Net primary productivity, allocation pattern and carbon use efficiency in an apple orchard assessed by integrating eddy-covariance, biometric and continuous soil chamber measurements

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Abstract

Carbon use efficiency (CUE) is a functional parameter that could possibly link the current increasingly accurate global estimates of gross primary production with those of net ecosystem exchange, for which global predictors are still unavailable. Nevertheless, CUE estimates are actually available for only a few ecosystem types, while information regarding agro-ecosystems is scarce, in spite of the simplified spatial structure of these ecosystems that facilitates studies on allocation patterns and temporal growth dynamics.

We combined three largely deployed methods, eddy covariance, soil respiration and biometric measurements, to assess monthly values of CUE, net primary production (NPP) and allocation patterns in different plant organs in an apple orchard during a complete year (2010). We applied a measurement protocol optimized for quantifying monthly values of carbon fluxes in this ecosystem type, which allows for a cross-check between estimates obtained from different methods. We also attributed NPP components to standing biomass increments, detritus cycle feeding and lateral exports.

We found that in the apple orchard both net ecosystem production and gross primary production on yearly basis, 380 ± 30 g C m⁻² and 1263 ± 189 g C m⁻² respectively, were of a magnitude comparable to those of natural forests growing in similar climate conditions. The largest differences with respect to forests are in the allocation pattern and in the fate of produced biomass. The carbon sequestered from the atmosphere was largely allocated to production of fruits: 49% of annual NPP was taken away from the ecosystem through apple production. Organic material (leaves, fine root litter, pruned wood and early fruit falls) contributing to the detritus cycle was 46% of the NPP. Only 5% was attributable to standing biomass increment, while this NPP component is generally the largest in forests.

The CUE, with an annual average of 0.71 ± 0.09, was higher than the previously suggested constant values of 0.47–0.50. Low nitrogen investment in fruits, the limited
root-apparatus, and the optimal growth temperature and nutritional condition observed at the site are suggested to be explanatory variables for the high CUE observed.

1 Introduction

Global greenhouse gases (GHGs) concentration in atmosphere has been growing since pre-industrial times due to anthropogenic forcing, in particular fossil fuel combustion and land use change (Canadell et al., 2007; Le Quere et al., 2009), with carbon dioxide (CO₂) being the prevalent anthropogenic GHG (IPCC-AR4, 2007; Peters et al., 2011). By sequestering a large amount of atmospheric carbon (C), terrestrial ecosystems are thought to offer a mitigation strategy for reducing global warming (Schimel et al., 2001). This is confirmed by the observation that the annual increment of atmospheric CO₂ is substantially smaller than the increment in anthropogenic emissions and, on a global scale, it has been estimated that the terrestrial biosphere is able to take up about 30% of anthropogenic CO₂ emissions annually (Schulze, 2006; Canadell et al., 2007).

Several studies have been carried out to assess the capacity of ecosystems in different natural biomes to sequester C from the atmosphere, most of which related to FLUXNET synthesis activity (Baldocchi, 2008; http://www.fluxnet.ornl.gov/) and results have been recently reviewed in a global dataset (Luyssaert et al., 2007; Schulze et al., 2010). An effective way to compare the ability of different ecosystems to sequester CO₂ from the atmosphere is to determine their carbon use efficiency (CUE). CUE, the ratio of net primary productivity (NPP) to gross primary productivity (GPP), is an intuitive and easily comparable index by which to assess the capacity of an ecosystem to transfer C from the atmosphere to terrestrial biomass (DeLucia et al., 2007). Increasing our knowledge on the magnitudes and spatial distribution of CUE and heterotrophic respiration (Rₘ) could allow for a better linkage of the GPP estimates with those of net ecosystem productivity (NEP), for which reliable climatic and biological predictors are still unavailable at the global scale.
Despite the great effort being made on the study of natural ecosystems, less knowledge is available from this ecological point of view regarding intensively managed ecosystems, particularly woody agro-ecosystems, although some crop classes may assume great importance especially at regional scale (Testi et al., 2008). Agricultural practices such as soil tillage, fertilization, irrigation, pruning and the reduced biodiversity occurring in non-natural ecosystems, may significantly alter the capacity of ecosystems to exchange C with the atmosphere (Smith, 2004; Osborne et al., 2010) and thus their potential to act as a sink of C as compared to natural ecosystems growing in similar environmental conditions.

Woody agro-ecosystems are among the least quantified and most uncertain elements in the terrestrial biogeochemical cycle. In the present study we hypothesize that the main ecosystem carbon fluxes of a woody agro-ecosystem are of the same magnitudes with respect to those of a natural forested ecosystem of the same biome rank (temperate-humid deciduous forest), while the main differences between the two land use types take place in the allocation pattern of fixed C within tree organs.

We investigated an apple orchard (*Malus domestica* Borkh.) growing in a temperate-humid area and compared it with data taken from literature of temperate-humid forests (Curtis et al., 2002; Luyssaert et al., 2007; DeLucia et al., 2007). We used CUE as a comparison index and we biometrically measured the NPP of the main ecosystem compartments to assess the C allocation pattern in the year 2010. In order to test the robustness of the measurements of C fluxes involved in CUE determination (GPP, NPP and autotrophic respiration, R_a), we adopted an experimental protocol which allowed us to obtain a cross check between independent estimates of each flux.
2 Materials and methods

2.1 Site description

The study site is located in the intensively cultivated valley bottom of the Adige River, in the municipality of Caldaro, South Tyrol, Italy (46°21’ N, 11°16’ E; 240 m a.s.l.). Apple trees (*Malus domestica* var. Fuji grafted on dwarfing M9 rootstock) were planted in rows in the year 2000, following a regular frame of 3 × 1 m, where 1 m is the distance between plants along the row and 3 m is the distance between tree rows. The average pruned tree height was 3.6 m. A 1.2 m-wide strip of soil centered on the tree-row was kept free of grass through periodic tillage. In the 1.8 m wide alleys between tree rows, grasses were free to grow, and were cut 3 times during the year. Apple tree plantations with the same characteristics were present around the selected field for a minimum distance of 300 m in all directions.

Budburst occurred in the second half of March, trees maximum LAI was 2.8 m² m⁻² in July and major leaf fall started at the end of October. The 30-yr average mean annual temperature was 11.5 °C, while the mean annual temperature during 2010 was 11.6 °C. Total water input for 2010 was 1770 mm, of which 1050 mm come from precipitation and 720 mm from irrigation. The soil is a Calcaric Cambisol according to the FAO Soil Taxonomy, with a pH of 7.4. In the upper 20 cm the soil bulk density was 1.49, the organic carbon concentration was 1.74 % and the nitrogen concentration was 0.20 %. The orchard was managed according to the guidelines of organic tree production. In 2010, 35.5 gCm⁻² and 7.5 gNm⁻² were applied by organic fertilizers.

2.2 Experimental set up

The site was selected based on the favorable conditions for eddy-covariance (EC) measurement in terms of regular terrain and homogeneity of land surface cover. An 8-m tower was set up at the beginning of 2009. Instruments for EC measurements were installed at the top of this tower. Additionally, the tower was equipped with a series
of meteorological instruments. Solar radiation components were measured by CNR1, Kipp & Zonen, Delft, Holland; air temperature and relative humidity by CS215, Campbell Scientific Incorporated, Logan, Utah, United States (CSI hereafter), rainfall by a professional rain gauge (RAIN-O-MATIC, Pronamic, Silkeborg, Denmark) and soil water content by multiple TDRs (CS616, CSI). All meteorological data were logged by a CR3000 (CSI).

Close to the tower, 16 collars (20 cm diameter) for soil respiration measurement were placed along a selected row at 35–55 cm from the trees, 8 on control plots and 8 on trenching plots. Soil trenching (50 × 50 × 60 cm) was carried out in summer 2009. Practical limitations for expanding the survey area to other tree lines and grassed alleys were encountered, since the measurement chambers would have been an obstacle to the farm machinery. This limitation was overcome by carrying out a parallel independent campaign for assessing the spatial variability of soil respiration in the field.

