are therefore attributed to more lignin phenols from angiosperm (flowering) plants. These ratios have to be handled with care, though, because the preferential degradation of syringyl phenols by white- and brown-rot fungi on land can also alter S/V ratios (Hedges et al., 1988). S/V values for the Laptev Sea transect increased with increasing water depth from ~0.65 for the inner shelf to ~1.0 for the slope/rise sediments (Fig. 5A). This trend can either be explained by preferential degradation of gymnosperm material or sorting during transport. Tesi et al. (2014) measured generally lower values for S/V (ESAS average: 0.47, for only Lena watershed dominated locations: 0.42) recording no trend with water depth (Fig. S1 for comparisons with other studies). Their deepest station was located at only 69 m water depth, though, whereas in this study sediments from down to 3146 m water depth were analyzed. S/V ratios in Buor-Khaya Bay surface sediments
(Winterfeld et al., 2015a) were also lower (0.43 ± 0.02 on average) and displayed no trend with water depth. Within the water depth interval they studied (5.8-17 m), however, the samples analyzed in this study had also quite homogeneous S/V ratios (0.64 ± 0.02). Two sediment cores from the East Siberian Sea (Bröder et al., 2016) showed also lower S/V values (inner shelf surface sediment: 0.62, outer shelf surface sediment: 0.50) displaying no clear trends over time/down-core. For the Beaufort Sea shelf Goñi et al. (2000) detected rather high values (0.54-1.71), which (besides the very high value at 61 m water depth) agree with the data from this study. Other transects across the North American Arctic margin (Goni et al., 2013) had slightly lower S/V ratios with no observed trends with water depth.

The ratio of cinnamyl to vanillyl phenols (C/V) is associated with the relative contributions of woody versus soft material, because only non-woody vascular plants synthesize cinnamyl phenols (Hedges and Mann, 1979a). This ratio admittedly decreases with ongoing degradation (Opsahl and Benner, 1995) and may therefore not be used as an unambiguous source indicator. We observed that C/V values strongly decreased across the Laptev Sea Shelf from ~0.5 (close to the Lena River outlet) to ~0.1 (on the slope/rise, Fig. 5B), which may reflect the preferential degradation of soft tissues. This trend is not likely caused by hydrodynamic sorting, since typically the larger, low-density, woody plant fragments are retained in shallower water, whereas finer material is transported further across the shelf (e.g. Keil et al., 1994; Tesi et al., 2016).

C/V ratios in Buor-Khay Bay sediments (Winterfeld et al., 2015a) in shallow waters were on average lower and more homogeneous (0.17 ± 0.03) than those measured in this study (0.41 ± 0.12 for the corresponding depth interval) (Fig. S1 for comparisons with other studies). C/V values for the entire ESAS were on average 0.15 (0.14 ± 0.07 for only Lena dominated waters) with no water depth trend (Tesi et al., 2014). In shallow sediment cores from the East Siberian Sea, Bröder et al. (2016) measured C/V ratios of 0.20 (inner shelf) and 0.13 (outer shelf) for the surface sediments with no significant trend over sediment depth. For the Mackenzie Shelf C/V values ranged between 0.16 and 0.32 and slightly increased with increasing water depth (Goñi et al., 2000). In contrast, in the Bering Sea, Chuckchi Sea,
Barrow Canyon, Canadian Archipelago, Lancaster sound and Davis Strait there were no C/V trends observed (Goni et al., 2013), with lower values in the Canadian part (0.10 ± 0.12) and highest values on the Beaufort Sea slope, where values slightly decreased with increasing depth (0.39 ± 0.07).

A comparison to the S/V-C/V signatures of potential Arctic plant end-members (compiled by Amon et al., 2012, and citations therein, Tesi et al., 2014, and Winterfeld et al., 2015a) showed that lignin phenols likely derive from both angio- and gymnosperm soft tissues in the shallower samples, closely matching with willow (Salix) tissues measured by Winterfeld et al. (2015a). With increasing water depths, angiosperm wood became the most important source material, while gymnosperm wood, grasses and mosses did not appear to contribute significantly to the overall lignin phenol fingerprint (Fig. 5C). As discussed earlier, this trend may well be a result of preferential degradation and sorting during cross-shelf transport and not derive from actual changes in source material.

