Seasonal and size-dependent variations in the phytoplankton growth and microzooplankton grazing in the southern South China Sea under the influence of the East Asian monsoon

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Abstract

To examine seasonal and size-dependent variations in the phytoplankton growth and microzooplankton grazing in oligotrophic tropical waters under the influence of seasonal reversing monsoon, dilution experiments were conducted during the summer 2009 (21 May to 9 June) and winter 2010 (9 to 18 November) in the southern South China Sea (SSCS). The results showed that environmental variables, phytoplankton biomass, phytoplankton growth rate ($\mu$), microzooplankton grazing rate ($m$), and correlation (coupling) between the $\mu$ and $m$, but the microzooplankton grazing impact on phytoplankton ($m/\mu$) significantly varied between the two seasons. Higher relative preference index (RPI) for and $m$ on the larger-sized (> 3 µm) phytoplankton than pico-phytoplankton (< 3 µm), indicating significant size-selective grazing by microzooplankton on the larger-sized phytoplankton, were also observed. The $\mu$ and $m$ were significantly correlated with salinity and dissolved inorganic nutrients, which indicated that salient seasonal variations in the phytoplankton growth and microzooplankton grazing in the SSCS were closely related to the environmental variables under the influence of the East Asian monsoon. We propose that intermittent arrivals of the northeast winter monsoon could lead to the low $\mu$ and $m$, and the decoupling between the $\mu$ and $m$ in the SSCS, through influencing nutrient supply to the surface water, and inducing surface seawater salinity decrease. The low $m/\mu$ (< 50% on average) indicates low remineralization of organic matter mediated by microzooplankton and the increased importance of the phytoplankton-mesozooplankton grazing pathway, and thus probably accounts for part of the high vertical biogenic particle fluxes in the prevailing periods of the monsoons in the SSCS. The size-selective grazing suggests that microzooplankton grazing contributes to the pico-phytoplankton dominance in the oligotrophic tropical waters such as that of the SSCS.
1 Introduction

Phytoplankton growth and microzooplankton grazing are crucial processes relating to ocean biogeochemistry and energy flow in the pelagic food web. Phytoplankton growth provides almost all the primary production in the upper ocean. Microzooplankton (< 200 µm) consumes most of the daily primary production in the sea, and regulates phytoplankton community composition, and affects the ultimate fate of the phytoplankton-derived primary production (Banse, 2007, 2013; Landry and Calbet, 2004; Schmoker et al., 2013).

The dilution technique introduced by Landry and Hassett (1982) has extensively been used to estimate phytoplankton growth and synchronous microzooplankton grazing rates in varied marine ecosystems, although later modification and increasing use has also brought scrutiny and critiques (Dolan and Mckeon, 2004; Gallegos, 1989; Gutiérrez-Rodríguez et al., 2009; Teixeira and Figueiras, 2009). Based on global data collection, Schmoker et al. (2013) pointed out that seasonality of environmental variables and phytoplankton community indicates seasonal variations in the phytoplankton growth and microzooplankton grazing in polar and sub-polar, temperate regions, and tropical waters with salient seasonal reversing monsoon. However, there are few studies investigating both the phytoplankton growth and microzooplankton grazing in tropical oceans (Caron and Dennett, 1999; Chen et al., 2013; Landry et al., 1998, 2011), and the seasonality of phytoplankton growth and microzooplankton grazing in tropical waters influenced by salient seasonal reversing monsoon is still less known.

Microzooplankton may modify their food preference depending on the morphology, abundance and quality of prey, which would cause phytoplankton community shift in species and/or size composition (Teixeira and Figueiras, 2009). Phytoplankton community in oligotrophic subtropical and tropical waters is usually dominated by small-sized pico-cells (< 3 µm); the pico-phytoplankton consists of three major groups including Prochlorococcus, Synechococcus and diverse types of pico-eukaryotic phytoplankton (Zubkov et al., 2000). Microzooplankton including both autotrophic and het-
erotrophic nanoflagellates can actively consume pico-phytoplankton-sized prey (An-Yi et al., 2009; Christaki et al., 2005; Frias-Lopez et al., 2009). Landry et al. (1998) proposed that the balance between microzooplankton grazing and phytoplankton growth was consistent with the dominance of pico-plankton in oligotrophic offshore regions in the Arabian Sea. Significantly higher microzooplankton grazing rates on the large-sized phytoplankton (> 5 μm) than the smaller one (< 5 μm) have been recently reported in the oligotrophic subtropical Northeast Atlantic (Cáceres et al., 2013). Our recent results suggest that size-selective grazing by microzooplankton on nano-sized (3–20 μm) phytoplankton contributes to the pico-phytoplankton dominance in the oligotrophic tropical waters of the South China Sea (SCS) in summer (Zhou et al., 2015a). However, until now, the role played by microzooplankton in the pico-phytoplankton dominance in oligotrophic tropical waters is still less examined.