After a characterization of tree diameters that was conducted over the whole site, six groups of five plants each were selected in order to represent the probability density distribution of the observed tree diameter. Nets for litter collection were placed under selected trees and biometric measurements were carried out during the 2010 growing season to assess the total NPP and carbon allocation within the studied ecosystem. Along with measurements and litter collection, 9 branches (3 per plant level) were cut from randomly selected trees and brought to the laboratory for analysis at each sampling date. The collected material was used to determine: the mean dry weight of each organ (after drying in oven to a constant weight at 65°C), the mean carbon and nitrogen content of each NPP component (FlashEA™ 1112 Elemental Analyzer, Thermo Fisher Scientific, Germany) and the mean leaf surface (LI-3000 + LI-3050 Portable Area Meters, Li-Cor Biosciences, Lincoln, Nebraska, USA, Li-Cor hereafter).
2.3 Measurement techniques

2.3.1 Eddy-covariance measurements

Net Ecosystem Exchange (NEE) of CO₂ was measured continuously by the eddy covariance technique since March 2009. Measurements and calculation were performed following the Euroflux methodology as described by Aubinet et al. (2000) with a 3-D sonic anemometer (Gill R3-50, Gill-Instruments, Lymington, UK) at a height of 8 m above ground (4 m above the canopy) and a close path CO₂/H₂O infrared gas analyzer (IRGA, LI-7000, Li-Cor). Air was sampled through a polyethylene tube (4 mm inner, 6 mm external diameter) at a distance of 0.3 m from the anemometer with a flux rate of 10 l min⁻¹ provided by an external pump (N838 KNDC, KNF Nueberger GmbH, Freiburg, Germany). Calibration was performed bi-weekly with reference gases: nitrogen and 380 ppm CO₂ flasks (produced by Messer, Grugliasco, Italy) were used to set the zero and the CO₂ span, respectively. Zero-level of CO₂ and H₂O in the reference cell of the analyzer was assured by the use of chemicals (respectively Ascarite II for CO₂ and magnesium perchlorate for water vapor), which were substituted bi-weekly.

The software Eddysoft (Kolle and Rebmann, 2007; Mauder et al., 2008) was used to calculate eddy fluxes with the following criteria: no detrending, no high or low pass filtering corrections were used; a two-axis rotation of coordinates was applied each 30 min. The software automatically calculated the lag-time for CO₂ at each half-hour to maximize the covariance between fluctuations in vertical wind velocity and gas molar fraction. In addition, the analysis of stationarity conditions for CO₂ turbulent flux and of the Integral Turbulent Characteristic (ITC) following Foken and Wichura (1996) was performed.

Gaps in CO₂ flux time series were filled with look-up tables (LUT) based on meteorological seasonal conditions. The whole year was separated into six bi-monthly periods and for each period two different tables were compiled: one for nighttime and one for daytime. For both tables only quality checked data were used as input. Average flux for the nighttime table was calculated using a moving LUT with 15-day sliding windows,
compiled for 21 classes of relative soil water content and 26 classes of air temperature. The ranging interval of a meteorological variable in a specific class depended on the ratio between the overall range of the variable in the bimonthly period and the number of classes. Gaps in the fifteen central days of the LUT were filled and then the temporal window of the LUT was moved 15 days ahead. For the daytime the moving LUT was compiled for 26 classes of air temperature and 26 classes of global radiation (Rg). Rg was also used to distinguish day from night (Rg > 20 W m\(^{-2}\)).

The partitioning of the observed NEE into gross primary productivity (GPP) and ecosystem respiration (R\(_{\text{eco}}\)) was achieved through night time LUT used for gap filling. Indeed, day time R\(_{\text{eco}}\) values for a bimonthly period were extrapolated from the nocturnal LUT according to air temperature and soil humidity for the specific day time half-hour period.

As a form of control, gap-filling was also performed by the marginal distribution sampling method which accounts for temporal auto-correlation of fluxes, replacing missing data with the average value under similar meteorological conditions (Reichstein et al., 2005) and by the light response curve approach described by Lasslop and colleagues (2010), using the online standard tool (http://www.bgc-jena.mpg.de/bgc-mdi/html/eddyproc/index.html). GPP and R\(_{\text{eco}}\) calculation was performed using the same tool.

### 2.3.2 Biometric measurements

Biometric measurements were conducted over six representative plots following Law et al. (2008). Six NPP components were considered separately: leaves (NPP\(_l\)); fruits (NPP\(_f\)); aboveground woody tissues (NPP\(_{\text{wAG}}\)) which include trunk, branches and shoots; belowground woody tissues (NPP\(_{\text{wBG}}\)) which include coarse roots and the belowground part of trunk; fine roots (NPP\(_{\text{fr}}\)); understory production (NPP\(_u\)). Each of these fluxes was assessed directly and independently. We did not consider in this budget the volatile organic compounds (VOC), non-\(\text{CO}_2\) carbon emission and root exudates production. Data and samples were collected once a month and the following
The equation was used to calculate the total NPP produced within each sampling date and for the whole season:

$$\Delta \text{NPP} = (L_{t+1} + \text{Sb}_{t+1}) - (\text{Sb}_t)$$  \hspace{1cm} (1)

Where $L$ is the litter collected from the nets and Sb is the standing biomass. Using this procedure, shrinkage effects in tree diameter may determine apparent reduction in wood biomass. Variations in C storage within tree organs were not considered beside their relevance. Details on the sampling procedures for each NPP component are described below.

NPP$_l$ – one tree out of each plot for a total of 6 trees was selected. In April 2010, we divided the trees according to 3 height levels (low = 0–120 cm, medium = 120–240 cm, high = 240–360 cm) to represent vertical variability within the trees (e.g. Rayment et al., 2002). We also numbered and tagged all the branches and we counted the number of leaves and flowers on each branch.

Three branches per tree level were then chosen, and from May till November the number of leaves and fruits of these 9 branches per tree were monitored avoiding any sampling collection. We used the complete characterization carried out in April to determine a multiplicative factor, specific to each plant level, to upscale measured values to the whole tree. The derived total number of leaves per tree was multiplied by the mean dry weight of leaves of that period, and thus by leaf carbon concentration, to determine the total amount of C in the leaves.

Leaf abscission was monitored by collecting the litter from nets placed under each selected plot on the same sampling date. NPP$_l$ was thus calculated using Eq. (1).

NPP$_f$ – once a month, flowers (April) and fruits (from May till October) were counted on the selected branches. The total number of fruits per plant was counted at harvest (October) and a multiplicative factor was used to extrapolate fruit number reduction due to early drop from selected branches to the whole tree, thus obtaining an estimate of total number of fruits per tree throughout the season. In a similar way as for the leaves, in order to assess C allocation to fruits, the total number of fruits was multiplied by the
mean dry weight of fruit at each sampling date and by the mean C content. Equation (1) was applied to assess NPP_f.

NPP_{w_{AG}} – monthly records of tree circumferences at 10 cm above grafting points were collected for each tree of the selected plots (n = 30). Allometric (\(y = ax^b\)) equation parameters were determined by excavating 11 apple trees of the same age and size, that were grown in very similar environmental and soil conditions in a nearby orchard. Since apple trees are pruned each year during winter, the same plant diameter may lead to a significant difference in aboveground (AG) woody biomass estimate depending on whether allometric equations are applied to pruned or unpruned trees. To consider this effect, we built first allometric equation with pruned trees (wood_{AG,p}) and a second equation with unpruned trees (wood_{AG,np}, see respective parameters in Table 1). Based on our measurements, pruning material was quantified in 11.5 % of AG woody biomass. To calculate the NPP_{w_{AG}} at monthly time-steps, a third allometric equation (wood_{AG}, see Table 1 for equation parameters) was determined with an initial value (April) on the ordinate set by the equation for pruned trees and the final value (November) on the ordinate set by equation for unpruned trees.

\[
NPP_{w_{AG}} = a(x^{b_{t+1}} - x^{b_t})
\]

Where \(x\) is the diameter at 10 cm above the grafting point, \(a\) and \(b\) are parameters of the appropriate power equation and \(t\) is the time of biometric measurement. When considering wood production over more than one year, the overall procedure results in a saw-tooth diagram.

NPP_{w_{BG}} – on the same excavated trees, the belowground (BG) biomass was determined to a horizontal distance of 15 cm, and to a vertical depth of 1 m. This value was integrated in space by considering the coarse roots excavated through soil coring (see below). Coarse roots where considered if they measured > 2 mm in diameter. Spatial interpolation was performed by ordinary kriging, assuming a maximum root depth of 1 m. A power allometric equation was established to relate diameter and belowground
woody organs (wood\textsubscript{BG}). The following equation was then used to estimate NPP\textsubscript{w\textsubscript{BG}}:

\[
\text{NPP}_{w_{BG}} = c(x_{t+1}^d - x_t^d)
\] (3)

Where \(x\) is the diameter at 10 cm above the grafting point and \(c\) and \(d\) are the parameters of the best fitting power equation, reported for wood\textsubscript{BG} in Table 1.

We assumed that all coarse root growth accounted for standing biomass increase, so not contributing to detritus cycle.

NPP\textsubscript{fr} – in March 2010, an intensive soil sampling campaign was carried out in order to assess the mean root biomass and its distribution. One tree per plot was selected and 17 soil cores were taken at each of the six plots, along two parallel lines across tree rows at different distances (> 15 cm) from the tree trunks. Each soil core was divided into 3 depth levels: 0–20, 20–40 and 40–60 cm. Each soil sample \((n = 306)\) was sieved to extract roots, separating them into coarse (diameter > 2 mm) and fine (< 2 mm) roots. Interpolation in space of fine root density values was performed as for coarse roots.