3.3 Degradation status of organic matter

During degradation, syringyl and vanillyl phenol aldehydes are oxidized to carboxylic acids of the same phenol group. Increasing Sd/Si and Vd/Vl ratios can therefore qualitatively indicate ongoing degradation of lignin phenols (Ertel and Hedges, 1984; Hedges et al., 1988). For fresh plant material typical acid-to-aldehyde ratios are around 0.1-0.2 (Hedges et al., 1988). Winterfeld et al. (2015a), however, found values as high as Sd/Si = 0.80 and Vd/Vl = 0.67 for a moss species (Aulacomnium turgidum), Sd/Si = 0.87 for larch (Larix) needles and Sd/Si = 0.49 Vd/Vl = 0.41 for wild rosemary (Ledum palustre). Sedges (Carex spp.), dwarf birch (Betula nana) and willow (Salix) range between Sd/Si = 0.13-0.24 and Vd/Vl = 0.18-0.23. The ratio of CuO oxidation-derived 3,5-dihydroxybenzoic acid to vanillyl phenols (3,5-Bd/V) also serves as a proxy for degradation as 3,5-Bd is formed during humification likely occurring in soils (Gordon and Goñi, 2004; Hedges et al., 1988; Prahl et al., 1994; Tesi et al., 2014). For this reason, this proxy can trace mineral rich soil organic matter in contrast to
vascular plant debris (e.g. Dickens et al., 2007; Prahl et al., 1994) as well as degradation during cross shelf transport (Tesi et al., 2016). Sd/Sl, Vd/Vl and 3,5-Bd/V all increased along the transect, implying more degraded material with increasing residence time in the shelf system (Fig. 6A). There appeared to be no differences between outer shelf/slope and rise, which may indicate that TerrOC on the slope is already highly reworked. In contrast, Tesi et al. (2014) found no correlation between Sd/Sl or Vd/Vl and distance from the coast, while 3,5-Bd/V significantly increased with increasing distance from the coast (Fig. S2 for comparisons with other studies). Sd/Sl values for the Buor-Khaya Bay from Winterfeld et al. (2015a) were slightly higher (1.04 ± 0.24) than samples from the corresponding water depths in this study (0.66 ± 0.15), whereas Vd/Vl values were significantly higher (1.28 ± 0.30 compared to 0.59 ± 0.14). Measurements for the Mackenzie Shelf agreed with the ones in this study (Sd/Sl = 0.81 ± 0.25 compared to 1.01 ± 0.33 for the corresponding water depths; Vd/Vl = 0.69 ± 0.14 to 0.86 ± 0.26; 3,5-Bd/V = 0.19 ± 0.04 to 0.31 ± 0.15), but did not show a trend with water depth (Goñi et al., 2000).