The southern SCS (SSCS) is characterized with permanent water stratification and oligotrophic conditions in the upper layer, and is affected by seasonal reversing monsoon. During the middle of May to September, the SSCS is under the influence of the southwest summer monsoon, while this area is influenced by the stronger northeast winter monsoon during November to the next April (Su, 2004). Mesoscale eddies with obvious seasonal variation (Fang et al., 2002, 1998; Zhang et al., 2014), and seasonal pattern of higher phytoplankton biomass, primary production and vertical biogenic particle fluxes occur during the prevailing periods of the monsoons and wintertime in the SSCS (Liu et al., 2002; Ning et al., 2004; Wan et al., 2010). There are hundreds of coral shoals, reefs and islands located in the SSCS, called the Nansha Islands. Many researchers including Qingchao Chen, Liangmin Huang and their co-workers, from the South China Sea Institute of Oceanology, Chinese Academy of Sciences, have previously investigated the environment variables, marine species diversity and ecological processes in the coral reef lagoons of the Nansha Islands and its adjacent waters during the 1980s to 1990s (e.g. Chen and Mutidisciplinary Expedition to Nansha Islands, 1991, 1989, 1994, 1996, 1998; Huang and Multidisciplinary Expedition to Nansha Islands, 1997). Their works provide valuable contributions to the understanding
of the taxonomic composition and distribution of phytoplankton, bacteria, zooplankton and fish, and ecological processes such as primary production in the SSCS. However, these results are seldom published in international media even scientific journals in Chinese, and thus less known to the scientific community. So far, there is no data reported on the microzooplankton grazing in this region. We hypothesize that seasonal changes in both the phytoplankton growth and microzooplankton grazing is expected in the tropical waters influenced by salient seasonal reversing monsoon, and microzooplankton contributes to the phytoplankton size composition through size-selective grazing in the SSCS.

To test the hypotheses, we conducted a series of dilution experiments in adjacent waters of the Nansha Islands in the SSCS during two cruises in May to June 2009 and November 2010. Significant seasonal variations in the phytoplankton growth and microzooplankton grazing and apparently size-selective grazing were observed.

2 Material and methods

Dilution experiments (Landry and Hassett, 1982) were conducted at 14 stations during 21 May to 9 June 2009 (summer), and at 10 stations during 9 November to 18 November 2010 (winter). Stations KJ28 and KJ32 were located at the basin waters northwest to the Nansha Islands; station YS was located at the lagoon of the Yongshu Reef, the other stations were distributed in waters around the Nansha Islands (Fig. 1).

Surface seawater was collected, and pre-screened with a 200 µm nylon netting for dilution experiments at each station. Particle-free seawater was obtained by filtering the seawater through a filter with a pore size of 0.22 µm. All the bottles, containers and filters were soaked in 10 % HCl for more than 10 h, and thoroughly washed with deionized water and MiliQ-water before each cruise. The bottles and containers were washed with 10 % HCl, deionized water and ambient seawater before each experiment. Measured amount of particle-free seawater was firstly added to the 2.4 L polycarbonate bottles, and unfiltered seawater was added and filled the bottles. Four dilution treatments of
25, 50, 75 and 100 % unfiltered seawater were prepared for the summer experiments, and another four dilution treatments of 37.5, 58.3, 79.2 and 100 % unfiltered seawater were prepared for the winter experiments. All the bottles were enriched with additional nutrients (final concentrations of 0.5 µM NH₄Cl, 0.03 µM KH₂PO₄, 1.0 nM FeSO₄ and 0.1 nM MnCl₂) to promote constant phytoplankton growth. Two bottles were filled with unfiltered seawater without nutrient enrichment served as no nutrient controls. Another two bottles filled with unfiltered seawater were sacrificed for initial samples of chlorophyll a (Chl a). All of the bottles were incubated for 24 h in a deck incubator cooled by running surface seawater and covered with neutral-density screens to simulate in situ light regime. These measures have been proved effective to avoid phytoplankton photoacclimation during the incubation (Zhou et al., 2015a).

Seawater was filtered through a sequence of 3 µm pore size polycarbonate filter and GF/F filter for size-fractionated Chl a of the larger-sized (> 3 µm) and pico- (< 3 µm) phytoplankton. Total Chl a was calculated as the sum of the two size fractions or directly sampled by filtering 0.5 to 1 L seawater on the GF/F filter. The filters were extracted in 90 % acetone at −20 °C for 24 h. The Chl a concentration was measured by fluorometry using a Turner Designs Model 10 Fluorometer (Parsons et al., 1984).

Seawater temperature, salinity and dissolved inorganic nutrients were also measured. Temperature and salinity were determined by Conductivity–Temperature–Depth probes. Dissolved inorganic nutrients such as silicate, phosphate and nitrate plus nitrite were analyzed with a flow-injection autoanalyzer (Quickchem 8500, Lachat Instruments) following the standard manuals.

Assuming an exponential growth model, the net growth rate \( k_d \) of phytoplankton in a dilution treatment was calculated according to the formula \( k_d = \ln(P_t/dP_0) \), where \( d \) is the dilution factor (the proportion of unfiltered seawater), \( P_t \) is the Chl a concentration after incubation, \( P_0 \) is the initial Chl a concentration. Phytoplankton growth rates with nutrient amendment (\( \mu_n \)) and microzooplankton grazing rates (\( m \)) were estimated from Model I linear regressions of net growth rate (\( k \)) vs. dilution factor (\( d \)). In situ phy-
toplankton instantaneous growth rates ($\mu$) were calculated as the sum of $m$ and net growth rate in control bottles without added nutrients.

When saturated or saturated-increasing grazing was observed as a departure from the assumed linear model (Gallegos, 1989; Teixeira and Figueiras, 2009), $m$ and $\mu$ were calculated on the basis of the method of Chen et al. (2009a).

Size-fractionated (> 3 and < 3 µm) phytoplankton growth and mortality rates were estimated for experiments at YS and KJ35 during the summer cruise, and all the experiments during the winter cruise.

Nutrient limitation index indicating nutrient sufficiency for phytoplankton growth was assessed by the ratio $\mu/\mu_n$, which is expected to vary with system trophic state (Landry et al., 1998). Net growth rate was also used as a proxy for the actual trophic state of the system being investigated (Calbet et al., 2011). The microzooplankton grazing impact on phytoplankton was assessed by the ratio of microzooplankton grazing to phytoplankton growth rates ($m/\mu$), and measures the extent to which the daily phytoplankton production is consumed and balanced by microzooplankton grazing (Landry et al., 1998).