In summer 2009, several minirhizotrons were installed in the apple orchard, at distances of 15, 35, 55 and 150 cm from the tree. They consisted of transparent Plexiglas tubes (8 cm diameter, 1 m length) inserted into the soil on an angle of 45° for approximately 90 cm, thus exploring a soil depth of 60 cm. Starting from 18 March 2010, root growth was monitored by collecting periodic images inside the minirhizotrons using a root scanner (CI-600 Root Scanner, CID-Inc, Camas, WA, USA). After a first screening of the collected images, 8 representative minirhizotrons were considered for the analysis. To assess fine roots NPP, the relative growth rate was calculated by image analysis (WinRHIZO software, Regent Instruments, Canada). The growth coefficient obtained was applied to the value of initial fine root biomass assessed by soil coring. No distinction was made between the growth pattern of grass and tree roots.

We assumed that a constant ratio existed between the standing biomass of fine and coarse roots in the dormant season, and that the excess in fine root production was annually feeding the detritus cycle.
NPP\(u\) – in the grassed alley, close to the monitored trees, we selected 6 control plots of 1.8 m\(^2\) from which we mowed the aboveground grass production on a monthly basis. Grass root growth, observed with minirhizotrons, was considered as NPP\(fr\) component. The herbaceous biomass grown along the tree row in periods between tillage events was assumed to be negligible.

### 2.3.3 Soil respiration measurements

An automatic multichambers CO\(_2\) soil flux measurement system (LI-8100 + LI-8150 with 8 chambers type LI-8100-104, Li-Cor) was used to measure soil respiration (\(R_s\)) and its heterotrophic component (\(R_h\)) along the tree row. Four chambers were kept on the same collars for the whole season, while the other four were rotated over 12 different positions on a weekly basis. Fluxes in each chamber were taken every half hour. Out of the 16 collars installed as described in Sect. 2.2., eight were on control plots (to assess \(R_s\)) and eight were on trenching plots (to assess \(R_h\)). CO\(_2\) concentration values were taken every second for a measurement period of 2 min and 35 s. The first 45 s were considered as a mixing period and excluded from the calculation of the soil CO\(_2\) efflux, which was obtained from the linear regression of the increasing CO\(_2\) concentration within the chamber over 1 min and 50 s.

Overall, more than 26 000 data points for both \(R_s\) and \(R_h\) were recorded. As quality control, we used the correlation coefficient of the linear relation between time and CO\(_2\) concentration, discarding values with \(R^2\) below 0.95, a more restrictive threshold than that 0.90 proposed by Savage et al. (2008). Gap-filling was performed on data collected at each collar following the LUT method, with air temperature and soil humidity as independent variables. Total \(R_s\) and \(R_h\) were calculated by summing the gap-filled time series.

A possible source of systematic error occurring with automated system is linked with the limited spatial representativeness of the point of \(R_s\) measurement (Savage et al., 2008), imposed by the nature of the system and the logistics of the study site. In order to obtain reliable values of \(R_s\), \(R_h\) and \(R_{a,BG}\) (below-ground autotrophic respiration,
calculated as \( R_s - R_h \)), for the whole ecosystem, in June 2010 a parallel soil respiration measurement campaign was carried out in order to: (1) take into account the spatial variability of soil respiration in the tree-lines (the soil strips under the tree canopy, 1.2 m width); (2) estimate the soil respiration in the areas corresponding to the grassed alleys (1.8 m width). Six collars (10 cm diameter) were placed at different distances from the trees (5 in the tree-line and 1 in the grassed alley) in each plot used for biometric measurements (36 collars in total) and 7 measurement cycles of \( R_s \) over one week were performed for each collar by a second LI-8100 with a LI-8100–102 survey chamber. The relations of \( R_s \) with air temperature observed in the collars located in the tree-lines and in the grassed alleys during this campaign were used to upscale continuous measurements to the whole orchard.

### 2.4 \( R_a \) assessment

Autotrophic respiration was not measured directly, but derived in three independent ways from measurements that rely on different methodologies. The first method follows the equation

\[
R_a = GPP - NPP
\]  
(4)

where GPP is derived from EC and NPP is assessed biometrically.

The second method follows the equation

\[
R_a = R_{eco} - R_h
\]  
(5)

where the first right hand term is EC-derived and the latter is measured with the soil chamber system.

The third method to assess \( R_a \) is based on the estimate of the autotrophic component of the soil respiration (\( R_{a,BG} = R_s - R_h \)) and on the approach suggested by Reich et al. (2006), who observed a consistent near–isometric scaling of total and aboveground plant respiration (\( R_{a,AG} \)) to total and aboveground plant N content across different taxa, environments and experiments. We firstly calculated the amount of N present
in the root biomass of the grassed alleys and in the root biomass of the tree-lines; then, we upscaled $R_{a,BG}$ to the whole ecosystem according to the N distribution in plant organs.

By this method,

$$R_a = k(R_{a,BG})$$

(6)

where the coefficient $k$ accounts for the ratio between measured total and belowground N content (Table 2).

### 2.5 NPP, GPP and CUE estimates

NPP was assessed biometrically ($NPP_{biom}$), by adding the cumulated values of each NPP component considered in the biometric measurements, and by summing the yearly NEP and $R_h$ ($NPP_{flux}$), thus involving eddy-covariance and the soil respiration chamber system respectively. The daily carbon uptake rate was calculated by dividing both the cumulated $NPP_{biom}$ and $NPP_{flux}$ at each sampling date by the number of days between the current sampling date to the previous one.

GPP was calculated following two methods. The first annual estimate of it ($GPP_{EC}$) was obtained from flux partitioning of EC-derived NEE fluxes, while the second yearly GPP value ($GPP_{BS}$) was calculated avoiding any involvement of the eddy covariance methodology by summing $NPP_{biom}$ with the $R_a$ assessed using the third method described in Sect. 2.4.

Carbon Use Efficiency (CUE) is defined as the ratio between net (NPP) and gross (GPP) primary production. To assess the uncertainty of CUE estimates, we used different approaches to determine both NPP and GPP, so obtaining four partially independent CUE values.

The seasonal trend of CUE was assessed in two partially independent ways ($CUE_{biom}$ and $CUE_{flux}$), by dividing the $NPP_{biom}$ and $NPP_{flux}$ values at each sampling date by the respective $GPP_{EC}$ amount. The yearly value of CUE was calculated by
dividing $\text{NPP}_{\text{biom}}$ and $\text{NPP}_{\text{flux}}$ by both $\text{GPP}_{\text{EC}}$ and $\text{GPP}_{\text{BS}}$ on an annual basis, thus obtaining four independent estimates of this index.

### 2.6 Statistical analysis and uncertainty estimate

Random and systematic errors affecting biometric, eddy covariance and soil respiration measurements (Clark et al., 2001; Loescher et al., 2006; Savage et al., 2008) contribute to the uncertainty in the estimate of each flux component. The different spatial representativeness of the measurement techniques deployed is an additional source of uncertainty affecting the overall carbon cycle estimate. We evaluated the uncertainty in EC based fluxes estimates by comparing results from different gap-filling and partitioning methods.

Additive and multiplicative random errors in biometric estimates were calculated by means of the error propagation theory (Taylor, 1982). When two means ($X$ and $Y$) with their standard errors of the mean ($\text{SEM}_x$ and $\text{SEM}_y$) were added yielding the value $Z$, the standard error of $Z$ ($\text{SEM}_z$) was calculated as follows

$$\text{SEM}_z = \sqrt{(\text{SEM}_x)^2 + (\text{SEM}_y)^2}$$  \hspace{1cm} (7)

while if $X$ and $Y$ were multiplied, the resulting $\text{SEM}_z$ was calculated as follows

$$\text{SEM}_z = Z \times \sqrt{\frac{(\text{SEM}_x)^2}{X} + \frac{(\text{SEM}_y)^2}{Y}}$$  \hspace{1cm} (8)

### 3 Results

#### 3.1 The ecosystem carbon stocks and fluxes

Soil was found to be by far the highest carbon pool, containing much more carbon than the standing biomass (17.1 vs. 1.2 kgC m$^{-2}$). Data relative to the stocks of carbon
within the standing biomass in the apple orchard at the beginning of the growing season (March 2010) are reported in Table 2: 72% of C was allocated aboveground and 28% belowground, while N was allocated 66% aboveground and 34% belowground.

### 3.1.1 Fluxes from eddy covariance

Figure 1 shows the daily ecosystem carbon fluxes of the apple orchard for the year 2010 assessed via eddy-covariance. Budburst occurred on the 18 March and NEE started to become negative (sink of C) during the first decade of April and returned positive (source of C) in the first decade of November, when leaf abscission was almost complete and only the vegetation on grassed alleys carried out photosynthesis. The maximum rate of NEE was $-7.21 \text{g C m}^{-2} \text{d}^{-1}$ (25 June). On a yearly basis, GPP, $R_{\text{eco}}$, and NEP ($= -\text{NEE}$) accounted for $1263 \pm 189$, $883 \pm 160$, and $380 \pm 30 \text{g C m}^{-2} \text{yr}^{-1}$, respectively, according to the LUT method. Uncertainties were determined as standard error obtained from comparison with other interpolation algorithms for gap filling and flux partitioning. Following Reichstein et al. (2005), NEP was 351, GPP 1074 and $R_{\text{eco}}$ 723 g C m$^{-2}$ yr$^{-1}$. Following Lasslop et al. (2010), these values were 512, 945, and 1433 g C m$^{-2}$ yr$^{-1}$, respectively.