Tesi et al. (2016) observed lower acid/aldehyde ratios for the lignin-rich low-density fraction compared to the other fractions (high-density with different grain sizes and settling velocities) in coastal surface sediments from the ESAS. With increasing distance from the coast, these values increased, whereas for the other fractions there were no apparent trends. These findings were interpreted as relatively fresh lignin in the low-density fraction (rich in large plant fragments) compared to the relatively degraded lignin that had likely experienced leaching and adsorbed to the fine mineral fractions (i.e. mineral bound OC). Tesi et al. (2016) found no difference in acid/aldehyde ratios between different density, grain size or settling velocity fractions of surface sediments from three (shorter) transects on the ESAS; it can thus be assumed that these proxies are not affected by hydrodynamic sorting during transport. Degradation-caused changes were limited to the lignin-rich low-density fraction, where Vd/Vl and Sd/Sl increased with increasing distance from the coast. Bulk 3,5-Bd/V values are potentially affected by both sorting and degradation, as they increased with
decreasing particle size (fine and ultrafine fractions had the most degraded signal and are preferentially transported to the outer shelf) and across the shelf in each of the fractions. The carbon preference indices for HMW n-alkanes and HMW n-alkanoic acids have also been widely applied as degradation proxies for plant waxes in marine sediments (for the ESAS, e.g. van Dongen et al., 2008; Fahl and Stein, 1997; Fernandes and Sicre, 2000; Vonk et al., 2010). It measures the ratio of odd-to-even numbers of carbon chain-lengths of HMW lipids and is based on the preference of odd carbon chain-lengths for HMW n-alkanes in fresh plant material (even carbon chain-lengths for HMW n-alkanoic acids; Eglinton and Hamilton, 1967). With ongoing degradation this preference is lost and the CPI approaches 1 (Bray and Evans, 1961). We observed that the HMW n-alkane CPI presented the same a similar pattern as the lignin phenol based degradation indices, however, the HMW n-alkanoic acid CPI did not show as much of a degradation trend (HMW n-alkane CPI: ~5.7 close to the coast, ~2.2 for the deep stations; HMW n-alkanoic acids: ~5.4 close to the coast, ~4.1 for the deep stations; Fig. 6B). Karlsson et al. (2011) measured lipid CPIs in the Buor-Khaya Bay with 10-80 km distance to the coast and obtained similar results to this ~800 km cross-shelf study, with higher values closer to the river delta (Fig. S2 for comparisons with other studies). Their data appears to have a wider spread, though, which might be due to either the narrower dynamics range, or a different definition of high-molecular weight: in this study, carbon chain-lengths of ≥ 23 for n-alkanes and ≥ 24 for n-alkanoic acids were defined as HMW, whereas Karlsson et al. (2011) used ≥ 21 for both compound classes. Fahl and Stein (1997) also reported a large range of n-alkane CPI values (< 0.2- > 5) for Laptev Sea sediments. Fernandes and Sicre (2000) analyzed sediments from the Kara Sea and from the major rivers discharging into this sea, Ob and Yenisey rivers. In the marine environment and the Ob River, they observed HMW n-alkane CPI values between 4.8 and 5.3, similar to those found at shallow water depths in this study. For the Yenisey River and mixing zone, they found higher CPI values, pointing to fresher material being transported there. Vonk et al. (2010) recorded HMW n-alkane CPI values for sediments along the East Siberian Sea Kolyma paleoriver transect.
(across the East Siberian Sea) shelf that decreased from > 7.5 to < 4.0 with increasing distance from the river mouth, overall higher than in this study but confirming the general trend to more degraded material on the outer shelf. Tesi et al. (2016) found HMW \( n \)-alkanoic acid CPI values to decrease with decreasing particle size with no significant trends across the shelf in all but the low-density fraction, which is largely retained close to the shore. The HMW \( n \)-alkane CPI values in that study, however, showed no systematical differences between different fractions, but an overall decreasing trend with increasing distance from the coast.

When undergoing degradation, HMW \( n \)-alkanoic acids may also lose their functional groups, turning them into HMW \( n \)-alkanes (Meyers and Ishiwatari, 1993). The slightly decreasing ratio of HMW \( n \)-alkanoic acids to \( n \)-alkanes also hints at more degraded material with increasing water depth, although, due to a rather large variability, this trend is not significant. For the Buor-Khaya Bay surface sediments Karlsson et al. (2011) obtained similar results (0.48-10.7, here 1.1-10.9) with higher values closer to the river delta (Fig. S2 for comparisons with other studies). Along the Kolyma paleoriver transect, Vonk et al. (2010) measured HMW \( n \)-alkanoic acid to \( n \)-alkane ratios between 1 and 6 with no clear trend with increasing distance from the river mouth. Tesi et al. (2016) found decreasing values with increasing distance from the coast with no differences between the fractions. Two sediment cores from inner and outer East Siberian Sea recording about one century of sedimentation showed no clear trend in CPI or HMW \( n \)-alkanoic acid/\( n \)-alkane towards more degraded TerrOC with increasing sediment depth (Bröder et al., 2016), but displayed a similar difference between inner and outer shelf as seen in this study. This contrasting behavior for cross-shelf and down-core trends may be caused by significantly different timescales for the two processes: about one century in situ/after burial compared to potentially several millennia long lateral transport. Furthermore, the degradation efficiency is possibly-likely higher under the oxic conditions prevailing during cross-shelf lateral transport (Keil et al., 2004), than in the anoxic conditions that predominate below a few millimeters of sediments on the ESAS
Comparing in situ to transport-related oxygen exposure 624 times on the wide Arctic shelves could potentially resolve the observed discrepancies.
4 Concluding remarks and future research directions