Grazing selectivity for size-fractionated phytoplankton was analyzed following the relative preference index (RPI) provided by Obayashi and Tanoue (2002) as $\text{RPI} = \frac{\sum [m]\text{chla}}{\sum [\text{chla}]},$ where $[m]$ is the amount of daily grazed Chl a ($\mu$g L$^{-1}$ d$^{-1}$), subscript $i$ refers to each size fraction analyzed and $[\text{chla}]$ is the concentration of Chl a ($\mu$g L$^{-1}$). $\text{RPI} > 1$ indicates positive selection and vice versa.

All the statistical analyses were conducted by using the SPSS 17.0. The average value of the ratio variables such as $m/\mu$ was presented as the geometric mean, while other average values were presented as the arithmetic mean.
3 Results

3.1 Environmental variables and Chl a

Environmental variables and Chl a in surface seawater were different between the two cruises (Tables 1 and 2). Surface seawater temperature (SST) and salinity (SSS), and the concentration of nitrate plus nitrite were significantly lower (independent t test, \( p < 0.05 \) or 0.01), while the concentrations of silicate and phosphate were significantly higher in the winter cruise (independent t test, \( p < 0.05 \) or 0.01) than those in the summer cruise. The concentration of phosphate (mean ± sd: 0.04 ± 0.03 µmolL\(^{-1}\)) during the summer cruise and that of nitrate plus nitrite (mean ± sd: 0.04 ± 0.02 µmolL\(^{-1}\)) during the winter cruise were almost undetectable. The concentration of Chl a in the winter cruise (mean ± sd: 0.104±0.024 µgL\(^{-1}\)) was about two times that in the summer cruise (mean ± sd: 0.066 ± 0.022 µgL\(^{-1}\)) (independent t test, \( p < 0.01 \)). Pico-phytoplankton (< 3 µm) accounted for most (> 80 %) of the total Chl a during both the cruises. The proportion of pico-phytoplankton in the winter cruise (mean ± sd: 81.9 ± 5.0%) was similar to that in the summer cruise (mean ± sd: 83.0 ± 1.8%) (independent t test, \( p > 0.1 \)).

3.2 Feeding responses in dilution experiments

The detailed results of each dilution experiment were listed in Tables 1 and 2. Except for linear feeding responses, both the saturated and saturated-increased feeding responses described by Teixeira and Figueiras (2009) occurred in the dilution experiments during both the cruises (Fig. 2a–c). Non-linear feeding responses occurred at eight of the 14 experiments during the summer cruise, while those occurred at four of the 10 experiments during the winter cruise (Tables 1 and 2). During the winter cruise, substantially negative phytoplankton growth rates \( (\mu) \) and high microzooplankton grazing rates \( (m) \) were observed at KJ53 (Fig. 2d–f). The negative \( \mu \) obtained at KJ53 and the derivative parameters (e.g. \( m/\mu \)) were not included for the comparison between seasons and size fractions.
3.3 Comparison of the phytoplankton growth and microzooplankton grazing between the two seasons

The phytoplankton growth and microzooplankton grazing were significantly different between the two cruises. The $\mu$ in the winter cruise (mean ± sd: 0.54 ± 0.22 d$^{-1}$) were significantly lower than those in the summer cruise (mean ± sd: 0.92 ± 0.32 d$^{-1}$) (independent $t$ test, $p < 0.01$). The $m$ in the winter cruise (mean ± sd: 0.27 ± 0.13 d$^{-1}$) were also significantly lower than those in the summer cruise (mean ± sd: 0.46±0.20 d$^{-1}$) (independent $t$ test, $p < 0.01$). However, the $m/\mu$ ratio was almost equally low both in the summer (geometric mean ± sd: 49±23%) and winter (geometric mean ± sd: 48±33%) cruises.

3.4 Size-dependent phytoplankton growth and microzooplankton grazing

The $\mu$ and $m$ varied between size fractions. Higher $m$ on the larger size fraction were observed at YS (0.65 d$^{-1}$ for the size fraction > 3 µm vs. 0.33 d$^{-1}$ for the size fraction < 3 µm) and KJ35 (0.39 d$^{-1}$ for the size fraction > 3 µm vs. 0.30 d$^{-1}$ for the size fraction < 3 µm) during the summer cruise. The $\mu$ of the larger-sized (> 3 µm) and pico- (< 3 µm) phytoplankton were 1.07 and 0.75 d$^{-1}$ at YS, and 0.69 and 0.90 d$^{-1}$ at KJ35, respectively. During the winter cruise, the $m$ on the larger size fraction (mean ± sd: 0.50±0.41 d$^{-1}$) were higher than those on pico-phytoplankton (mean ± sd: 0.27 ± 0.27 d$^{-1}$) at seven of the 10 experiments, and the $\mu$ of the larger size fraction (mean ± sd: 0.88 ± 0.38 d$^{-1}$) were also higher than those of pico-phytoplankton (mean ± sd: 0.42 ± 0.31 d$^{-1}$) at seven of the nine experiments (exclude data at KJ53) (Table 3). The $m/\mu$ was higher for the larger size fraction (57.3 % at KJ35 and 61.2 % at YS) than pico-phytoplankton (33.3 % at KJ35 and 44.0 % at YS) during the summer cruise, while the $m/\mu$ was not significantly different between the two size fractions (41.1±82.8 % for the size fraction > 3 µm, and 39.4±21.1 % for the size fraction < 3 µm) during the winter cruise (Table 3).
Higher RPI for the larger-sized phytoplankton than pico-phytoplankton was observed during both the cruises (Fig. 3). The RPI for the larger-sized phytoplankton was higher than one (geomean ± sd: 1.44 ± 0.31), while the RPI for pico-phytoplankton was lower than one (geomean ± sd: 0.90 ± 0.07) during the summer cruise. The RPI was significantly (Mann–Whitney test, p < 0.05) higher for the larger-sized phytoplankton (geomean ± sd: 1.44 ± 1.57) than pico-phytoplankton (geomean ± sd: 0.61 ± 0.35) during the winter cruise.