### 3.1.2 Fluxes from soil respiration system

Measured soil respiration ($R_s$) and its heterotrophic component ($R_h$) fluxes over the season are shown in Fig. 2. According to results from the spatial variability campaign, we found that continuous chamber measurements were taken in a spot where the CO$_2$ efflux from the soil was above the average, since they were done in a tree line and in positions, close to the trees, having high emission (Fig. 3 and Fig. 4a). A two-sample $t$-test was carried out to compare $R_s$ obtained from the two measurement systems. $R_s$ was significantly higher in multiplexed system (mean = $8.21 \mu\text{mol CO}_2 \text{m}^{-2} \text{s}^{-1}$) than in survey chamber mode (mean = $6.54 \mu\text{mol CO}_2 \text{m}^{-2} \text{s}^{-1}$; $t = 9.996$, $p < 0.001$, degree of freedom = 147). Observed temperature sensitivity was similar, but the basal
respiration, $R_b$, was larger in the spot where continuous measurements were taken (Table 3). Assuming that the observed relation between continuous and survey measurements was constant along the year, we calculated a constant multiplicative factor of 0.797 to upscale continuous measurements to the tree lines, and a second multiplicative factor of 0.831 to estimate the Sr in the grassed alleys (Fig. 4a). Total Rs resulted as the weighted average of these estimates.

By using the LUT method, upscaled $R_s$ and $R_h$ accounted for $801 \pm 65$ and $614 \pm 67 \text{gCm}^{-2} \text{yr}^{-1}$ respectively. To assess the robustness of these estimates we also modeled $R_s$ by the $Q_{10}$ (van’t Hoff, 1884) and by the Lloyd and Taylor (1994) models, and with both models the yearly amount of $R_s$ and $R_h$ were within the uncertainty levels mentioned above.

### 3.1.3 Fluxes from biometric sampling

The monthly results of the biometric measurements of NPP are shown in Fig. 5 and in Table 4 with related uncertainties.

Leaf production, $106 \pm 5 \text{gCm}^{-2} \text{yr}^{-1}$ represented 11% of NPP. Leaf growth occurred mostly between April and early June, when leaves represented 44% of NPP. A reduced leaf growth was observed until September.

Fine roots production showed three distinct peaks: the first one occurred before maximum leaf production, the second one just after it and a third one in October, after a reduced growth period in summer. On annual basis, NPPfr was 14% of NPP. Assuming that the increase in fine roots follows the same growth pattern as coarse roots, C allocated to standing fine roots is $6 \pm 1 \text{gCm}^{-2} \text{yr}^{-1}$, while the remainder, $124 \pm 27 \text{gCm}^{-2} \text{yr}^{-1}$, is supposed to be shed annually and to feed the detritus cycle.

The relative growth of woody organs was larger during spring and early summer (21% of total NPP) and decreased to 8% from the end of August till the end of the vegetative season. On annual basis, NPPwAG was $173 \pm 53 \text{gCm}^{-2} \text{yr}^{-1}$ (18% of total NPP); the largest part of this component was accounted by pruned wood feeding the detritus cycle ($147 \pm 45 \text{gCm}^{-2} \text{yr}^{-1}$), while the increment in standing wood biomass
was 26 ± 8 g C m\(^{-2}\) yr\(^{-1}\). Carbon allocated to belowground structural organs accounted for only 1% of total woody NPP (NPP\(_{BG}\) = 13 ± 3 g C m\(^{-2}\) yr\(^{-1}\)).

From June until the harvest, fruits represented the largest sink. Carbon allocation to this organ was more than 65% of total NPP in July and August and more that 75% of total NPP in September and October. On annual basis, carbon allocated to fruits was 495 ± 35 g C m\(^{-2}\), equal to 52% of total NPP.

Aboveground primary production of the understory (NPP\(_u\)) was a significant component of total NPP, accounting for 42 ± 3 g C m\(^{-2}\), 5% of total NPP, with a long growth period (March–November) and a relatively constant growth rate throughout the season.

The total NPP\(_{biom}\), obtained by the sum of the cumulated values of each considered NPP component (Table 5), was 960 ± 70 g C m\(^{-2}\) yr\(^{-1}\). Summarizing the fate of NPP components, we obtained that 45 ± 9 g C m\(^{-2}\) yr\(^{-1}\) represent an increase in standing biomass, 471 ± 35 g C m\(^{-2}\) yr\(^{-1}\) are exported, and 444 ± 53 g C m\(^{-2}\) yr\(^{-1}\) feed the detritus cycle.

### 3.2 Independent assessment of NPP, \(R_a\), GPP and the overall carbon cycle

The equation NPP + \(R_a\) = GPP shows the relation among these three fluxes and their importance for the determination of CUE. In this study, the annual amount of each of these three C fluxes was obtained following at least two independent pathways.

The results of biomass accumulation of the ecosystem components considered at each sampling date are reported in Table 4, while Fig. 6 shows the daily carbon uptake rate of both NPP\(_{biom}\) and NPP\(_{flux}\). Daily NPP\(_{biom}\) showed a less regular annual pattern of C uptake with respect to NPP\(_{flux}\), which was bell-shaped. Maximum daily NPP occurred from mid June until mid July for both NPP\(_{biom}\) (7.80 ± 2.18 g C m\(^{-2}\) d\(^{-1}\)) and NPP\(_{flux}\) (10.32 g C m\(^{-2}\) d\(^{-1}\)).

Although some discrepancies emerge when comparing the two curves (NPP\(_{flux}\) − NPP\(_{biom}\) = −2.9 g C m\(^{-2}\) d\(^{-1}\) in May, −1.5 g C m\(^{-2}\) d\(^{-1}\) in September, +1.3 g C m\(^{-2}\) d\(^{-1}\) in November), we found a good agreement between these two independent methods.
of determining NPP, confirmed by the surprisingly close value of yearly NPP_{biom} and NPP_{flux} values, which were 960 ± 68 and 994 ± 70 g C m^{-2} yr^{-1} respectively.

As mentioned above, R_a was the only element of the C cycle that was not measured directly. Each of the three equations applied implies the integration of different methodologies and leads to sensible different estimates of R_a (Table 6). Following Eq. (4), R_a was 303 ± 201 g C m^{-2} yr^{-1}, a value close to result of Eq. (5), which leads to a R_a of 269 ± 174 g C m^{-2} yr^{-1}. The third method, Eq. (6), relying on soil chamber methodology and measurements on N content in the biomass, gives estimates significantly higher with respect to the previous two, with R_a being 545 ± 44 g C m^{-2} yr^{-1}.

On a yearly basis, it was possible to estimate GPP independently from EC measurements. GPP_{BS} was obtained by summing NPP_{biom} with R_a determined by Eq. (6), therefore relying only on biometric and soil chamber measurements. While GPP_{EC} was 1263 ± 189 g C m^{-2} yr^{-1}, GPP_{BS} was found to be approximately 250 g C m^{-2} yr^{-1} greater (1505 ± 83 g C m^{-2} yr^{-1}).

All the C fluxes measured within the ecosystem during 2010, and lateral import and export, are shown in Fig. 7. The R_a value reported in Fig. 7 (372 ± 140 g C m^{-2} yr^{-1}) refers to the average ± se of R_a obtained by Eqs. (4), (5) and (6) presented in Table 6.

### 3.3 CUE: seasonal trend and yearly value

The seasonal trend of CUE_{biom} and CUE_{flux} is shown in Fig. 8. Significant differences between the two estimates occurred at the beginning of the growing season and at its end, after harvest. When analyzed separately, CUE_{biom} showed an irregular pattern with a decreasing trend throughout the season, while CUE_{flux} showed its highest values during the summer months. In both cases, CUE was above 0.5 for the whole growing season. Based on results shown in the previous sections, on a yearly basis we obtained four independent estimates (Table 7), giving an average value of CUE for the apple orchard of 0.71 ± 0.09.
4 Discussion

4.1 Magnitude of C fluxes

The first hypothesis of this study was that the main ecosystem C fluxes of a woody agro-ecosystem have the same order of magnitude as natural forest ecosystems growing in the same biome rank. Table 8 shows a comparison between meteorological and biological variables measured at the study site and at the temperate-humid deciduous forests reported in the global forests database published by Luyssaert et al. (2007). Regarding the average ecosystem characteristics, it appears that apple trees are smaller in size (height, AG and BG biomass) respect to trees growing in forests, and have a smaller LAI. Climate characteristics of the study site are very similar to the average observed in temperate-humid forests, with the exception of summer precipitation: in the apple orchard water was additionally supplied by irrigation, a practice that eliminates major stress due to drought in the summer period. Based on our results, EC ecosystem carbon fluxes (GPP, NEP and \( R_{\text{eco}} \)) obtained in the studied agro-ecosystem are quantitatively similar to a forested stand. We obtained slightly lower GPP and \( R_{\text{eco}} \) values and a larger NEP, but differences were within 100 g C m\(^{-2}\) yr\(^{-1}\).