Across the Laptev Sea from the Lena River mouth to the deep sea of the Arctic interior a considerable loss of terrigenous organic matter has been observed on both bulk and molecular level. All terrigenous biomarkers display a massive decline with increasing water depth along this high-resolution transect due to hydrodynamic sorting and degradation during transport. Terrigenous organic matter (TerrOC) seems to be also qualitatively more degraded on the outer shelf, slope and rise compared to inner shelf and coastal areas. These results corroborate and expand previous findings for the East Siberian Arctic Shelf, showing that the shelf seas in this region function as an active reactor for TerrOC. Since the East Siberian Arctic Shelf belongs to the widest and shallowest continental margins on Earth, cross-shelf transport times and thus the time spent in oxic sediments are expected to be comparatively long. This stands in sharp contrast to e.g. the Mackenzie basin, which is thought to act as a geological sink for organic carbon due to its efficient TerrOC burial (Hilton et al., 2015). For narrower Arctic shelves in general, where transport times can be expected to be much shorter, organic matter transfer towards the deeper basins appears to be much more efficient, with high TerrOC concentrations in surface sediments even at greater water depths (e.g. Barrow Canyon, Goni et al., 2013). It can therefore be assumed that the cross-shelf transport time exerts first-order control over the extent of TerrOC degradation. With ongoing global warming, rising permafrost-derived organic carbon input from river-sediment discharge and coastal erosion is expected to reach the marine environment. It is therefore crucial to better constrain cross-shelf transport times in order to determine a TerrOC degradation rate and thereby contribute to quantifying potential carbon-climate feedbacks.

