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# Salinization alters fluxes of bioreactive elements from streams and soils across land use

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## Abstract

There has been increased salinization of fresh water over decades due to the use of road salt deicers, wastewater discharges, human-accelerated weathering, and ground-water irrigation. The effects of increased salinization on biogeochemical cycles in fresh-water ecosystems are still not well understood. We investigated potential impacts of increased salinization on fluxes of bioreactive elements from stream sediments and riparian soils to overlying stream water. Two-day incubations of sediments and soils with stream and deionized water across 3 salt levels were conducted at 8 routine monitoring stations at the Baltimore Ecosystem Study Long-Term Ecological Research (LTER) site in the Chesapeake Bay watershed. Ambient stream chemistry was also monitored before and after a snow event coinciding with road salt additions. Results indicated: (1) salinization typically increased sediment releases of labile dissolved organic carbon (DOC), dissolved inorganic carbon (DIC), total dissolved Kjeldahl nitrogen (TKN) (ammonium + ammonia + dissolved organic nitrogen), and sediment transformations of nitrate, (2) salinization generally decreased DOC aromaticity and fluxes of soluble reactive phosphorus (SRP) from both sediments and soils, (3) the effects of increased salinization on sediment releases of DOC and TKN and DOC quality increased with percentage watershed urbanization. The differential responses of riparian soils and sediments to increased salinization were likely due to differences in organic matter amounts and composition. Results of the sediment and soil incubations were used to interpret changes in ambient stream chemistry before and after a snow event. Our results suggest that short-term increases in salinization can cause releases of significant amounts of labile organic carbon and nitrogen from stream substrates and organic transformations of nitrogen and phosphorus. Given that salinization of fresh water will increase in the future, potential impacts on coupled biogeochemical cycles and water quality should be expected.

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# 1 Introduction

Salt concentrations in freshwaters are rapidly increasing at a regional scale in the United States, especially in urban watersheds with high population density (e.g., Kaushal et al., 2005; Findlay and Kelly, 2011; Steele and Aitkenhead-Peterson, 2011; Corsi et al., 2015). Most of the increased salinization can typically be attributed to road salt deicers and other industrial uses, wastewater discharges, groundwater irrigation, and human-accelerated weathering (e.g., Findlay and Kelly, 2011; Aitkenhead-Peterson et al., 2009; Kaushal et al., 2013). Increased salinization can have important environmental consequences for drinking water supplies, freshwater biodiversity, degradation of soils and groundwater, degradation of vehicles and infrastructure, and mobilization of inorganic and organic contaminants (Nielson et al., 2003; Kaushal et al., 2005; Findlay and Kelly, 2011; Corsi et al., 2015). Recent studies have further shown that increased salinization can influence biogeochemical cycles of bioreactive elements such as carbon and nitrogen (Green et al., 2008, 2009a, b; Green and Cresser, 2008; Compton and Church, 2011; Lancaster, 2012; Steele and Aitkenhead-Peterson, 2013) as well as phosphorus and sulfur (Compton and Church, 2011; Kim and Koretsky, 2011, 2013). Evidence is accumulating that increased salinization is an important process during the urban evolution of watersheds globally from decades to centuries (Kaushal et al., 2014a), and salinization has significant ecosystem effects over broader spatial and temporal scales (e.g., Findlay and Kelly, 2011; Kaushal and Belt, 2012; Corsi et al., 2015).

Although there has been increasing research, much more work needs to be done regarding the effects of increased salinization on biogeochemical cycles (Green et al., 2008, 2009a, b; Green and Cresser, 2008; Compton and Church, 2011; Kim and Koretsky, 2011; Lancaster, 2012; Steele and Aitkenhead-Peterson, 2013). Prior studies have commonly investigated the effects of salinization on fluxes and transformations of individual bioreactive elements or bulk concentrations of dissolved organic carbon (DOC) (e.g., Green et al., 2008, 2009; Steele and Aitkenhead-Peterson, 2011),

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sediments and soils under ambient and field conditions, similar water quality measurements were conducted before and after a snow event to compare with the laboratory salinization experiments.

## 2.1 Site description

5 The 8 stations were located at the US National Science Foundation supported Baltimore Ecosystem Study (BES) Long-Term Ecological Research (LTER) site, with land use varying from forest to low-density residential, agricultural, to suburban and urban (Table 1). The main focal watershed of the BES LTER site is the Gwynns Falls, a 17 150 ha watershed in the Piedmont physiographic province that drains into the northwest branch of the Patapsco River that flows into the Chesapeake Bay (Fig. 1). The Gwynns Falls sites traverse a rural/suburban to urban gradient from Glyndon (GFGL), Gwynnbrook (GFGB), Villa Nova (GFVN) to Carroll Park (GFCP) (Table 1). An agricultural stream (MCDN) is a small tributary to the Gwynns Falls draining a watershed dominated by row crop agriculture (corn, soybeans), while Dead Run (DRKR) is an urbanized tributary of the Gwynns Falls between GFVN and GFCP. Samples were also taken from a small urban tributary to the Gwynns Falls (GFGR), approximately 700 m above GFCP, which is highly contaminated with sewage (Kaushal et al., 2011). Baisman Run (BARN) is a low-density residential watershed located in the nearby Gunpowder Falls watershed that drains primarily forest land cover (Table 1; Fig. 1). The BES LTER site provides access to extensive background information and long-term monitoring of major anions, nutrients, and carbon concentrations and fluxes in streams (www.beslter.org; Groffman et al., 2004; Kaushal et al., 2008, 2011). Previous work has shown that watersheds of the BES LTER site can have considerably elevated levels of chloride and sodium (Kaushal et al., 2005; Kaushal and Belt, 2012).