The estimate of \( R_h \) was obtained by measuring \( R_s \) from trenching plots. This methodology for separating microbial and root respiration is widely applied for its simplicity and low cost, although it has several disadvantages (Subke et al., 2006; Lambert et al., 2011). Among them, the most important is probably the fact that part of the measured C may come from decomposition of roots that are excised during the trenching process (Hanson et al., 2000). We avoided accounting for the “priming effect” due to an excess of decomposable matter (Kuzyakov et al., 2000) starting the measurements approximately 10 months after the trenching plots were set. Another problem could arise because of the higher soil water content of trenching with respect to the control sample, due to the absence of root absorption. This may cause a change in the microbial community and in the rate of CO\(_2\) emission (Diaz-Pines et al., 2010) that
we prevented starting from June 2010 by installing a plastic shelter approximately 1 m above the trenching plot. It has also been suggested that small trenching acts as a sink for CO₂ from surrounding soil (Rachpal et al., 2006), thus causing an overestimation of \( R_h \). Despite all limitations of the trenching approach and the assumptions that need to be made, the meta-analytical review published by Subke et al. (2006) reports a general good agreement among different methodologies in soil respiration partitioning, reinforcing the reliability of our estimates.

The ratio \( R_h/R_s \) obtained in the present study (0.77) is within the range of the studies carried out over temperate forests, as reviewed by Subke et al. (2006), and higher than the value (0.65) reported in an apple orchard by Panzacchi et al. (2012). Our average \( R_h \) (614 ± 67 g C m\(^{-2}\) yr\(^{-1}\)) is also higher as compared to the average \( R_h \) values reported by Luyssaert et al. (2007) for temperate humid deciduous forests (387 ± 4 g C m\(^{-2}\) yr\(^{-1}\), Table 3). Besides the uncertainties of the methodology itself, this may also be due to a relatively high soil organic carbon content and to the superficial soil tillage which was periodically carried out along the apple tree strip (1.2 m wide) to control the growth of grass below the trees, a common practice in an organic production system (Reganold et al., 2001; Smith, 2004).

The protocol that was used to biometrically assess NPP accounts for four (out of six) hierarchical levels of the framework for net primary production that was proposed by Luyssaert and colleagues (2007). We did not account for neither photosynthates released to symbionts or as root exudates, nor for non-CO₂ carbon emission (VOC, CO, CH₄). Thus, besides uncertainties in the estimates of the other NPP components, the biometric value we found (960 ± 70 g C m\(^{-2}\) yr\(^{-1}\)) is likely to be an underestimation of the real NPP. There are few reliable estimations of the magnitude of the root exudates component and non-CO₂ carbon emissions on total NPP in the literature (Grayston et al., 1997; Millard et al., 2007). Those are often accounted for as missing NPP due to the intrinsic difficulty of their direct assessment (Luyssaert et al., 2007) under field conditions. In the review of 14-C labeling studies on plant-soil interactions published by Farrar et al. (2003), it is suggested that exudation may account for 5–10 % of net
C assimilation although Jones et al. (2004) revised this estimation to 2–4 % and other studies point out the dependency of the amount of root exudates on plant species, soil type and fertility and other climatic variables (Cheng et al., 2007; Jones et al., 2009; Vicca et al., 2012). The NPP found in this study was about 200 gCm$^{-2}$yr$^{-1}$ higher with respect to natural forests of the same biome rank (Table 8). In a less productive and less vigorous apple genotype, Panzacchi et al. (2012) have also found an average NPP value of 785 gCm$^{-2}$yr$^{-1}$. The presence of a large amount of fruit on an apple tree is known to enhance specific leaf photosynthesis as a consequence of their role as a sink, that allows for a more rapid download of photosynthates from the phloem respect to apple trees not bearing fruits (Giuliani et al., 1997; Tartachnyk and Blanke, 2004).

4.2 Annual and seasonal C allocation pattern

The second hypothesis of this study was that the main differences between natural and agricultural woody ecosystems lie in the allocation pattern of fixed C instead of in a different magnitude of C fluxes. Results from this study confirm this hypothesis, highlighting how this agro-ecosystem is strongly oriented to fruit production. While deciduous forests of temperate-humid biomes allocate the fixed C primarily into leaves, wood and roots, with an incidence on total NPP of 30, 43 and 27 %, respectively (Table 8), these three NPP components in the studied orchard accounted for only 11, 18 and 15 % of total NPP, with fruits being by far the major NPP contributor ($495 \pm 35$ gCm$^{-2}$yr$^{-1}$, 52 % of total NPP). When analyzing the seasonal trend of the C allocation pattern, it can be noticed that roughly 70 % of the leaves are set within 2 months after budburst, while aboveground woody organs show a constant growth until mid-August. Most root NPP is due to fine roots production, since apple trees are grafted on dwarfing rootstock and thus new coarse roots production is rather limited. Results on fine roots growth are consistent with findings published by Eissenstadt et al. (2006). The overall amount of fine root released to detritus cycle also agrees with findings of Wells and Eissenstadt (2001).
The apple orchard is also affected by significant lateral flows of C due to human activities which occur, with a different magnitude, in both inputs and outputs. As lateral C input, we must consider organic fertilization, a common agricultural practice repeated every year for ensuring the reintegration of exported mineral nutrients, which accounted for about 35 g C m\(^{-2}\) yr\(^{-1}\). The lateral C output flow is quantitatively much more important since it is represented by the harvested apples which are taken away from the ecosystem for commercial purposes. In 2010 this component was quantified as 471 ± 35 g C m\(^{-2}\) yr\(^{-1}\) (74 t ha\(^{-1}\) fresh weight, corresponding to 95 % of NPP), while the remaining 5 % is accounted for by early drops and uncollected fruits (24 ± 3 g C m\(^{-2}\) yr\(^{-1}\)) that enter the detritus cycle. The estimated fruit production is slightly higher than the average yields observed in this area of intense apple production (approx. 60 t ha\(^{-1}\)).

The fate of the C contained in the abscised leaves of apple trees during their decomposition on the soil surface has been reported by Tagliavini et al. (2007) who showed that approximately 80 % of initial amounts are lost in the first two years after leaf abscission. Ventura et al. (2009) reported that the decomposition of peach leaf litter is complete after 3 yr and about 10 % of initial amounts of leaf C are likely to be transformed into more stable C forms in the soil.

Our findings are consistent with other studies on apple trees of different growing conditions and varieties (Palmer, 1988; Minchin et al., 1997; Faqi et al., 2008), while a similar incidence of fruit production on total NPP was also found in other agro-ecosystems, such as a coconut palm plantation (Navarro et al., 2008), peach (Chalmers and van den Ende, 1975), orange (Liguori et al., 2009) and kiwifruit orchards (Rossi et al., 2007). For comparison with other croplands see Ciais et al. (2010).

The fate of allocated carbon is partially different from forests, with a fraction of C exported from the ecosystem through apple production representing nearly half of total net primary production. About 46 % of annual NPP feeds the detritus cycle, and this is similar to natural forests if tree mortality is not considered (e.g. Tan et al., 2010), giving ample potential for the soil of the fruit tree ecosystem to act as a net carbon sink. The
amount of NPP which increases the standing biomass, contributing to ecosystem C storage function (5% of total NPP), is conversely much lower than in forests.

### 4.3 Independent assessment of NPP, $R_a$ and GPP

The methodological approach employed in the present study gave the opportunity to assess C fluxes involved in CUE determination through independent pathways. This allowed a cross check of the estimated fluxes, thus obtaining an important feedback on the robustness of the estimation.

NPP was assessed by using only biometric measurements of different ecosystem components ($NPP_{biom}$) and through the sum of CO$_2$ fluxes obtained by EC (NEP) and soil chamber ($R_h$) methodology ($NPP_{flux}$). The seasonal trends, as well as the yearly cumulated values, are very close to each other, supporting the reliability of the employed methodological approach.