Acknowledgements

We thank crew and personnel of the IB ODEN, the RV Yakob Smirnitskyi and the TB0012. The SWERUS-C3 and the International Siberian Shelf Study 2008 (ISSS-08) expeditions were supported by the Knut and Alice Wallenberg Foundation, Headquarters of the Far
Eastern Branch of the Russian Academy of Sciences, the Swedish Research Council (VR Contract No. 621-2004-4039, 621-2007-4631 and 621-2013-5297), the US National Oceanic and Atmospheric Administration (OAR Climate Program Office, NA08OAR4600758/Siberian Shelf Study), the Russian Foundation of Basic Research RFFI (08-05-13572, 08-05-00191-a, and 07-05-00050a), the Swedish Polar Research Secretariat, the Nordic Council of Ministers and the US National Science Foundation (OPP ARC 0909546). L. Bröder also acknowledges financial support from the Climate Research School of the Bolin Climate Research Centre. T. Tesi also acknowledges EU financial support as a Marie Curie fellow (contract no. PIEF-GA-2011-300259), contribution no. XXXX of ISMAR-CNR Sede di Bologna. J.A. Salvadó also acknowledges EU financial support as a Marie Curie grant (FP7-PEOPLE-2012-IEF; project 328049). I. Semiletov thanks the Russian Government for financial support (mega-grant #14.Z50.31.0012). O. Dudarev thanks the Russian Science Foundation (grant No. 15-17-20032).
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Figure 1: Map of the study area in the Laptev Sea. Red filled circles mark the sediment sampling sites. The transect reaches from the Lena River mouth and the Buor-Khaya Bay (water depths ~10 m) across the Laptev Sea Shelf (mean depth ~50 m) to the slope/shelf break and rise (water depths ~3000 m). Arrows show the directions of the prevailing ocean currents.
Figure 2: (A) The ratio of total organic carbon (TOC) to mineral surface area (SA) decreases with increasing water depth by a factor of 7 or 86%. Typical values for deep sea, river-suspended sediments and high TOC supply are taken from Blair and Aller (2012). (B) The stable carbon isotopic signal ($\delta^{13}C$, open boxes) increases with increasing water depth from −26‰ to −22‰, suggesting a change in source material from terrigenous to marine dominated. The radiocarbon isotopic signal ($\Delta^{14}C$, filled diamonds) increases toward the outer shelf, supporting an increase in fresh marine organic carbon. The slope sediments show an older (more depleted) $\Delta^{14}C$ signal, possibly due to ageing during transport and in situ. (C) The relationship between TOC/SA and $\delta^{13}C$ can help to disentangle two processes occurring simultaneously during cross-shelf transport: The net loss (i.e. degradation) or sorting (i.e. hydraulically retaining) of TerrOC leads to a shift towards lower TOC/SA ratios, whereas the replacement/dilution with marine OC shifts the isotopic signature towards higher values.
Figure 3: Terrigenous biomarker loadings display a strong decrease with increasing water depth across the shelf: (A) for lignin phenols and cutin acids by a factor of 130 and 230 respectively, (B) for HMW $n$-alkanes and HMW $n$-alkanoic acids by a factor of 12 and 44 respectively. (C) Comparison between the different biomarkers along the transect: lignin phenols, cutin acids, HMW $n$-alkanoic acids and $n$-alkanes where each is normalized to respective highest value (corresponding to 100%).
Figure 4: A comparison of lignin phenol data from this project to values from published studies around the Arctic Ocean. Similar decreasing trends with increasing water depth are observed for all systems but Barrow Canyon, where elevated lignin phenols concentrations are found even at depth of > 1000 m.
Figure 5.4: The lignin phenol composition carries source information: (A) an increasing ratio of syringyl to vanillyl phenols (S/V) suggests relatively more angiosperm material on the outer shelf/slope possibly due to selective degradation or sorting during transport. (B) The decreasing ratio of cinnamyl to vanillyl phenols (C/V) decreases with increasing water depths, which implies an increasing relative contribution of woody material compared to soft tissues. (C) Comparison of S/V and C/V with the end-members for different Arctic plants as compiled from different studies by Amon et al. (2012, and citations therein, here marked with 1); ice-complex deposit and topsoil permafrost as determined by Tesi et al. (2014, here marked with 2) and more plant species measured by Winterfeld et al. (2015a, here marked with 3). The boxes indicate typical ranges of S/V and C/V for different vascular plant tissues in different locations (e.g. Goñi et al., 2000). The surface sediment lignin phenol compositions along the Laptev transect appear to be a mix of angio- and gymnosperm soft tissues most similar to willow and shift towards angiosperm wood with increasing water.
Figure 86: Degradation proxies for TerrOC, blue triangles point toward lower extent of degradation: (A) CuO-oxidation derived ratios Sd/Si, Vd/Vl and 3,5-Bd/V all display a trend toward more degraded material with increasing distance from the shore with no difference between outer shelf and slope/rise sediments. (B) The carbon preference indices (CPI) of HMW n-alkanes and n-alkanoic acids show the same trend, yet not as pronounced for the n-alkanoic acids. The ratio of HMW n-alkanoic acids to HMW n-alkanes has a wider scatter, but also hints at more degraded material with increasing water depth.
Table 1: List of surface sediment samples from the Laptev Sea transect.