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## 2.2 Sample collection and processing

Stream water, sediments, and soils for laboratory salinization experiments were collected on 8 March 2013, one day before a snow storm in the Baltimore-Washington D.C. metropolitan region. Three litres of stream water were collected at each site for the experiments and water quality analyses. Sediment samples were taken simultaneously along 4 cross-sections perpendicular to stream flow within 50 m of the primary sampling site (Duan and Kaushal, 2013). Along each stream cross section, surface sediments at three sites (left, middle and right) were collected. All sediments collected at these sites were well-mixed to make a composite sample. Soil samples from the riparian zone were also collected similar to sediment samples. Because the sites GFCP and GFGR were located very close to each other, only one soil sample was collected to represent these two sites. The sediment and soil samples were transferred to glass jars, and placed immediately into a cooler and brought back to lab. In the lab, sediments were sieved through a 2 mm sieve, and the < 2 mm fractions of sediments and soils were homogenized for incubation experiments (e.g., plant roots were picked from soils and discarded). Stream water was re-sampled 2 days later, when snow was melting, in order to examine the changes in water quality before and after the snow storm. Stream waters were temporarily stored at 2–4 °C along with sediments and soils prior to laboratory experiments. Approximately 100 mL aliquots of stream water were filtered through pre-combusted GF/F Whatman filters, and the filtrates were stored in a refrigerator for analyses of optical properties and dissolved inorganic carbon (DIC) measurements. Another aliquot was similarly filtered but frozen prior to analyses for concentrations of dissolved organic carbon (DOC), nutrients, and major anions.

## 2.3 Laboratory salinization experiments

For each laboratory salinization experiment, 60 g sub-samples of homogenised sieved sediments (< 2 mm) were inserted into a series of 125 mL glass flasks to cover the bottom of the flasks, and 100 mL of unfiltered stream waters were carefully added with

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a pipette in order to not disturb the sediments. In order to evaluate the potential effects of salinization, pure NaCl salt was amended to unfiltered stream water to obtain 3 concentration levels ( $0 \text{ g Cl}^{-1}$ ,  $2 \text{ g Cl}^{-1}$ , and  $4 \text{ g Cl}^{-1}$ ). These concentration levels were within the range reported for salt concentrations in ambient stream water at the Baltimore LTER site (Kaushal et al., 2005). All laboratory salinization experiments were conducted in duplicate. Simultaneously, streamwater samples without sediments were also incubated at the same 3 levels of salinization as controls, in order to separate potential contributions of sediments from that of stream water. The laboratory salinization experiments were conducted in the dark and at ambient temperatures ( $19\text{--}22^\circ\text{C}$ ), and the flasks were gently stirred for 2 days with a shaker table to simulate water movement in streams. During the 2 day incubations, the flasks were loosely capped to avoid external contamination while allowing for air entry. Laboratory salinization experiments for riparian soils were conducted similar to stream sediments, except that (1) deionized (DI) water (rather than stream water) was used for soil leaching, and (2) the samples were not stirred during the incubations assuming much slower hydrologic flow rates during soil infiltration than those for a stream. These experimental designs may have introduced potential artefacts such as no soil infiltration, constant temperature and no exposure to sunlight, which could influence results. However, previous studies investigating the potential impacts of salinization on soil biogeochemistry have used similar approaches (e.g., Green et al., 2008, 2009a; Compton and Church, 2011; Kim and Koretsky, 2013; Steele and Aitkenhead-Peterson, 2013). At the end of the incubations, the incubated stream water and DI water were filtered through pre-combusted GF/F Whatman filters for water chemistry analyses.

### 2.4 Chemical analysis

All filtrates were analyzed for major anions, nitrate, soluble reactive phosphorus (SRP), DOC, dissolved inorganic carbon (DIC), total dissolved nitrogen (TDN), and optical properties (absorbance and fluorescence). DOC, TDN and DIC concentrations were measured on a Shimadzu Total Organic Carbon Analyzer (TOC-V CPH/CPN)

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(Duan and Kaushal, 2013). Chloride, nitrate, and sulfate concentrations were measured with a Dionex ion chromatograph (ICS-1500, Dionex INC., USA), with an eluent of 3.5 mM of  $\text{Na}_2\text{CO}_3$  with 1.0 mM  $\text{NaHCO}_3$  and a flow rate of  $0.3 \text{ mL min}^{-1}$ . SRP was measured on an automated QuikChem 8500 Series 2 FIA System, using the ascorbic acid-molybdate blue method (Murphy and Riley, 1962). Total Kjeldahl nitrogen (TKN), including dissolved organic nitrogen, ammonia, and ammonium, was calculated by subtraction of nitrate-N from TDN.

Ultraviolet (UV) absorbance and fluorescence spectroscopy were used in characterization of DOC source and lability. Filtrates were scanned for absorbance from 200 to 800 nm with a Shimadzu UV-1800 Spectrophotometer. UV absorbance at 254 nm was used to calculate specific UV absorbance (SUVA) by normalizing for DOC concentration. SUVA is strongly correlated with percent aromaticity of organic matter as determined by  $^{13}\text{C}$  NMR (Weishaar et al., 2003), and thus can be a useful parameter for estimating terrestrial organic carbon sources in aquatic systems. Fluorescence measurements were made on a FluoroMax-4 Spectrofluorometer (Horiba Jobin Yvon, Edison NJ, USA) using the method that was described previously by Duan and Kaushal (2013). Basically, we used a 1 cm quartz cuvette with slit widths set to 5 nm. Excitation emission matrix scans (EEMs) were obtained by collecting a series of emission wavelengths ranging from 300 to 600 nm (2 nm increments) at excitation wavelengths ranging from 240 to 450 nm (5 nm increments). EEMs data were corrected for instrument biases, inner filtering and scatter removal, and calibrated values of fluorescence intensities at excitation/emission = 275 nm/340 nm and 350 nm/480 nm were recorded as protein-like and humic-like fluorophores (Coble, 1996; Stolpe et al., 2010). Relative to the humic-like fluorophore, the intensity of the protein-like fluorophore is generally higher in labile DOC sources (e.g., wastewater; Hudson et al., 2007) and positively correlated with DOC bioavailability (Balcarczyk et al., 2009; Lønborg et al., 2010). Thus, the ratio of the protein-like to the humic-like fluorophore (P/H) was calculated here as an index of organic carbon lability.

Ash free dry weight (AFDW) of the sediment and soil samples was analyzed as an index of organic matter content. Sediment and soil ash weights were calculated as the difference in weights before and after combustion at 550 °C for 4 h (APHA, 1998). Prior to combustion, sediments were dried at 105 °C for 4 h to remove water. Ash free dry weights were determined in triplicates.