Since no direct measurements of $R_a$ were carried out, a multiple approach was applied to assess it. As shown in Table 6, $R_a$ was estimated by: (i) coupling EC with biometric measurements (GPP-NPP), (ii) coupling EC with soil chamber measurements ($R_{eco} - R_h$) and (iii) coupling soil chambers measurements and biomass nitrogen determination. The difference between the results of the latter method and the previous two highlights the most important discrepancy we registered in the present study, which is due to the relatively higher amount of C fluxes obtained via soil chambers with respect to the EC derived $R_{eco}$. This problem is also reported in other studies (Ryan et al., 1997; Law et al., 1999). EC measurements may be affected by a series of different systematic and random errors (Baldocchi, 2003; Richardson et al., 2006) which may lead to uncertainties in the yearly NEE estimate, as well as the derived $R_{eco}$ and GPP. We tried several methodologies for the gap-filling and flux partitioning procedure of our dataset ranging from marginal distribution sampling (Reichstein et al., 2005), to light response curve (Lasslop et al., 2010), and to LUT (Moffat et al., 2007; Rossini et al., 2010 and present study) and we used the results from methodologies different from EC to assess the uncertainty in C flux estimates. Since the yearly amount of $R_s$
measured by our multiplexed system was consistent with other published data (Blanke et al., 1995; Koerber et al., 2009; Ceccon et al., 2011) and we are quite confident about the results derived from biometric measurements (NPP\textsubscript{biom}). LUT was the methodology which gave the most reliable estimate of EC-derived GPP and $R_{\text{eco}}$. We chose it as the reference method, using the estimates from other methods to assess flux uncertainty.

The third method to assess $R_a$ allowed us to obtain a second estimate of yearly GPP ($\text{GPP}_{BS}$) completely independent of eddy covariance measurements. This yielded higher figures with respect to $\text{GPP}_{EC}$ by approximately 250 g C m\(^{-2}\) yr\(^{-1}\), suggesting an underestimation of C fluxes with EC.

4.4 Annual and seasonal CUE

The hypothesis that CUE is constant among forests (Gifford, 1994, 2003; Dewar et al., 1998; Ryan et al., 1997), with a possible appropriate universal value of 0.47 (Waring et al., 1998), was rejected by De Lucia and colleagues (2007) who reported a systematic large variation of CUE among forest types (from 0.23 to 0.83) over a wide range of published data. Amthor (2000) suggested a theoretical possible interval of CUE between 0.2 and 0.65, confirmed also by experimental results on herbaceous species by van Iersel (2003), with crops generally having a higher value with respect to “natural” vegetation (Amthor, 1989). More recently, Van Oijen et al. (2010) proposed a theoretical approach, based on the law of mass conservation, to analyze the quantitative relationships between photosynthesis, respiration, growth and carbon storage in vegetation, suggesting a narrowly constrained respiration to photosynthesis ratio. Vicca et al. (2012) observed a significant spread in CUE values among different ecosystems, suggesting the nutritional status, and the consequent reduced allocation of photosynthates to symbionts, as the main driver for the variation found. The high CUE value found in this agro-ecosystem ($0.71 \pm 0.09$) can be explained by a relatively low level of plant respiration ($R_a$), by a lower amount of photosynthates allocated to non-biomass component of NPP, or both.
A possible explanation for a relatively low $R_a$ level may be found in both structural characteristics of the orchard and climatic conditions of the site. Regarding the first aspect, despite the fact that our understanding of plant $R_a$ is still incomplete and poorly parameterized in current models (Piao et al., 2010), several studies (Ryan et al., 1997; Arneth et al., 1998; Law et al., 1999) highlighted the importance of the biomass composition in determining the total $R_a$, with foliage respiration having the greatest incidence on total $R_a$ when considered over unit of biomass. In our apple orchard, leaves represent only 11% of total NPP with respect to the average value of 30% for deciduous forests ($n = 32$) calculated from the database of Luyssaert et al. (2007). Crops, on the contrary, allocate most of the C in storage organs such as seeds and tubers which have a relatively small growth respiration and a generally low maintenance respiration (Amthor, 2000). In our case, 52% of NPP was represented by fruits. Apples fruits have a specific cost for synthesis and maintenance of approximately 1.15 g glucose g$^{-1}$, significantly lower of that of apple leaves (1.44 g glucose g$^{-1}$) and wood (1.30–1.33 g glucose g$^{-1}$) (Walton et al., 1999) and a specific dark respiration that rapidly declines as growth by cell expansion begins (Jones, 1981; Bepete and Lakso, 1997). Moreover, the yearly pruning of apple trees, favors the exposition of apple leaves to direct light, thus enhancing their potential photosynthetic capacity. Our findings confirm the hypothesis that apple trees have relatively low autotrophic respiration rates compared to many other plants due to the low construction costs of fruits (Lakso et al., 1999). The fact that apple trees grafted on dwarfing rootstocks, like in our study orchard, have a relatively small tree framework and root system, likely contributes to explaining the high value of CUE. Additionally, the low nitrogen content of apple fruits (0.29%) associated with their elevated incidence on total plant biomass (33 ± 7%), are in line with Reich et al. (2006; see also Ryan et al., 1996), who observed an almost linear correlation between total plant nitrogen and total plant respiration.

Climatic conditions may also contribute to explain the high CUE observed: following findings of Piao et al. (2010) over a wide range of ecosystems, and the related debate (Enquist, 2011; Chen et al., 2011), it emerges a possible role of the mean annual
temperature (MAT) in controlling the $R_a$ to GPP ratio, and the MAT of our site (11.5°C) is very close to the indicated one (~11.0°C) at which this ratio reaches its minimum at global scale. In addition, optimal conditions of water availability prevent CUE drop due to drought (Metcalfe et al., 2011; Panzacchi et al., 2012).

The ratio between biomass production and GPP observed in the studied orchard is well above the regression line found in Vicca et al. (2012) even for the most fertile forest ecosystems. This suggests that multiple factors enhancing CUE have to be considered, including climate and plant physiological traits.

This study helped assess the seasonal trend of CUE, using both NPP$_{\text{biom}}$ and NPP$_{\text{flux}}$ over the same GPP$_{\text{EC}}$ values. According to Campioli et al. (2011), it is important to specify that the variability of GPP estimates over a short period, depending on the selected partitioning method, may have a great effect on the CUE value, thus making it difficult to speculate on the absolute values observed along the season in each estimate, which in our case was occasionally above the unit. As shown in Fig. 8, the greatest discrepancy between the two curves of CUE$_{\text{biom}}$ and CUE$_{\text{flux}}$ occurs in spring, particularly in the first two months after budburst, where NPP$_{\text{biom}}$ estimate was greater than NPP$_{\text{flux}}$ and in the autumn, when the opposite was observed. Although the variation of carbon reserves along the year in mobile forms, such as starch, was not measured, we interpreted these results as a clear sign of remobilization (Mauler et al., 2004; Millard et al., 2007), with apple trees using stored carbohydrates in the first two months after budburst and likely re-allocating C to storage organs after harvest. Although a spring peak of CUE followed by a quick decline after May was observed by Campioli et al. (2011) in a temperate beech forest, in our case the NPP : GPP ratio was high also throughout the summer, mainly due to the continued biomass accumulation in fruits. This suggests a lower accumulation of nonstructural soluble carbohydrates, with respect to what is occurring in forests (Hoch et al., 2003).
5 Conclusions

This study demonstrates that the main ecosystem carbon fluxes of an apple orchard have a comparable magnitude with respect to deciduous forests growing in similar climate conditions. The major differences were a higher NPP in the apple orchard and a different allocation pattern of fixed C, with fruits representing approximately half of total yearly NPP. CUE, obtained from four partially independent methods, was 0.71 ± 0.09, indicating an elevated capacity of the orchard to allocate photosynthates to biomass production. The high amount of fruit biomass may be a possible explanatory reason for the high CUE found in the present study, because of both the reported low respiratory costs of fruits, and their low nitrogen content. The environmental conditions present in the orchards, obtained by pruning and supplying water and nutrients to the soil, represent another explanation for the high CUE, possibly limiting the transfer of photosynthates to non-biomass components, such as root exudates, of NPP.

Given the high amount of carbon allocated to apple fruit, mostly exported from the ecosystem every year, and the limited amount of C that increases the standing biomass of apple trees, the role of the apple orchard to act as a sink of carbon to the atmosphere largely depends on the persistence of the fraction of C which annually feeds the detritus cycle.

This work suggests that in global biogeochemical modeling it has to be considered that the CUE of agro-ecosystems can be higher than that of forests growing in a similar climate. Further research is needed to establish whether the high CUE observed is a common feature of agricultural systems.
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Carbon use efficiency in an apple orchard

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Table 1. Coefficients of the power allometric equation found for aboveground woody biomass (wood\textsubscript{AG}) and below ground woody biomass (wood\textsubscript{BG}). The “wood\textsubscript{AG}” equation is calculated using the “pruned above ground woody biomass” equation (wood\textsubscript{AG,p}) at the beginning of the season and the “not-pruned aboveground woody biomass” equation at the end of the season (wood\textsubscript{AG,np}), thus accounting for the pruning material. Reported are parameters with their relative standard errors.