<table>
<thead>
<tr>
<th>ID</th>
<th>Sample type</th>
<th>Lat °N</th>
<th>Long °E</th>
<th>Water depth m</th>
<th>OC mg·g⁻¹</th>
<th>SA m²·g⁻¹</th>
<th>δ¹³C %</th>
<th>Δ¹⁴C %</th>
<th>SiO₂ wt %</th>
<th>Al₂O₃ wt %</th>
<th>CaO wt %</th>
</tr>
</thead>
<tbody>
<tr>
<td>SW-1</td>
<td>0-0.5cm</td>
<td>78.942</td>
<td>125.243</td>
<td>3146 10.4</td>
<td>34.0</td>
<td>-22.34</td>
<td>-418</td>
<td>60.3</td>
<td>16.5</td>
<td>2.4</td>
<td></td>
</tr>
<tr>
<td>SW-2</td>
<td>0-0.5cm</td>
<td>78.581</td>
<td>125.607</td>
<td>2900 13.8</td>
<td>38.3</td>
<td>-22.70</td>
<td>-392</td>
<td>57.8</td>
<td>17.2</td>
<td>2.1</td>
<td></td>
</tr>
<tr>
<td>SW-3</td>
<td>0-0.5cm</td>
<td>78.238</td>
<td>126.150</td>
<td>2601 10.6</td>
<td>31.8</td>
<td>-22.61</td>
<td>-426</td>
<td>62.1</td>
<td>16.0</td>
<td>1.6</td>
<td></td>
</tr>
<tr>
<td>SW-4</td>
<td>0-0.5cm</td>
<td>77.855</td>
<td>126.664</td>
<td>2106 13.2</td>
<td>41.5</td>
<td>-22.51</td>
<td>-428</td>
<td>56.6</td>
<td>17.5</td>
<td>1.3</td>
<td></td>
</tr>
<tr>
<td>SW-14</td>
<td>0-1cm</td>
<td>76.894</td>
<td>127.798</td>
<td>64 8.9</td>
<td>19.4</td>
<td>-24.33</td>
<td>-314</td>
<td>71.3</td>
<td>12.5</td>
<td>1.5</td>
<td></td>
</tr>
<tr>
<td>SW-23</td>
<td>0-1cm</td>
<td>76.171</td>
<td>129.333</td>
<td>56 15.8</td>
<td>21.7</td>
<td>-24.96</td>
<td>-333</td>
<td>68.9</td>
<td>13.6</td>
<td>1.4</td>
<td></td>
</tr>
<tr>
<td>YS-4</td>
<td>0-1cm</td>
<td>75.987</td>
<td>129.984</td>
<td>50 13.4³</td>
<td>31.4</td>
<td>-24.76⁵</td>
<td>-284 ²</td>
<td>63.8</td>
<td>15.1</td>
<td>1.3</td>
<td></td>
</tr>
<tr>
<td>SW-24</td>
<td>0-1cm</td>
<td>75.599</td>
<td>129.558</td>
<td>46 10.7</td>
<td>37.0</td>
<td>-24.30</td>
<td>-437</td>
<td>62.5</td>
<td>15.4</td>
<td>1.2</td>
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<tr>
<td>YS-6</td>
<td>0-1cm</td>
<td>74.724</td>
<td>130.016</td>
<td>32 18.6³</td>
<td>31.6</td>
<td>-25.60⁵</td>
<td>-465 ²</td>
<td>62.1</td>
<td>16.1</td>
<td>1.3</td>
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</tr>
<tr>
<td>YS-9</td>
<td>Grab</td>
<td>73.366</td>
<td>129.997</td>
<td>23 13.1³</td>
<td>16.9</td>
<td>-26.10⁵</td>
<td>-415 ²</td>
<td>70.8</td>
<td>14.0</td>
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<td>YS-13</td>
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<td>131.701</td>
<td>19 18.9³</td>
<td>23.5</td>
<td>-25.90⁵</td>
<td>-543 ²</td>
<td>61.6</td>
<td>17.4</td>
<td>0.8</td>
<td></td>
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<td>YS-14</td>
<td>0-1cm</td>
<td>71.630</td>
<td>130.050</td>
<td>7 19.1³</td>
<td>11.4</td>
<td>-26.20⁵</td>
<td>-504 ²</td>
<td>69.6</td>
<td>15.0</td>
<td>1.6</td>
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<tr>
<td>TB-46</td>
<td>Grab</td>
<td>72.700</td>
<td>130.180</td>
<td>✅ 25.8³</td>
<td>12.0⁵</td>
<td>-26.50⁵</td>
<td>-436 ²</td>
<td>67.9</td>
<td>15.2</td>
<td>1.8</td>
<td></td>
</tr>
</tbody>
</table>