3.5 Nutrient limitation to the phytoplankton growth

Nutrient limitation index (\(\mu/\mu_n\)), as well as the net growth rate (\(\mu - m\), data not shown), was significantly higher (Mann–Whitney test, \(p < 0.05\)) during the summer cruise (geometric mean ± sd: 0.80 ± 0.25) than the winter cruise (geometric mean ± sd: 0.54 ± 0.30). The \(\mu/\mu_n\) was approximate to or larger than one at five of the 14 experiments in the summer. In contrast, the \(\mu/\mu_n\) was apparently lower than one at eight of the nine experiments (exclude data at KJ53) in the winter (Tables 1 and 2).

3.6 Correlations between the growth and grazing rates with environmental variables

Taken all the data from the two cruises together, the average \(\mu\) and \(m\) were 0.77 ± 0.34 d\(^{-1}\) and 0.39 ± 0.20 d\(^{-1}\). The \(\mu\) were positively correlated with SST (\(r = 0.43\), \(p < 0.05\)), SSS (\(r = 0.55\), \(p < 0.01\)), \(\mu/\mu_n\) (\(r = 0.50\), \(p < 0.05\)), net phytoplankton growth rate (\(r = 0.83\), \(p < 0.01\)) and \(m\) (\(r = 0.76\), \(p < 0.01\)), but were negatively correlated with phosphate (\(r = −0.57\), \(p < 0.01\)) and Chl a (\(r = −0.65\), \(p < 0.01\)) (Table 4). The \(m\) were also positively correlated with SST (\(r = 0.62\), \(p < 0.01\)), SSS (\(r = 0.48\), \(p < 0.05\)) and nitrate plus nitrite (\(r = 0.54\), \(p < 0.01\)), and was negatively correlated with Chl a (\(r = −0.66\), \(p < 0.01\)) and phosphate (\(r = −0.54\), \(p < 0.01\)) (Table 4).
3.7 Correlations between the phytoplankton growth and microzooplankton grazing

The positive correlation ($r^2 = 0.57$, $p < 0.01$) between $\mu$ and $m$ was observed during the summer cruise (Fig. 4a). However, there was no significant correlation between the total $\mu$ and $m$ during the winter cruise. By grouping $\mu$ and $m$ separately for each size fraction, it is found that the correlation was significant for the larger size fraction ($r^2 = 0.84$, $p < 0.05$) but not for the pico-phytoplankton ($r^2 = 0.41$, $p = 0.07$) (Fig. 4b).

4 Discussion

4.1 Comparisons with other studies

All the $\mu$ (except those obtained at KJ53 during the winter cruise) and $m$ were within the reviewed ranges based on global data collection (Calbet and Landry, 2004; Schmoker et al., 2013). Substantive $m$ but negative $\mu$ were observed at KJ53 (Fig. 2d–f), the only station located on the continental shelf with a bottom depth less than 200 m (Fig. 1).

4.1.1 Negative phytoplankton growth rates

Negative $\mu$ have been extensively reported in previous dilution experiments (e.g. Burkill et al., 1987; Loebl and Beusekon, 2008; Suzuki et al., 2002; Zhou et al., 2011). Zhou et al. (2013) reviewed that temperature and light regime during incubation, insufficient sampling, contamination of particle-free water and the added nutrients, and decay of phytoplankton blooms could be the possible reasons for the negative $\mu$ in dilution experiments. Our measures to mimic the in situ light and temperature during incubation exclude light and temperature from the factors for the substantially negative $\mu$. Insufficient sampling also unlikely contributed to the negative $\mu$ since sufficient mixing was conducted before Chl a sampling. The substantive $m$ could exclude contamination as the reason for the negative rates, because contamination could not only cause phy-
topplankton death, but also less microzooplankton grazing. We conjecture that phytoplankton community decay may occur and lead to the negative $\mu$ at KJ53. The lowest silicate concentration at KJ53 may be related to the negative $\mu$. Why the lowest silicate concentration occurred and to what extent the low nutrient condition was related to the negative phytoplankton growth is a topic that needs further assessment.

4.1.2 Non-linear feeding responses

Non-linear feeding responses including both the saturated and saturated-increased types occurred in our dilution experiments. Non-linear responses were usually observed in eutrophic waters with high prey abundance (Elser and Frees, 1995; Gallegos, 1989; Teixeira and Figueiras, 2009). Non-linear feeding responses in dilution experiments conducted in the oligotrophic subtropical Northeast Atlantic in summer have also been reported by Quevedo and Anadón (2001) and Cáceres et al. (2013). However, the authors did not explain the underlying reasons for these phenomena. The oligotrophic conditions and low phytoplankton biomass in their study area were similar to those in the oligotrophic tropical waters of the SSCS. Teixeira and Figueiras (2009) proposed that changes in the specific phytoplankton growth rate due to varied nutrient limitation in experimental bottles, in the morality rate related to microzooplankton abundance and feeding behavior and even virus infection could be related to the non-linear responses. As additional nutrients were added in the experiments bottles, we do not think that nutrient limitation was the factor causing the non-linear responses in our experiments. Rather we think that the sufficient nutrients added in the experimental bottles led to high phytoplankton abundance in the less diluted bottles. Microzooplankton may reach a maximum ingestion rate at high food concentration, and the maximum ingestion rate may remain constant despite further increase in prey abundance, which is often used to explain the occurrence of saturated feeding responses in dilution experiments (Gallegos, 1989; Moigis, 2006; Teixeira and Figueiras, 2009), and could explain those in our experiments. While the occurrence of saturated-increased responses implies that decrease in the ingestion rate should be occurred to the further increase in
food availability. There is no concluded explanation for the decrease in the ingestion rate. Teixeira and Figueira (2009) proposed that prey selection by microzooplankton in waters with high and diverse food abundance may account for the decrease. We consider that nutrient amendment in the experimental bottles may give rise to relatively higher phytoplankton (food) abundance, leading to decrease in the ingestion rate and accounting for the occurrence of saturated-increased responses in our experiments.