<table>
<thead>
<tr>
<th>Woody organs</th>
<th>n</th>
<th>Intercept ± se</th>
<th>Exponent ± se</th>
<th>$R^2$</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>wood\textsubscript{AG,np}</td>
<td>11</td>
<td>229.3158 ± 1.3820</td>
<td>1.6115 ± 0.1787</td>
<td>0.9105</td>
<td>&lt; 0.001</td>
</tr>
<tr>
<td>wood\textsubscript{AG,p}</td>
<td>11</td>
<td>202.9379 ± 1.3682</td>
<td>1.6115 ± 0.1787</td>
<td>0.9105</td>
<td>&lt; 0.001</td>
</tr>
<tr>
<td>wood\textsubscript{AG}</td>
<td>2</td>
<td>0.0384 ± na</td>
<td>6.2470 ± na</td>
<td>na</td>
<td>na</td>
</tr>
<tr>
<td>wood\textsubscript{BG}</td>
<td>11</td>
<td>46.7026 ± 1.9599</td>
<td>1.7694 ± 0.3716</td>
<td>0.7391</td>
<td>0.0014</td>
</tr>
</tbody>
</table>
Table 2. Table 2. Distribution of dry weight (DW), C and N stocks within the ecosystem at the beginning of the growing season (March 2010). \( \text{Wood}_{\text{AG}} \) includes trunks and standing branches of different age; \( \text{Wood}_{\text{BG}} \) represents coarse roots (Table 1). Fine roots were obtained by the intensive soil core sampling conducted in March 2010.

<table>
<thead>
<tr>
<th>Components (March 2010)</th>
<th>DW ( \text{kg m}^{-2} )</th>
<th>C ( \text{kg m}^{-2} )</th>
<th>N ( \text{kg m}^{-2} )</th>
</tr>
</thead>
<tbody>
<tr>
<td>( \text{Wood}_{\text{AG}} )</td>
<td>1.85</td>
<td>0.84</td>
<td>0.014</td>
</tr>
<tr>
<td>( \text{Wood}_{\text{BG}} )</td>
<td>0.42</td>
<td>0.19</td>
<td>0.003</td>
</tr>
<tr>
<td>Fine roots</td>
<td>0.29</td>
<td>0.13</td>
<td>0.004</td>
</tr>
<tr>
<td>Total</td>
<td>2.56</td>
<td>1.16</td>
<td>0.021</td>
</tr>
</tbody>
</table>
Table 3. Parameters of the $Q_{10}$ model fitted for the contemporary measurements of soil respiration obtained by the multiplexed system and the survey chamber system.

<table>
<thead>
<tr>
<th>Methodology</th>
<th>$R_b$</th>
<th>$Q_{10}$</th>
</tr>
</thead>
<tbody>
<tr>
<td>Continuous measurements</td>
<td>$6.079 \pm 0.406$</td>
<td>$1.221 \pm 0.061$</td>
</tr>
<tr>
<td>Survey chamber</td>
<td>$4.668 \pm 0.498$</td>
<td>$1.226 \pm 0.097$</td>
</tr>
</tbody>
</table>
Table 4. Biomass production (gDW m\(^{-2}\) ± standard error) in the monitored tree organs between each sampling date.

<table>
<thead>
<tr>
<th>Sampling date</th>
<th>Leaves</th>
<th>Fruits</th>
<th>Wood(_{AG})</th>
<th>Wood(_{BG})</th>
<th>Fine roots</th>
<th>Understory</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td>20 Apr 2010</td>
<td>32.7 ± 1.7</td>
<td>0</td>
<td>26.9 ± 59.1</td>
<td>−0.6 ± 3.4</td>
<td>85.9 ± 45.6</td>
<td>9.0 ± 1.2</td>
<td>153.8 ± 74.7</td>
</tr>
<tr>
<td>11 May 2010</td>
<td>108.2 ± 5.5</td>
<td>29.4 ± 4.6</td>
<td>75.8 ± 31.1</td>
<td>9.6 ± 4.0</td>
<td>22.7 ± 52.7</td>
<td>4.3 ± 0.1</td>
<td>250.1 ± 61.7</td>
</tr>
<tr>
<td>23 Jun 2010</td>
<td>48.6 ± 6.2</td>
<td>194.5 ± 41.2</td>
<td>79.6 ± 21.4</td>
<td>9.3 ± 1.7</td>
<td>87.8 ± 59.0</td>
<td>20.1 ± 4.4</td>
<td>440.0 ± 75.4</td>
</tr>
<tr>
<td>15 Jul 2010</td>
<td>22.8 ± 4.2</td>
<td>273.5 ± 54.9</td>
<td>75.1 ± 68.1</td>
<td>1.9 ± 3.2</td>
<td>17.9 ± 65.5</td>
<td>17.3 ± 1.5</td>
<td>408.5 ± 109.4</td>
</tr>
<tr>
<td>20 Aug 2010</td>
<td>7.8 ± 2.5</td>
<td>266.4 ± 57.7</td>
<td>89.8 ± 54.1</td>
<td>7.2 ± 1.9</td>
<td>15.0 ± 48.0</td>
<td>20.0 ± 2.6</td>
<td>406.2 ± 92.6</td>
</tr>
<tr>
<td>15 Sep 2010</td>
<td>12.4 ± 3.4</td>
<td>294.3 ± 61.4</td>
<td>30.4 ± 30.4</td>
<td>1.1 ± 1.1</td>
<td>10.8 ± 51.6</td>
<td>24.3 ± 3.8</td>
<td>373.3 ± 85.9</td>
</tr>
<tr>
<td>14 Oct 2010</td>
<td>0</td>
<td>181.2 ± 33.7</td>
<td>17.5 ± 9.3</td>
<td>2.6 ± 1.2</td>
<td>35.5 ± 75.2</td>
<td>8.3 ± 2.3</td>
<td>245.1 ± 83.0</td>
</tr>
<tr>
<td>16 Nov 2010</td>
<td>0</td>
<td>0</td>
<td>−13.3 ± 9.6</td>
<td>−1.8 ± 1.1</td>
<td>16.3 ± 78.1</td>
<td>0</td>
<td>16.3 ± 78.1</td>
</tr>
<tr>
<td>Total</td>
<td>232 ± 10</td>
<td>1239 ± 88</td>
<td>382 ± 117</td>
<td>29 ± 7</td>
<td>292 ± 62</td>
<td>103 ± 6</td>
<td>2278 ± 160</td>
</tr>
</tbody>
</table>
Table 5. Yearly NPP values of the ecosystem components measured biometrically (data are expressed in gDW m\(^{-2}\) yr\(^{-1}\) and g C m\(^{-2}\) yr\(^{-1}\) ± standard error of the mean).

<table>
<thead>
<tr>
<th>NPP component</th>
<th>DW (g m(^{-2}) yr(^{-1}))</th>
<th>DW (%)</th>
<th>gC gDW(^{-1})</th>
<th>C (g m(^{-2}) yr(^{-1}))</th>
<th>C (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>NPP(/)</td>
<td>232 ± 10</td>
<td>10</td>
<td>0.458</td>
<td>106 ± 5</td>
<td>11</td>
</tr>
<tr>
<td>NPP(f)</td>
<td>1239 ± 88</td>
<td>55</td>
<td>0.400</td>
<td>495 ± 35</td>
<td>52</td>
</tr>
<tr>
<td>NPP(_{AG})</td>
<td>382 ± 117</td>
<td>17</td>
<td>0.454</td>
<td>173 ± 53</td>
<td>18</td>
</tr>
<tr>
<td>NPP(_{BG})</td>
<td>29 ± 7</td>
<td>1</td>
<td>0.453</td>
<td>13 ± 3</td>
<td>1</td>
</tr>
<tr>
<td>NPP(_{fr})</td>
<td>292 ± 62</td>
<td>13</td>
<td>0.444</td>
<td>130 ± 28</td>
<td>14</td>
</tr>
<tr>
<td>NPP(_{u})</td>
<td>103 ± 6</td>
<td>5</td>
<td>0.410</td>
<td>42 ± 3</td>
<td>4</td>
</tr>
<tr>
<td>NPP total</td>
<td>2278 ± 160</td>
<td>100</td>
<td>960 ± 70</td>
<td>100</td>
<td></td>
</tr>
</tbody>
</table>
Table 6. Computed values of $R_a$ (± se) obtained from the three methodologies discussed in paragraph 2.4 (Eqs. 4, 5 and 6). The coefficient $k$ obtained from the ratio between total and BG nitrogen content in the standing biomass was 2.92. The average $R_a$ value (372 ± 140 g C m$^{-2}$ yr$^{-1}$) is reported in Fig. 7.