Data from: ³Vonk et al. (2012); ⁵Tesi et al. (2016); ²Karlsson et al. (2014).
Table 2: Biomarker results for surface sediment samples from the Laptev Sea transect.

<table>
<thead>
<tr>
<th>ID</th>
<th>Lignin µg m⁻²</th>
<th>Cutin µg m⁻²</th>
<th>HMW* alkanes µg m⁻²</th>
<th>HMW** acides µg m⁻²</th>
<th>S/V</th>
<th>C/V</th>
<th>Sd/Si</th>
<th>Vd/Vl</th>
<th>3,5Bd/V</th>
<th>CPI alk</th>
<th>CPI acids/alk</th>
</tr>
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<tr>
<td>SW-1</td>
<td>0.56</td>
<td>0.063</td>
<td>0.14</td>
<td>0.27</td>
<td>1.1</td>
<td>0.09</td>
<td>1.3</td>
<td>0.98</td>
<td>0.43</td>
<td>2.5</td>
<td>4.1</td>
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<td>SW-2</td>
<td>0.41</td>
<td>0.070</td>
<td>0.13</td>
<td>0.57</td>
<td>0.99</td>
<td>0.16</td>
<td>1.3</td>
<td>0.99</td>
<td>0.52</td>
<td>1.9</td>
<td>4.1</td>
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<td>SW-3</td>
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<td>0.75</td>
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<td>0.14</td>
<td>1.3</td>
<td>0.95</td>
<td>0.61</td>
<td>1.7</td>
<td>4.1</td>
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<td>SW-4</td>
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<td>0.093</td>
<td>0.10</td>
<td>1.1</td>
<td>0.08</td>
<td>1.4</td>
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<td>0.58</td>
<td>2.7</td>
<td>4.1</td>
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<td>SW-6</td>
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<td>0.18</td>
<td>0.67</td>
<td>0.90</td>
<td>0.19</td>
<td>1.2</td>
<td>1.0</td>
<td>0.46</td>
<td>2.9</td>
<td>4.4</td>
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<tr>
<td>SW-14</td>
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<td>0.15</td>
<td>0.46</td>
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<td>0.08</td>
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<td>0.18</td>
<td>1.6</td>
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<td>0.19</td>
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<td>1.1</td>
<td>0.43</td>
<td>4.0</td>
<td>3.9</td>
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<tr>
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<td>11</td>
<td>0.83^d</td>
<td>6.6^d</td>
<td>0.64</td>
<td>0.53</td>
<td>0.55</td>
<td>0.47</td>
<td>0.12</td>
<td>5.4^d</td>
<td>5.0^d</td>
</tr>
</tbody>
</table>

* carbon chain-lengths 23-34; ** carbon chain-lengths 24-30.  
^d recalculated data from Karlsson et al. (2011).
Figure S1: The ratios of high-over-low molecular weight n-alkanes and n-alkanoic acids decrease along the transect due to dilution with marine OC.
Figure S1: A comparison of biomarker concentrations and ratios measured in this study to literature values from different studies on Arctic margins: a Tesi et al. (2014), b Winterfeld et al. (2015a), c Goñi et al. (2000), d Goni et al. (2013), e Karlsson et al. (2011), f Vonk et al. (2010), g Belicka et al. (2004), h Yunker et al. (1993).
Figure S2: A comparison of the degradation proxies measured in this study to literature values from different studies on Arctic margins: a Tesi et al. (2014), b Winterfeld et al. (2015a), c Goñi et al. (2000), d Karlsson et al. (2011), e Vonk et al. (2010).