## 2.5 Data analysis and statistics

Changes in concentrations of DOC and other water quality variables in stream water and DI water during the two-day incubations were calculated, and the changes were calibrated with the controls (incubations with unfiltered water or DI water only) to obtain fluxes from sediments or soils. Differences in chemical fluxes between incubations at the three salinization levels were tested using one-way ANOVA. Relationships between water quality variables and land use or between two sediment/soil fluxes were examined using Spearman's rank correlation. For all statistical tests, significance was accepted at  $\alpha = 0.05$ . Analyses were performed using SPSS version 17.0 (2014).

## 3 Results

### 3.1 Water and sediment chemistry

In stream waters that were used for laboratory salinization experiments, water chemistry varied considerably (Table 2). In general, concentrations of chloride ion ( $\text{Cl}^-$ ), sulfate-S ( $\text{SO}_4^{2-}$ -S) and DOC, and protein-like to humic-like fluorophore (P/H) ratios of DOC in stream water increased with watershed impervious surface cover (ISC) ( $r^2 = 0.77$ – $0.83$ ,  $p < 0.05$ ,  $n = 8$ ). SUVA showed an opposite trend and decreased with watershed ISC ( $r^2 = 0.79$ ,  $n = 8$ ,  $p < 0.05$ ).  $\text{Cl}^-$  concentrations also increased with ISC, but the coefficient was not significant ( $r^2 = 0.40$ ,  $n = 8$ ,  $p > 0.05$ ), and the highest value was not observed at the site GFGR with highest ISC. Nitrate-N ( $\text{NO}_3^-$ -N) and SRP con-

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centrations did not vary with watershed ISC, and the highest concentrations occurred at the agricultural site (MCDN; Table 2).

Sediment ash free dry weight (AFDW) also displayed an increasing trend with ISC (from 0.61 to 1.90 %) except one surprisingly high value (3.98 %) observed at GFCP. AFDW of the riparian soils (6.17–8.84 %) were significantly higher than the sediments ( $p < 0.05$ , one-way ANOVA) but did not vary significantly with watershed ISC (Table 2).

### 3.2 Influence of salinization on C fluxes and DOC composition

Sediments were a net source for both DOC and DIC. DOC release from sediments consistently increased with increasing salinization (Fig. 2a), and DOC values at 4 g CIL<sup>-1</sup> were 7.8 ± 1.9 times higher than those at 0 g CIL<sup>-1</sup>. Although salinization also increased the release of DIC (with one exception at GFCP), the highest DIC releases generally occurred at 2 g CIL<sup>-1</sup> (6 in 8 cases), and the highest DIC values were only 1.4 ± 0.02 times higher than those at 0 g CIL<sup>-1</sup> (Fig. 2b). Meanwhile, maximal sediment releases of DOC and DIC were 3.9 ± 0.8 and 0.51 ± 0.12 times higher than their ambient concentrations in streamwater.

Moreover, the effects of salinization on sediment releases differed among DOC fractions. Salinization consistently and considerably increased releases of the protein-like fluorophore, showing significant increases that were 6.7 ± 1.0 times higher at 4 g CIL<sup>-1</sup> relative to those at 0 g CIL<sup>-1</sup> (Fig. 2c). The effects of salinization on humic-like fluorophore releases, however, were not consistent and were much less (increase by 1.2 ± 0.1 time) (Fig. 2d). As a result, salinization consistently and considerably increased (by 5.9 ± 0.7 times) the protein to humic (P/H) ratio (Fig. 2e) – an index of DOC lability (Lønborg et al., 2010; Duan and Kaushal, 2013). The effects of salinization on DOC lability using fluorescence spectroscopy were further supported by absorbance measurements. Absorbance measurements showed a general decreasing trend in SUVA changes with increasing salinization (7 out of 8 cases; Fig. 2f).

Effects of laboratory salinization on DOC and DIC releases from soils were relatively more complex and not as consistent. In 4 out of 7 cases (POBR, MCDN, GFGB and

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soils in this study (Fig. 3). However, neither this model nor the flocculation/microbial-suppression mechanism can explain the consistent observation of DOC mobilization from sediments in our laboratory salinization experiments (Fig. 2), probably owing to differences in organic carbon composition and sources between stream sediments and riparian soils.

Our results from DOC characterization can provide further information for interpreting the differences in salinization effects on DOC releases between sediment and soils. Our results showed that only protein-like fluorophores were consistently and significantly remobilized from sediments with salinization (Fig. 2), which suggested that the increased DOC releases from sediments were mainly attributed to the releases of protein-like labile DOC fractions. Similar findings were also reported by Li et al. (2013), which showed that KCl can significantly increase the mobility of microbially-derived labile organic matter (indicated by the fluorescence index). Meanwhile, chemical analyses suggest that the protein-like fluorophores consist primarily of proteinaceous materials (e.g., proteins and peptides; Yamashita and Tanoue, 2003; Maie et al., 2007), and this DOC fraction is generally hydrophilic and low molecular weight (LMW) compounds (e.g., Sommerville and Preston, 2001). Results of Chen et al. (1989) and Fuchs et al. (2006) showed that solubility of the proteinaceous materials in LMW is generally neither affected by pH within normal range 6–9 nor by colloid coagulation. Therefore, increasing ionic strength (or salinization) can enhance the solubility of the proteinaceous materials via sodium dispersion (Green et al., 2008, 2009a) or through nonspecific electrostatic interactions at low salinities (called a “salting in” effect) (Tanford, 1961; Chen et al., 1989). Furthermore, because stream sediments are generally enriched in these labile, proteinaceous materials derived from biofilms (algae and microbes) and wastewater organics in urban watersheds (Kaushal et al., 2011, 2014b; Kaushal and Belt, 2012; Newcomer et al., 2012; Duan et al., 2014b), it is reasonable that salinization can mobilize a large amount of protein-like labile dissolved organic matter from stream sediments.