<table>
<thead>
<tr>
<th>Model</th>
<th>Methodology</th>
<th>$R_a$ (g C m$^{-2}$ yr$^{-1}$)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1 GPP - NPP$_{biom}$</td>
<td>EC – biometric</td>
<td>303 ± 202</td>
</tr>
<tr>
<td>2 $R_{eco} - R_h$</td>
<td>EC – soil chambers</td>
<td>269 ± 174</td>
</tr>
<tr>
<td>3 $k \times R_{a_{BG}}$</td>
<td>soil chambers</td>
<td>545 ± 44</td>
</tr>
</tbody>
</table>
Table 7. The four independent approaches used to assess the CUE (± se) of the studied apple orchard (average = 0.71 ± 0.09).

<table>
<thead>
<tr>
<th>Model</th>
<th>CUE</th>
</tr>
</thead>
<tbody>
<tr>
<td>NPP_{biom}/GPP_{EC}</td>
<td>0.76 ± 0.13</td>
</tr>
<tr>
<td>NPP_{flux}/GPP_{EC}</td>
<td>0.79 ± 0.13</td>
</tr>
<tr>
<td>NPP_{biom}/GPP_{BS}</td>
<td>0.64 ± 0.06</td>
</tr>
<tr>
<td>NPP_{flux}/GPP_{BS}</td>
<td>0.66 ± 0.06</td>
</tr>
</tbody>
</table>
Table 8. Table of comparison between natural woody ecosystems of temperate humid biomes (dataset of Luyssaert et al., 2007) and the studied apple orchard, a woody agro-ecosystem.

<table>
<thead>
<tr>
<th>Stand characteristics (mean ± SD)</th>
<th>Temperate humid deciduous forest (2010)</th>
<th>apple orchard (2010)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Latitude (°)</td>
<td>44 ± 9</td>
<td>46</td>
</tr>
<tr>
<td>Max LAI (m² m⁻²)</td>
<td>6.1 ± 3.5</td>
<td>2.8</td>
</tr>
<tr>
<td>Tree height (m)</td>
<td>19 ± 7</td>
<td>4</td>
</tr>
<tr>
<td>Tree density (number ha⁻¹)</td>
<td>1723 ± 2439</td>
<td>3330</td>
</tr>
<tr>
<td>Stand age</td>
<td>75 ± 50</td>
<td>11</td>
</tr>
<tr>
<td>AG biomass (g C m⁻²)</td>
<td>10882 ± 5670</td>
<td>841 ± 81</td>
</tr>
<tr>
<td>BG biomass (g C m⁻²)</td>
<td>2565 ± 2609</td>
<td>320 ± 15</td>
</tr>
</tbody>
</table>

**Stand climate (mean ± SD)**

<table>
<thead>
<tr>
<th></th>
<th>Temperate humid deciduous forest (2010)</th>
<th>apple orchard (2010)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mean winter temperature (°C)</td>
<td>2 ± 9</td>
<td>1 ± 4</td>
</tr>
<tr>
<td>Mean summer temperature (°C)</td>
<td>20 ± 5</td>
<td>22 ± 5</td>
</tr>
<tr>
<td>Precipitation sum winter (mm)</td>
<td>183 ± 164</td>
<td>152</td>
</tr>
<tr>
<td>Precipitation sum summer (mm)</td>
<td>356 ± 259</td>
<td>293(+350*)</td>
</tr>
<tr>
<td>Net radiation winter (W m⁻²)</td>
<td>150 ± 100</td>
<td>9 ± 93</td>
</tr>
<tr>
<td>Net radiation summer (W m⁻²)</td>
<td>425 ± 78</td>
<td>162 ± 258</td>
</tr>
<tr>
<td>Mean winter air humidity (%)</td>
<td>79 ± 11</td>
<td>72 ± 23</td>
</tr>
<tr>
<td>Mean summer air humidity (%)</td>
<td>77 ± 5</td>
<td>62 ± 23</td>
</tr>
</tbody>
</table>

**Mean C fluxes (mean ± SE)**

<table>
<thead>
<tr>
<th></th>
<th>Temperate humid deciduous forest (2010)</th>
<th>apple orchard (2010)</th>
</tr>
</thead>
<tbody>
<tr>
<td>GPP</td>
<td>1375 ± 12 (n = 22)</td>
<td>1263 ± 189</td>
</tr>
<tr>
<td>NPP</td>
<td>738 ± 8 (n = 52)</td>
<td>960 ± 70</td>
</tr>
<tr>
<td>NPP/</td>
<td>235 ± 2 (n = 32)</td>
<td>106 ± 5</td>
</tr>
<tr>
<td>NPP\textsubscript{AG}</td>
<td>329 ± 10 (n = 21)</td>
<td>173 ± 53</td>
</tr>
<tr>
<td>NPP\textsubscript{fr} + NPP\textsubscript{BG}</td>
<td>207 ± 3 (n = 52)</td>
<td>143 ± 28</td>
</tr>
<tr>
<td>NPP\textsubscript{f}</td>
<td>n.a.</td>
<td>495 ± 35</td>
</tr>
<tr>
<td>NPP\textsubscript{u}</td>
<td>n.a.</td>
<td>42 ± 3</td>
</tr>
<tr>
<td>NEP</td>
<td>311 ± 7 (n = 29)</td>
<td>380 ± 30</td>
</tr>
<tr>
<td>(R_{\text{eco}})</td>
<td>1048 ± 13 (n = 24)</td>
<td>883 ± 160</td>
</tr>
<tr>
<td>(R_a)</td>
<td>673 ± 22 (n = 15)</td>
<td>372 ± 140</td>
</tr>
<tr>
<td>(R_i)</td>
<td>387 ± 4 (n = 40)</td>
<td>614 ± 67</td>
</tr>
<tr>
<td>CUE</td>
<td>0.54 ± 0.01</td>
<td>0.71 ± 0.09</td>
</tr>
</tbody>
</table>

* indicates the cumulated irrigation of the summer period
Fig. 1. Eddy covariance measured and derived C fluxes (g C m$^{-2}$ d$^{-1}$). Blue dots show NEE, with negative values indicating days in which the ecosystem is acting as a sink of C. Red triangle and green squares represent daily $R_{\text{eco}}$ and GPP obtained from flux partitioning of NEE data via LUT method.
Fig. 2. Average daily soil respiration (g C m\(^{-2}\) d\(^{-1}\)) measured in control (a) and trenching plots (b) with the multiplexed system along the tree-line. Data from each single collar are plotted with gray asterisk (*). Dots represent daily \(R_s\) (a) and \(R_h\) (b), when data from at least three chambers passed the quality test. Bars are standard deviation of the mean.
Fig. 3. Comparison of $R_s$ data ($\mu$mol CO$_2$ m$^{-2}$ s$^{-1}$) obtained from the multiplexed system (a) with those obtained from the survey chamber (b). Crosses indicate the average value, blue bars indicate the standard error. Letters (A to F) represent the six plots of the survey campaign. All the data refers to the same time period (24–30 June 2010).
Fig. 4. Soil respiration ((a), µmol CO$_2$ m$^{-2}$ s$^{-1}$) and N content in root biomass ((b), g N m$^{-3}$ soil) measured across the tree-line, where zero indicates the position of the apple tree. Measurements were performed in the six different plots used for the spatial survey. Bars represent standard error (se) of the mean.
Fig. 5. NPP (gC m\(^{-2}\)) between each sampling date interval calculated in the six considered ecosystem compartments. Bars width reflects the time (in days) occurred between successive biometric samplings.
Fig. 6. Annual pattern of NPP (gC m$^{-2}$ d$^{-1}$). The dashed line represents daily NPP obtained from biometric measurements (NPP$_{biom}$), and the solid line represents the 15-days moving average of NPP obtained by summing the daily fluxes of $R_h$ and NEP (NPP$_{flux}$). Bars represent standard error (se) of the mean daily NPP$_{biom}$. 
Fig. 7. Sankey plot of the carbon cycle of the studied agro-ecosystem for the year 2010. Data are in gCm$^2$y$^{-1}$ for each component of the C cycle. Arrow’s width reflects the flux size. GPP = gross primary production; $R_a$ = autotrophic respiration; $R_{eco}$ = ecosystem respiration; $R_h$ = heterotrophic respiration; NPP = net primary productivity, NPP$_{AG}$ = aboveground wood NPP; NPP$_l$ = leaf NPP; NPP$_f$ = fruit NPP; NPP$_u$ = understory NPP; NPP$_{BG}$ = belowground wood NPP; NPP$_{fr}$ = fine roots NPP; OC$_{fert}$ = organic carbon content of fertilizer; NEP = net ecosystem productivity; fruit harvest = fruit production exported from the ecosystem.
Fig. 8. Seasonal trend of the carbon use efficiency. Triangles represent CUE values obtained by dividing $\text{NPP}_{\text{biom}}$ and $\text{GPP}_{\text{EC}}$ (CUE\textsubscript{biom}); circles indicate CUE values obtained from the ratio between $\text{NPP}_{\text{flux}}$ and $\text{GPP}_{\text{EC}}$ (CUE\textsubscript{flux}). Time period considered is the growing season 2010, from DOY 77 till DOY 291.