4.1.3 Phytoplankton growth and microzooplankton grazing rates in low latitude tropical waters

There is no study on microzooplankton grazing in the SSCS, especially in waters around the Nansha Islands. If any, Chen et al. (2009b) reported an average $\mu$ of $0.75 \pm 0.62$ d$^{-1}$ and an average $m$ of $0.65 \pm 0.51$ d$^{-1}$ in the western SCS northwest to the present study waters in summer. These rates were similar to our results observed during the summer cruise (Table 5).

There are few studies on microzooplankton grazing in low latitude tropical waters such as the SSCS. Landry et al. (2011) reported a low average $\mu$ of $0.43 \pm 0.14$ d$^{-1}$ and $m$ of $0.31 \pm 0.11$ d$^{-1}$ in the high-nutrient-low-chlorophyll (HNLC) equatorial Pacific Ocean. These rates are similar to the rates observed during the winter cruise in the present study. Yang et al. (2004) investigated the phytoplankton growth and microzooplankton grazing in the western Pacific waters with latitudes similar to those of the SSCS. Their reported $\mu$ and $m$ in summer were $0.35$ to $0.75$ d$^{-1}$, and $0.51$ to $0.67$ d$^{-1}$, which are similar to our results observed during the summer cruise. Landry et al. (1998) showed an average $\mu$ of $0.5$ d$^{-1}$ and an average $m$ of $0.6$ d$^{-1}$ at the oligotrophic stations in the subtropical and tropical Arabian Sea. Caron and Denett (1999) reported the $m$ of $0.35 \pm 0.18$ d$^{-1}$ and $0.30 \pm 0.17$ d$^{-1}$ during the northeast monsoon season and spring intermonsoon season in the Arabian Sea, respectively. The $m$ were similar to the low $m$ ($0.27 \pm 0.13$ d$^{-1}$) observed during the northeast monsoon season in the present study.

The growth and grazing rates of pico-phytoplankton estimated in the present study fall into the middle range of those rates of the main pico-phytoplankton groups
(Prochlorococcus, Synechococcus and eukaryotic pico-phytoplankton) reported in previous studies (reviewed data in Table II in Hirose et al., 2008). Based on cell cycle analysis, Yang and Jiao (2002) reported the in situ Prochlorococcus growth rate of 0.54 d\(^{-1}\) at the chlorophyll maximum layer in the SSCS in May. This rate is lower than the pico-phytoplankton growth rate observed (at YS and KJ35) during the summer cruise, but slightly higher than that (0.42 d\(^{-1}\) on average) observed during the winter cruise. Lower average growth rate (0.15 d\(^{-1}\)) of Prochlorococcus in the subtropical and tropical Atlantic has been reported by Zubkov et al. (2000).

4.2 Seasonal variations in phytoplankton growth and microzooplankton grazing

Our results showed pronounced seasonal changes in the phytoplankton growth and microzooplankton grazing, but not the microzooplankton grazing impact on phytoplankton (\(m/\mu\)), in the SSCS. Many studies have found pronounced seasonality in the phytoplankton growth and microzooplankton grazing in temperate and high latitude waters (e.g. Calbet et al., 2008; Kim et al., 2007; Lawrence and Menden-Deuer, 2012; Loebl and Beusekon, 2008). Nevertheless, there are few studies on the seasonality of the microzooplankton grazing in oligotrophic tropical waters. From the perspective of phytoplankton growth, Landry et al. (2011) reported that \(\mu\) was slightly higher in December than September in the HNLC waters of the equatorial Pacific, but the rate was not significantly different between periods of the southwest monsoon and the early northeast monsoon in the Arabian Sea (Landry et al., 1998); while Caron and Dennett (1999) demonstrated that \(\mu\) was approximately twice as high during the northeast monsoon season than the spring intermonsoon period in the Arabian Sea. Chen et al. (2013) observed that \(\mu\) in surface water of the northern SCS was significantly higher in the summer than winter. However, the previous studies showed no seasonal variation in microzooplankton grazing in the HNLC waters of the equatorial Pacific (Landry et al., 2011), the Arabian Sea (Caron and Dennett, 1999; Landry et al., 1998) and oligotrophic surface waters of the northern SCS (Chen et al., 2013). Significantly lower \(m\) in the
summer than winter at the chlorophyll maximum layer in the northern SCS has been reported by Chen et al. (2013). From the data published by Quevedo and Anadón (2001), we calculated that the $\mu$ and $m$ were higher in the summer ($0.61 \pm 0.43 \text{d}^{-1}$ and $0.49 \pm 0.28 \text{d}^{-1}$) than in the spring ($0.33 \pm 0.27 \text{d}^{-1}$ and $0.29 \pm 0.18 \text{d}^{-1}$) in the oligotrophic subtropical Northeast Atlantic. Lower average $\mu$ and $m$ for pico-phytoplankton in the winter than summer have been recently reported in the subtropical East China Sea (Guo et al., 2014). These results are consistent with the significant seasonal variations in the phytoplankton growth and microzooplankton grazing in the SCS in the present study.