Relative to the proteinaceous materials, humic substances are larger hydrophobic molecules occurring in the colloidal size range (e.g., Aiken et al., 1985). This DOC

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fraction is readily subjected to flocculation (e.g., Sholkovit, 1976), sorption to mineral surfaces (Fox, 1991; Hedges and Keil, 1999), and pH suppression (Kipton et al., 1992; Li et al., 2007, 2013) with increasing ionic strength or salinization. The potential instability of the humic-like DOC fraction upon salinization was further supported by our present results and previous studies (e.g., Li et al., 2013), which showed that salinization consistently decreased SUVA of DOC released from soils and sediments (Figs. 2 and 3) – a parameter indicating DOC aromaticity (Weishaar et al., 2003). Relative to stream sediments, soil organic matter consists primarily of humic substances (up to 60–70%; Griffith and Schnitzer, 1975). In this study, we also observed much higher SUVA values in DOC leached from soils (around  $10 \text{ L}(\text{mg C})^{-1} \text{ m}^{-1}$ ) than from sediments ( $< 2 \text{ L}(\text{mg C})^{-1} \text{ m}^{-1}$ ) (Fig. 7). Due to larger fractions of humic substances, the effects of salinization on DOC leaching from soils were not consistent (Fig. 3), but depended on the relative importance of sodium dispersion and pH suppression (Green et al., 2008).

The effects of salinization on DIC fluxes from sediments and soils may involve organic carbon mineralization, dissolution of carbonate minerals, and  $\text{CO}_2$  exchange with the atmosphere. The potential importance of organic carbon mineralization and its influence on DIC releases during laboratory salinization experiments were supported by the observed increases in DIC concentrations with DOC releases (Fig. 6a and b). This mechanism seemed to fit better for soils, considering the strong linear relationship between DIC and DOC across soil salinization experiments (Fig. 6b). The complex relationships between DOC and DIC for sediment incubations (Fig. 6a) indicated the importance of other potential controls as well – e.g.,  $\text{CO}_2$  efflux to the atmosphere. It is known that the solubility coefficient for  $\text{CO}_2$  decreases with salinity (Weiss et al., 1974; Duan and Sun, 2003) and pH suppression, both of which could occur during salinization experiments (Green et al., 2008). Thus,  $\text{CO}_2$  efflux to the atmosphere may become a dominant control at higher salinities (e.g.,  $2\text{--}4 \text{ g Cl}^{-1}$ ), and further increases in salinization may decrease DIC releases despite of continuous releases of DOC (Fig. 2). Relative to organic carbon mineralization and  $\text{CO}_2$  efflux to the atmosphere, the ef-

fects of salinization on carbonate mineral solubility seemed relatively less important because crystalline rocks of igneous or metamorphic origin characterize the surface geology in the region (<http://www.mgs.md.gov/esic/geo/bal.html>). Additional evidence suggests that the solubility of carbonate minerals increases with salinization (Akin and Lagerwerff, 1965), but DIC releases from soils in laboratory salinization experiments of the present study did not follow this trend (Fig. 3).

## 4.2 Potential effects of salinization on N, P, and S leaching and transformation

We observed mobilization of TKN in response to salinization in both sediments and soils (Figs. 3 and 4), and this has also been observed in previous studies (mainly  $\text{NH}_4^+$ ; Duckworth and Cresser, 1991; Compton and Church, 2011; Kim and Koretsky, 2011). The consistent  $\text{NH}_4^+$  releases with salinization can be attributed to  $\text{Na}^+$  dispersion (Green and Cresser, 2008). That is, as a positively-charged ion,  $\text{NH}_4^+$  can be adsorbed on negatively-charged particles of soils and sediments (Nieder et al., 2011);  $\text{NH}_4^+$  retained on the cation exchange sites can be greatly reduced by the presence of sodium ions, causing flushing of  $\text{NH}_4^+$ -N with salinization (Duckworth and Cresser, 1991; Compton and Church, 2011; Kim and Koretsky, 2011). For DON releases from stream sediments or soils, it is reasonable to argue that the effects of salinization on DON are similar to those of the protein-like fluorophores for the reasons we discussed previously. However, previous studies have shown no consistent effects of salinization on DON leaching from soils or plant litter (Steele and Aitkenhead-Peterson, 2013; Green et al., 2008, 2009; Compton and Church, 2011). Thus, the increased TKN fluxes from sediments or soils in response to increased salinization (Fig. 4 and 5) were more likely attributed to  $\text{NH}_4^+$  mobilization.

In contrast to DOC, DON, and  $\text{NH}_4^+$ , nitrate is a highly soluble, negatively-charged ion. Mechanisms such as pH suppression,  $\text{Na}^+$  dispersion/exchange, and colloid coagulation do not apply for nitrate to interpret the salinization effects, while biologically-mediated transformations may play a relatively more important role. According to previous studies on soils, salinity can decrease the rates of both nitrification and denitri-

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colloids in estuaries in response to mixing of fresh water with seawater (e.g., Sholkovit, 1976). Thus, increased salinization may decrease stability of the colloidal humic- Fe (Al-) phosphate complexes, leading to reduced releases of SRP from sediments and soils (Figs. 4 and 5) and a coupling between SRP and SUVA (Fig. 6). In addition to abiotic controls, inhibition of microbial activity at higher salinities (Srividya et al., 2009) could provide an alternative explanation for observed decreases in SRP mobilization with salinization.

Relative to C, N and P, effects of salinization on sulfur transformations are relatively less known. Kim and Koretsky (2011, 2013) reported salinization inhibited porewater sulfate reduction in lake sediments. However, our results show that stream sediments were generally a source of sulfate (Figs. 4d and 5d), and thus sulfate reduction seemed to be an unimportant process in free-flowing streams. Effects of increased salinization on sulfate releases warrant further investigation in future studies, however.

### 4.3 Interactive effects of watershed urbanization and salinization

Overall, our results suggest that the effects of increased salinization on sediment DOC, TKN and DIC fluxes, DOC lability (indicated by P/H ratio and SUVA) increased from rural/agricultural, suburban to urban site (Fig. 7). The interactive effects of land use and salinization on sediment fluxes may be explained by coinciding changes in stream sediment organic matter content (indicated by ash free dry weight), which also showed an increase across the rural-urban gradient (Table 2 and Fig. 7a). Higher sediment organic matter content in urban streams has also been observed in our previous work (Duan and Kaushal, 2013), probably due to additional organic inputs from algal and anthropogenic sources (e.g., wastewater). Our recent work (Kaushal et al., 2014b) showed that gross primary production and organic matter lability increased significantly with watershed urbanization. Wastewater inputs from sewer leaks are common in the urban tributaries in the lower Gwynns Falls (DEPRM and Baltimore City Department of Public Works, 2004; Kaushal et al., 2011). As quantity and quality of sediment organic

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matter increase across the rural-urban land use gradient, so did the fluxes of labile DOC, total DOC, TKN and DIC in response to increased salinization.