4.3 East Asian monsoon influencing seasonal variations in phytoplankton growth and microzooplankton grazing by affecting nutrient supply and salinity decrease in surface water

Seasonal variations in environmental variables, related to the East Asian monsoon, may directly and/or indirectly influence phytoplankton growth and microzooplankton grazing. Significant seasonal changes in a host of environmental variables including salinity, dissolved inorganic nutrients, and phytoplankton biomass were indeed observed (Table 5). The correlation between $\mu$ and $m$ with the environmental variables indicate that seasonal variations in the phytoplankton growth and microzooplankton grazing were related to the contrasting environmental conditions under the influence of the East Asian monsoon (Table 4).

Many studies have showed increased $m$ during the warmest seasons and reduced $m$ and $\mu$ in winter months (e.g. Ara and Hiromi, 2009; Lawrence and Menden-Deuer, 2012). Both the $\mu$ and $m$ were positively correlated with temperature in the present study. However, the mean seasonal temperature discrepancy was less than 0.4 °C and not significant (independent $t$ test, $p = 0.055$) (Table 5). We do not think this slightly temperature variation alone could account for the substantial decrease in the $\mu$ and $m$ observed in the northeast monsoon season. Substantive $m$ at near freezing temperature in the Arctic Ocean have been reported (Franzè and Lavrentyev, 2014; Sherr
et al., 2013, 2009). This suggested that the physiological effect of temperature was less important for the $m$ variation in the Arctic Ocean, rather than in the tropical waters of the SCS.

We think that changes in dissolved inorganic nutrients and other factors associated with SSS may be the main divers for the variations in the phytoplankton growth and microzooplankton grazing observed in the present study. The significantly higher concentration of dissolved inorganic nutrients such as silicate and phosphate could support the nearly twice as high Chl $a$ concentration during the northeast monsoon season than the summer. The higher concentrations of silicate, phosphate and Chl $a$ are consistent with previous results of relatively higher dissolved nutrients, phytoplankton biomass and primary production in surface waters of the SCS in winter (Liu et al., 2002; Ning et al., 2004). It is easy to understand that the stronger northeast monsoon could increase nutrient supply to the surface layer by enhancing vertical mixing and basin-scale uplift of nutricline depth (Liu et al., 2002). However, this kind of nutrient supply in the SCS could be episodically influenced by intermittent arrivals of the strong northeast monsoon in the form of strong cold air. Therefore, we infer that nutrient supply to the surface water under the influence of the northeast monsoon may stimulate sporadic enhancement in phytoplankton production and biomass.

The arrival of strong northeast monsoon may bring not only nutrient supply to the surface water as a result of enhanced vertical mixing, but also SSS decrease due to heavy frontal rainfall when the cold air meet the warm and wet local air. A strong rainy season is usually developed from October to January in the SCS (Wyrtki, 1961). The remote sensing data from the Tropical Rainfall Measuring Mission indeed showed that apparently larger rainfall occurred during the winter than summer (Fig. 5). The large rainfall could dilute and freshen the surface seawater (as those observed during the winter cruise). In addition, after the passage of the strong cold air, the winds over the SCS are always weak. The low-wind condition could facilitate the less saline surface water overlying on the more saline deep water to form the “freshwater cap” (Zeng et al., 2009). The salinity discrepancy between surface water and that at 25 m in depth was
several times higher during the winter cruise (0.99) than the summer cruise (0.17) (Tables 1 and 2), indicating that a freshwater cap formed during the winter cruise in the SSCS.

The freshwater cap could enhance water stratification and block nutrient supply to the surface layer, thus nutrients such as nitrogen in surface water could be depleted by the phytoplankton community, and lead to nutrient limitation to the growth of the phytoplankton community with already increased biomass. The high $\mu/\mu_n$ (approximate to or higher than one) indicated that phytoplankton growth was less even not nutrient-limited during the summer cruise (Table 1). Similar results in the oligotrophic subtropical Northeast Atlantic have also been reported (Cáceres et al., 2013; Quevedo and Anadón, 2001). In contrast, extremely lower concentration of nitrate plus nitrite and severe nutrient limitation indicated by the lower $\mu/\mu_n$ (0.54 ± 0.30) were observed during the winter, which may account for part of the low $\mu$ (Table 5).

Except for impeding phytoplankton accesses to nutrients, other factors associated with the SSS decrease may impact phytoplankton growth. Previous studies showed that salinity played an important role in the pico-phytoplankton distribution in estuaries (e.g. Mitbavkar et al., 2012). Prochlorococcus has been found seldom occurred in less saline seawaters (Partensky et al., 1999). Guo et al. (2014) reported that salinity was positively correlated with Prochlorococcus abundance in summer, and with abundances of all the main pico-phytoplankton groups (Prochlorococcus, Synechococcus and eukaryotic pico-phytoplankton) in winter, but was negatively correlated with the growth rates of Synechococcus and eukaryotic pico-phytoplankton, across the salinity gradient from the inshore to the offshore regions in the subtropical East China Sea. Dissolved nutrients co-varied with salinity were suggested as the reason for the correlation. This could be true in the reported areas with significant spatial salinity gradients caused by river plume, as low salinity means high nutrients supply from the river plume. The correlation between SSS and dissolved inorganic nutrients in the present study may to some degree support this argument (Table 4). In addition, trace metals from wet atmospheric deposition (by rainfall) could be associated with the rainfall-induced
SSS decrease, and toxic to the pico-phytoplankton growth (Paytan et al., 2009), and lead to the lower $\mu$ of pico-phytoplankton than that of the larger-sized phytoplankton during the winter.

However, the positive correlation between phytoplankton growth rates and SSS, but negative correlation of SSS with silicate and phosphate in the present study may also suggest alternative explanation. Salinity has been found the major environmental determinant of microbial community (including the cyanobacteria) composition in the global level (Lozupone and Knight, 2007). Fu and Bell (2003) demonstrated that low salinity was harmful to the growth, Chl $a$ content, nitrogen fixation and alkaline phosphatase activity of the cyanobacteria *Trichodesmium*. We speculate that low salinity may also go against the growth of other cyanobacteria such as *Prochlorococcus* and *Synechococcus* in the oligotrophic seawater in the SCS, although there is little (if any) data examining the salinity impact on pico-phytoplankton growth, and thus lead to the lower $\mu$ of pico-phytoplankton during the winter.

The freshwater cap could also impact the microzooplankton grazing indirectly. First, the formation of freshwater cap may inhibit the migration of mesozooplankton (e.g. copepods) into the water with lower salinity (Grindley, 1964) and change the mesozooplankton composition in the water column (Zhou et al., 2015b), which can release the mesozooplankton grazing pressure on ciliates, then through trophic cascades increase the ciliate grazing on nanoflagellates (HNF) (Chen et al., 2012), reducing the abundance of HNF the main grazer on pico-phytoplankton (Safi and Hall, 1999), and releasing the grazing pressure on pico-phytoplankton (Klauschies et al., 2012). Second, as discussed above, the impeding effect of freshwater cap on phytoplankton accesses to nutrients could lead to poor food quality of phytoplankton as prey, and thus reduce the grazing activity of microzooplankton. Both the arguments suggest that the SSS decrease could result in low microzooplankton grazing rate on pico-phytoplankton such as that observed in the winter cruise.
4.4 Monsoon related vertical mixing and biological activity affecting the contrasting seasonal shifts of nutrients

Why the seasonal shift of nitrate plus nitrite was different from those of phosphate and silicate is open to discuss. The seasonal varied pattern and concentrations of phosphate and silicate, and the summer concentration of nitrate plus nitrite, in surface seawater of the SCS, were consistent with the results reported by Ning et al. (2004). They explained that the low phosphate concentration in surface seawater probably resulted from phytoplankton consumption and less supplementation from deep water because of the strong thermocline in summer. Relatively high concentrations of phosphate and silicate (Tables 2 and 5) could be supplied from deep water, as a result of vertical mixing caused by the strong northeast monsoon. However, the nitrate plus nitrite concentration in the present study was more than one order of magnitude lower in the winter than summer. The nearly undetectable nitrate plus nitrite concentration in the winter is different from the results reported by Ning et al. (2004), but similar to the results reported by Chen et al. (1997). Chen et al. (1997) documented that the concentration of nitrate plus nitrite was undetectable in adjacent waters of the Nansha Islands in winter, and proposed that nitrogen was the limiting factor for the primary production.

We propose that nutrients in surface waters were influenced by both vertical mixing and phytoplankton consumption and release. In the summer, the low even depleted concentrations of phosphate and silicate could be resulted from phytoplankton consumption and less supplementation from deep water because of the strong thermocline formed in summer. In addition, the probably high nitrogen fixation in the SCS (Wong et al., 2002) may not only provide a large input of nitrogen to the surface water, supporting the growth of nitrogen fixers and other phytoplankton (Foster et al., 2011; Groszkopf et al., 2012; Mulholland and Bronk, 2004) and preventing a depletion of the nitrate plus nitrite in the surface water, but may also lead to a more severe phosphorus-depleted condition, as nitrogen fixation consumes a large amount of phosphorus (Dyhrman et al., 2006; Zehr and Pae rl, 2008). In the winter, the strong northeast...
monsoon could induce vertical mixing and large rainfall, both of them may cause community decay of nitrogen fixers. The harmful effects of low salinity on *Trichodesmium* growth and nitrogen fixation, and the influence of rainfall on the occurrence of *Trichodesmium* blooms have been reported (Fu and Bell, 2003; Lugomela et al., 2002). As discussed in Sect. 4.3, after the passage of the strong northeast monsoon, nitrogen in the form of nitrate plus nitrite could be firstly depleted by phytoplankton consumption, leading to the extremely low concentration of nitrate plus nitrite observed during the winter cruise.

### 4.5 Decoupling between phytoplankton growth and microzooplankton grazing influenced by the winter monsoon

The different correlation between *μ* and *m* suggested that coupling between phytoplankton and microzooplankton also varied between the two seasons. Close coupling between phytoplankton and microzooplankton grazing indicated by the positive correlation between *μ* and *m* were broadly reported in previous studies (discussion in Zhou et al., 2011). Our results also showed that the microzooplankton grazing was closely coupled with the phytoplankton growth in the summer cruise, but was not in the winter cruise. We consider that the influence of the northeast monsoon could break the coupling between phytoplankton growth and microzooplankton grazing in winter. Firstly, the comings of strong northeast monsoon supply nutrients from deep water to the surface by enhancing vertical mixing. This episodic input of nutrients could break the coupling between phytoplankton and microzooplankton by stimulating *μ* overwhelming corresponding *m* (Irigoin et al., 2005). Secondly, large rainfall and the resulted SSS decrease may decouple the phytoplankton (especially the pico-phytoplankton) growth and microzooplankton grazing through directly or indirectly influencing the phytoplankton growth and microzooplankton grazing as discussed in Sect. 4.3.
4.6 Implications of the low microzooplankton grazing impact on phytoplankton