#### 4.4 Comparison of laboratory salinization experiments to field observations

5 Similar effects of laboratory salinization experiments on DOC and nutrient fluxes from sediments were only partially observed in the field measurements on ambient stream water quality before and after the snow event. The agreements included the consistently higher DOC concentrations and consistently lower nitrate concentrations during post-snow storm conditions than those of pre-snow conditions (Fig. 8c, d, i, j). The difference between laboratory salinization experiments and field observations during the snow event (Figs. 2–5 and 9) may be attributed to the different settings between field processes and laboratory experiments. First of all, the compositions of salts that was used for deicers are different from pure NaCl that was used in our laboratory experiments (Ramakrishna and Viraraghavan, 2005; Shi et al., 2010). The impurities in the deicers may cause increases in pH in soils and sediments – different from pH suppression with pure NaCl. Many studies have observed that soil pH actually increases after use of road deicers or groundwater irrigation (e.g., Aitkenhead-Peterson et al., 10 2009; Galuszka et al., 2011). In this study, consistent increases in SUVA (not shown) and decreases in P/H ratio at some sites during the snow event might be related to an increase in pH with deicer use (Fig. 8). In addition, snow melt might cause elevated leaching of bioreactive elements or “dilution effects”, in addition to increases in salinization (Cameron, 1996; Agren et al., 2012). There are also other major nonpoint sources of organic matter and nutrients (e.g., upland soils and plant litter) from watersheds besides stream sediments and riparian soils (Kaushal et al., 2011), which may respond differently to the episodic salinization. All these uncertainties might contribute to observed differences between laboratory and field observations and should be considered in future research.

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## 5 Conclusions

The potential effects of salinization on biogeochemical fluxes from soils and stream sediments are summarized in Fig. 9. As shown in this figure, releases of labile DOC (thus total DOC) and TKN from sediments can potentially increase during episodic stream salinization, due to “salting-in” effects (or Na dispersion) of proteinaceous organic matter and  $\text{NH}_4^+$  mobilization. The increased releases of labile DOC and TKN can result in increases in sediment releases of DIC and sediment retention of nitrate as a result of organic carbon mineralization and associated N transformations (e.g., denitrification and nitrate microbial immobilization). Moreover, the effects of salinization on sediment releases of labile DOC and TKN also increased with watershed urbanization (indicated by watershed ISC) due to higher sediment organic content at urban sites. DOC aromaticity (indicated by SUVA) and releases of SRP, however, decreased with stream salinization, likely due to coagulation of colloidal humic- Fe (Al-) complexes and pH suppression, which were associated with ion exchange. The sediment releases of labile DOC and TKN with increased salinization probably represents a significant, previously unrecognized flux of labile DOC and nitrogen in urban streams, which might have a large influence on carbon and nutrient biogeochemical cycles and water quality in urban waters. For soils, salinization effects on DOC leaching were not consistent, and there were no interactive effects of land use and salinization. Differences in effects of salinization on sediments and soils are likely attributed to differences in organic matter sources and lability. Nonetheless, our work suggest that increased salinization can have major effects on concentrations and fluxes of bioreactive elements in human-impacted watersheds and streams, and it is critical to conduct comprehensive investigations of the effects of salinization on all major bioreactive elements and also couple them together as a whole.

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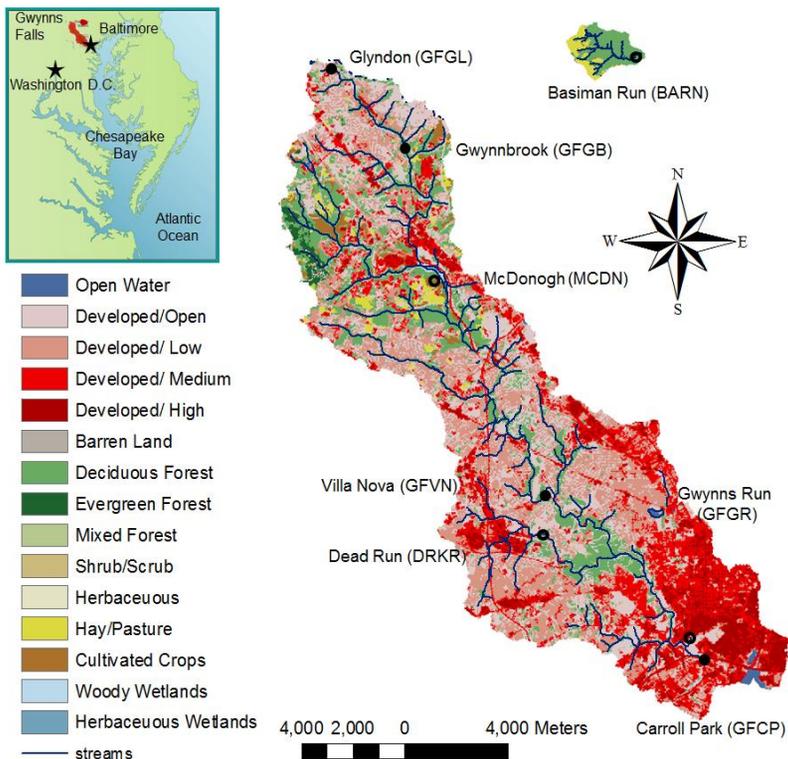
**Table 1.** Characteristics of study subwatersheds.

| Site Type        | BARN forest | MCDN agriculture | GFGB suburban | GFGL suburban | GFVN urban | GFCP urban | DRKR urban | GFGR urban |
|------------------|-------------|------------------|---------------|---------------|------------|------------|------------|------------|
| Area (ha)        | 3.86        | 0.1              | 11            | 0.8           | 84.2       | 170.7      | 14.3       | 6.5        |
| %ISC             | 0.3         | 0.1              | 15            | 19            | 17         | 24         | 45         | 61         |
| Developed/open   | 25.5        | 13.6             | 43.8          | 41.2          | 27.4       | 25.5       | 22.4       | 9.2        |
| Developed/low    |             | 5.4              | 28.9          | 21.5          | 25.2       | 28.8       | 38.8       | 27.5       |
| Developed/medium |             | 3.6              | 4.3           | 8.3           | 10.5       | 16.2       | 18.1       | 43.6       |
| Developed/high   |             |                  | 1.3           | 3.4           | 2.5        | 5.0        | 7.4        | 17.4       |
| Barren           |             |                  |               |               | 0.2        | 0.1        | 0.1        | 0.04       |
| Shrub            | 0.7         | 7.9              | 1.1           | 6.3           | 1.4        | 0.8        | 0.2        | 0.05       |
| %Forest          | 72.6        | 1.4              | 19.0          | 19.2          | 25.5       | 19.3       | 12.7       | 1.3        |
| Hay/pasture      | 1.0         | 30.4             | 0.9           | 0.1           | 3.1        | 1.7        | 0.1        |            |
| Cultivated Crops |             | 37.1             | 0.1           |               | 3.4        | 1.8        |            |            |
| Wetland          |             | 0.7              | 0.6           |               | 0.8        | 0.7        | 0.2        |            |
| Open water       |             |                  |               |               | 0.1        | 0.1        |            | 0.9        |

Watershed land cover and impervious surface (ISC %) data are from Shields et al. (2008) and the National Land Cover Database (NLCD). Both land cover and impervious statistics were based on 30 m resolution land cover data.

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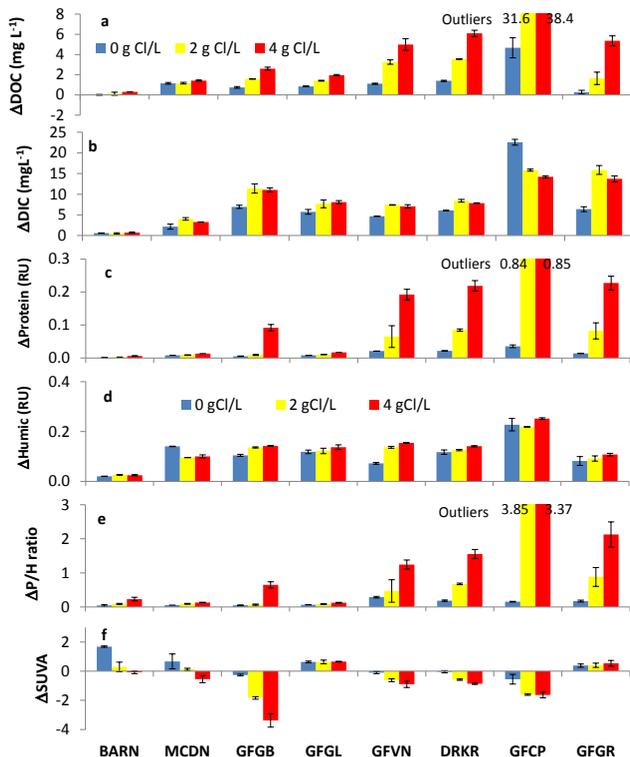
**Figure 1.** Land use of the Gwynns Falls and Baisman Run watersheds, showing sites from which sediment, soil and stream water were collected for salinization experiments. Baisman Run is a watershed with forest as the dominant land use, and it is located in the nearby Gunpowder River. Solid and open circles represent sites of the main stem and tributaries, respectively. Resolution of the land use data is 30 m, and land use and stream channel location data are from US Department of Agriculture ([datagateway.nrcs.usda.gov](http://datagateway.nrcs.usda.gov)) and US Geological Survey (<http://datagateway.nrcs.usda.gov>).

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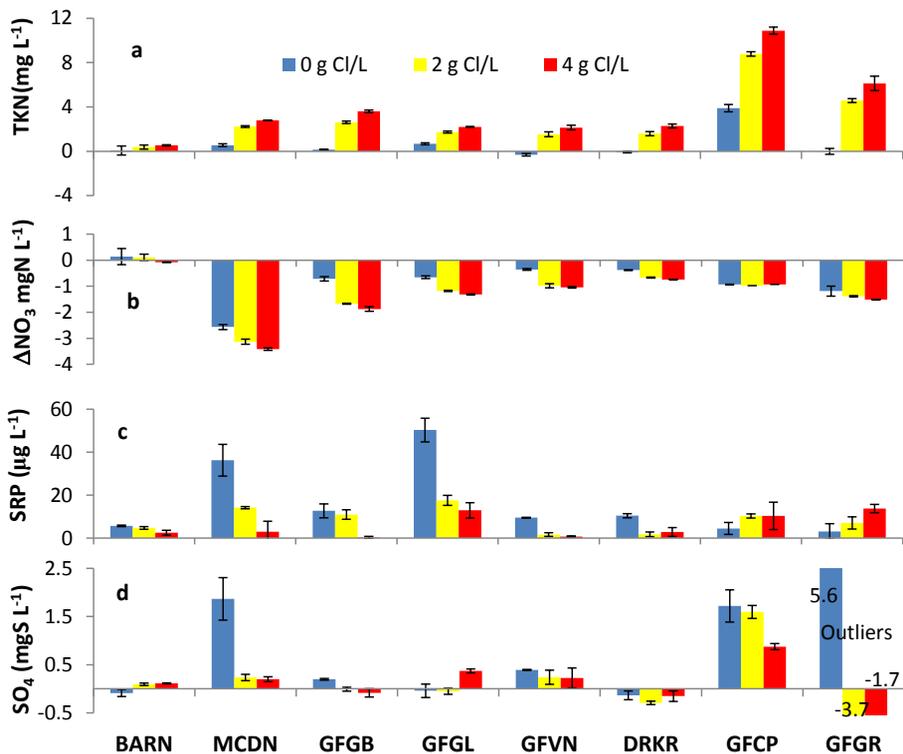
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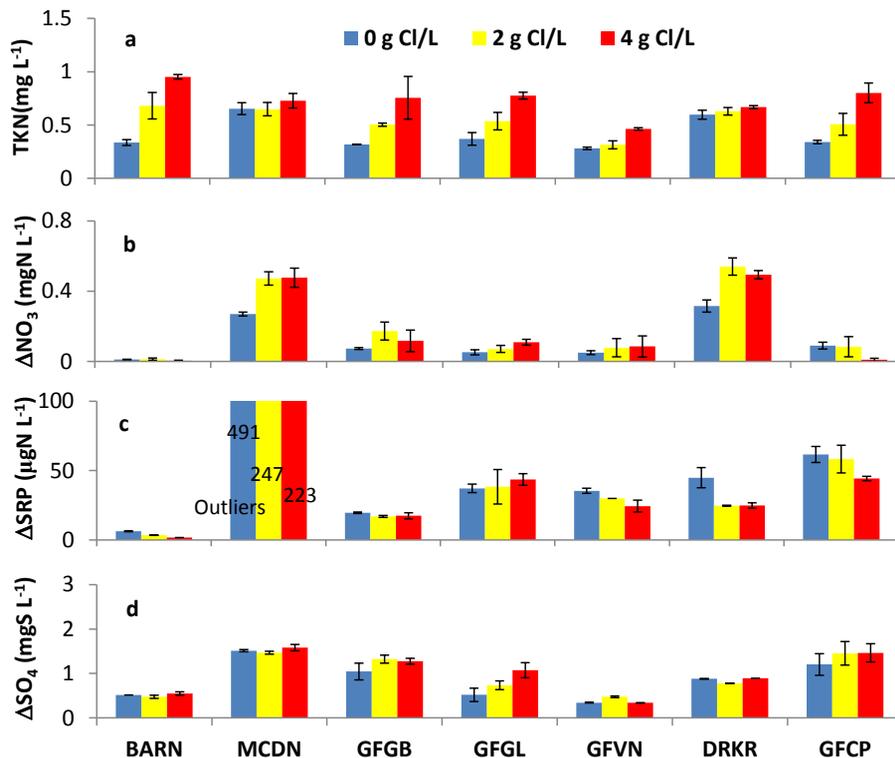


**Figure 2.** Changes in DOC, DIC, protein-like fluorophore, humic-like fluorophore, protein to humic (P/H) ratio and specific ultraviolet absorption (SUVA) during 2 day sediment incubations with NaCl-amended stream water (in 0, 2 and 4 g Cl L<sup>-1</sup>). The changes in controls (stream water only) were subtracted to obtain the contributions from sediments. Errors bars stand for SDs of duplicate incubations, and positive and negative values referred to release and retention, respectively. Humic- and proten-like fluorescence is in Raman Unit (RU).





**Figure 4.** Changes in TKN (DON + NH<sub>3</sub>-N + NH<sub>4</sub><sup>+</sup>-N), nitrate-N, SRP and sulfate during 2 day sediment incubations with NaCl-amended stream water (in 0, 2 and 4 g Cl L<sup>-1</sup>). The changes in controls (stream water only) were subtracted to obtain the contributions from sediments. Errors bars stand for SDs of duplicate incubations, and positive and negative values referred to release and retention, respectively.

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**Figure 5.** Release of TKN (DON + NH<sub>3</sub>-N + NH<sub>4</sub><sup>+</sup>-N), nitrate-N, SRP and sulfate during 2 day soil incubations with NaCl-amended DI water (in 0, 2 and 4 g Cl L<sup>-1</sup>). Errors bars stand for SDs of duplicate incubations.

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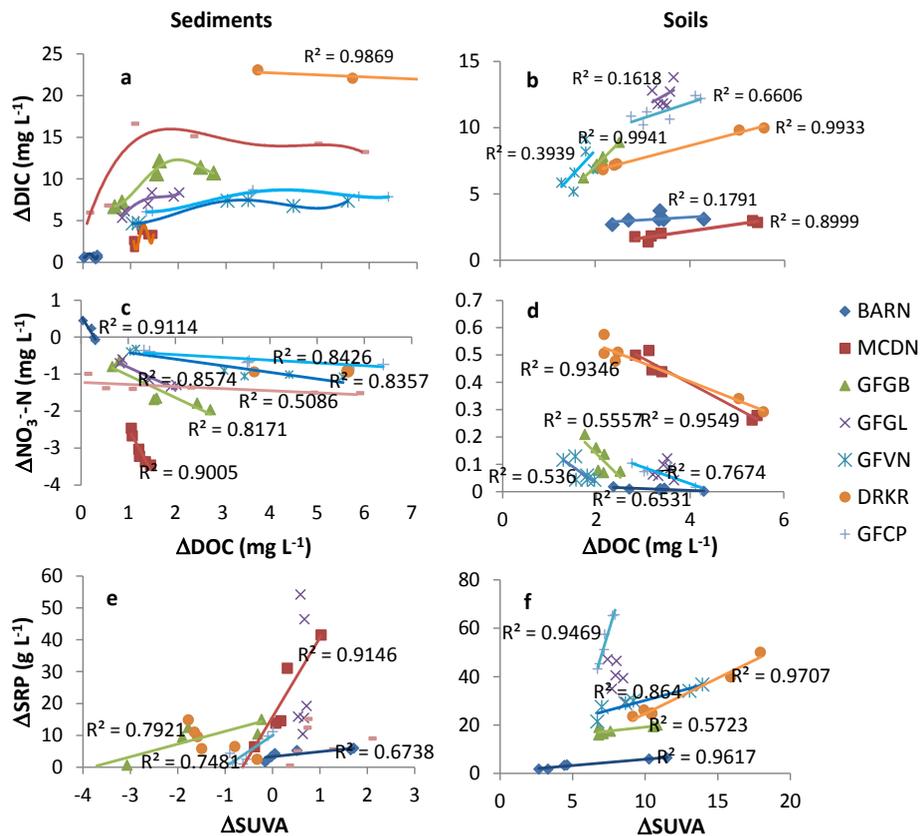
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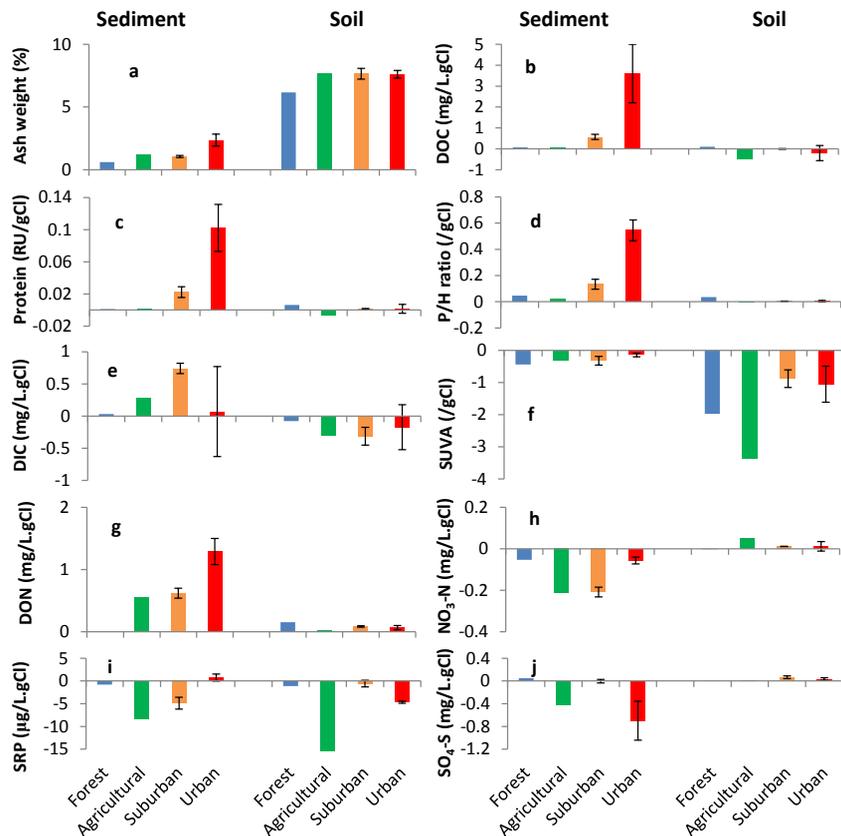
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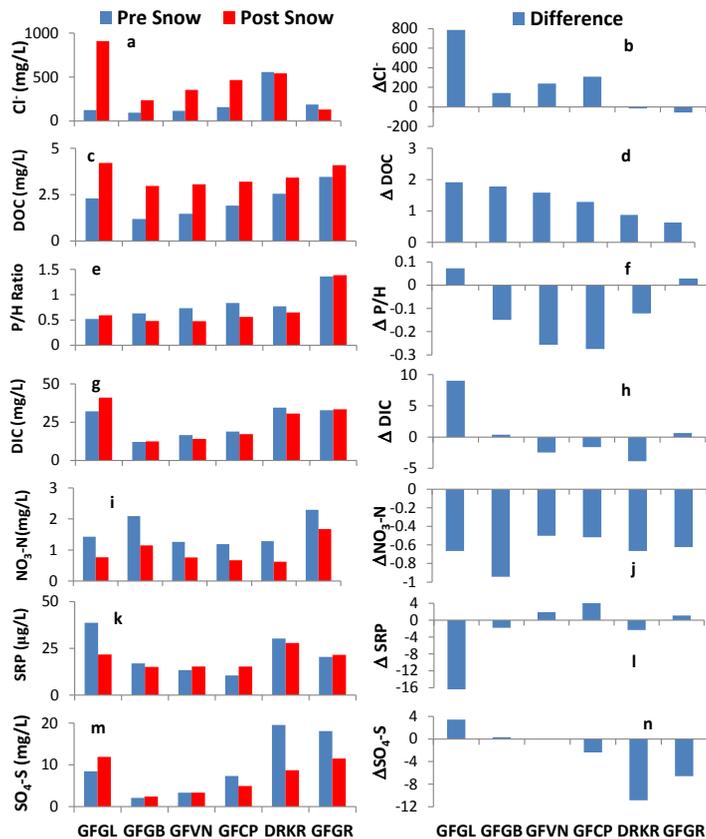
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**Figure 6.** Correlations between changes in DIC or nitrate and changes in DOC, and between changes in SRP and changes in SUVA during the incubations of sediment and soils with NaCl-amended stream or DI water.



**Figure 7.** Ash-free dry weight, and salinity effects on DOC, protein-like fluorophore, P/H ratio, DIC, SUVA, TKN, nitrate, SRP and sulfate for sediment and riparian soils at the forested (1), agricultural (1), suburban (3) and urban (3) sites. Salinity effects were estimated from changes in these variables across three different salinities used for the incubations. Positive and negative values represented retention/release in their standardized fluxes (or increases/decreases in P/H ratio) per g of Cl<sup>-</sup>.



**Figure 8.** Concentrations of  $\text{Cl}^-$ , DOC, DOC P/H ratio, DIC, nitrate, SRP and sulfate and their changes in ambient stream water observed in the field before and 2 days after snow storm event.

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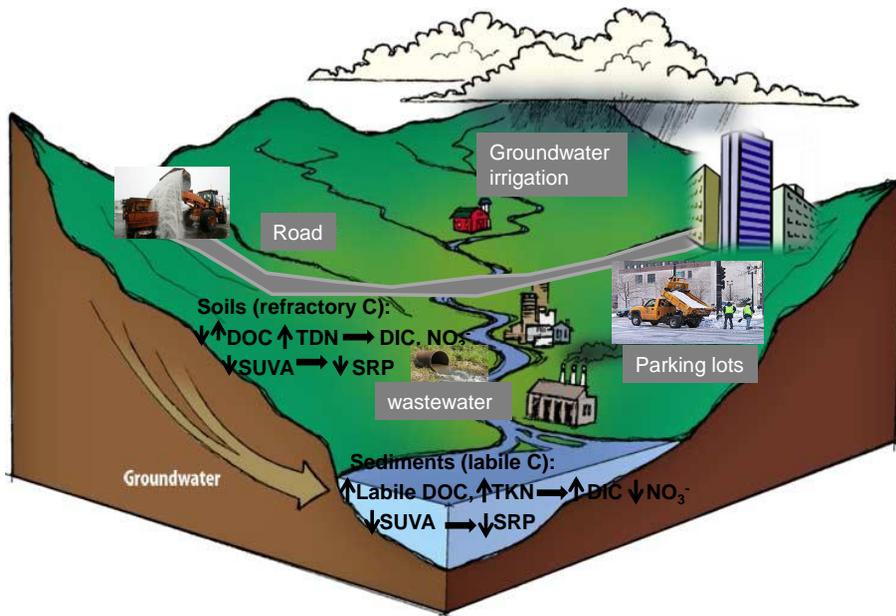
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**Figure 9.** A conceptual diagram summarizing potential effects of salinization on DOC quality, DOC and TKN releases from sediments and soils, as well as linkage to release/retention of DIC, nitrate and SRP during sediment and soil salinization.

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