As discussed in Sect. 4.1.3, our data of the $\mu$ and $m$ were similar to most of the previous results observed in low latitude tropical waters, but the $m/\mu$ in the SSCS was relatively lower (< 50 % on average) than those reported in the previous studies except that a similar value (49 %) in the Arabian Sea during the northeast monsoon season has been reported by Caron and Denett (1999). Our $m/\mu$ is also lower than the mean values in most of the global sea regions (Schmoker et al., 2013). The low $m/\mu$, i.e. the high growth differential over grazing indicates low remineralization of organic matter mediated by microzooplankton and the increased importance of the phytoplankton-mesozooplankton grazing pathway (Landry et al., 1998). As a result, potentially high vertical biogenic particle fluxes via the formation of phytoplankton aggregates and consumption of those aggregates by mesozooplankton could occur (Legendre and Rassoulzadegan, 1996; Schmoker et al., 2013). Higher vertical biogenic particle fluxes in the prevailing periods of the monsoons than between the monsoons in the SSCS was indeed observed (Wan et al., 2010). Our results suggest that the high growth differential over microzooplankton grazing may account for part of the high vertical biogenic particle fluxes in the SSCS.

4.7 Size-selective grazing contributes to the pico-phytoplankton dominance in the oligotrophic waters of the SSCS

Size selectivity of microzooplankton grazing have been proposed in previous studies (e.g., Burkill et al., 1987; Froneman and Perissinotto, 1996; Huang et al., 2011; Kuipers and Witte, 1999; Sun et al., 2004), and varied patterns of the size selectivity have been reported. For example, higher grazing rate on smaller phytoplankton has been reported accounting for the larger-sized phytoplankton dominance in eutrophic waters (e.g. Strom et al., 2007), but no such pattern were found in other studies (Lie and Wong, 2010; Safi et al., 2007; Strom and Fredrickson, 2008; Zhou et al., 2011). In contrast, Zhou et al. (2015a) found that microzooplankton selectively grazed on nano-
phytoplankton (3–20 µm) in the oligotrophic waters of the SCS in summer, and proposed that the size-selective grazing on nano-phytoplankton contributes to the pico-phytoplankton dominance there. Higher microzooplankton grazing rate on the large-sized phytoplankton (> 5 µm) than the smaller one (< 5 µm) has also been recently reported in the oligotrophic subtropical Northeast Atlantic (Cáceres et al., 2013).

The higher RPI (Fig. 3) for and $m$ (Tables 1 and 2) on the larger-sized phytoplankton than pico-phytoplankton suggests that microzooplankton selectively grazed on the larger-sized phytoplankton in the SSSCS both in the summer and winter. The size-selective grazing balanced part of the relative higher growth rate of the larger-sized phytoplankton (> 3 µm), and led to the nearly equal grazing impact ($m/\mu$) on both the larger-sized phytoplankton and pico-phytoplankton in the winter (Table 3). As a result, microzooplankton grazing maintained the pico-phytoplankton dominance in the study waters during the winter. Therefore, on the basis of the results observed in the present study and those reported by Zhou et al. (2015a) and Cáceres et al. (2013), we propose that microzooplankton grazing contributes to the pico-phytoplankton dominance in oligotrophic subtropical and tropical waters such as that of the SCS.

5 Conclusions

Significant seasonal variations in $\mu$ and $m$ as well as environmental variables under the influence of the East Asian monsoon were observed in the SSSCS. Nutrient supply to the surface influenced by vertical mixing and SSS decrease related to large rainfall were considered as the main factors accounting for the significant low $\mu$ and $m$, and the decoupling between the $\mu$ and $m$ in the SSSCS in the winter.

The $m/\mu$ did not significantly vary between the two seasons. The low $m/\mu$ (< 50 % on average), i.e. the high growth differential over microzooplankton grazing indicates low remineralization of organic matter mediated by microzooplankton and the increased importance of the phytoplankton-mesozooplankton grazing pathway, and may
account for part of the high vertical biogenic particle fluxes in the prevailing periods of the monsoons in the SCS.

Significant size-selective grazing on the larger-sized (> 3 µm) phytoplankton was observed in the SCS both in the summer and winter, which indicates that microzooplankton grazing contributes to the pico-phytoplankton dominance in the oligotrophic tropical waters of the SCS.

**Author contributions.** Linbin Zhou designed the experiments and carried them out. Yehui Tan and Liangmin Huang supervised the projects. Zifeng Hu provided the monthly rainfall data and the Fig. 5. Zhixin Ke provided part of the nutrient data. Linbin Zhou prepared the manuscript with contributions from all authors.

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**References**


Seasonal and size-dependent variations in the phytoplankton growth

L. Zhou et al.


Table 1. Summary of environmental variables, phytoplankton growth rates with ($\mu_n, \text{d}^{-1}$) and ($\mu, \text{d}^{-1}$) without nutrient amendment, and microzooplankton grazing rates ($m, \text{d}^{-1}$) in the southern South China Sea in May–June 2009. SST, surface seawater temperature ($^\circ$C); SSS, surface seawater salinity; 25-T (S) seawater temperature (salinity) at 25 m in depth; Si, silicate (µmolL$^{-1}$); P, phosphate (µmolL$^{-1}$); N, nitrate plus nitrite (µmolL$^{-1}$); Chl a, chlorophyll a (µgL$^{-1}$); $n$, the number of data points in the linear part; $R^2$, the determination coefficient of the regression of the linear part; the superscript “a” denotes a saturated feeding response; the superscript “b” denotes a saturated-increased feeding response. The significant level ($\rho$) of regressions when saturated or saturated-increased feeding responses occurred was in some occasions > 0.05 owing to the few points included in the regression.

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Table 2. Summary of environmental variables, phytoplankton growth rates with ($\mu_n, \text{ d}^{-1}$) and without ($\mu, \text{ d}^{-1}$) nutrient amendment, and microzooplankton grazing rates ($m, \text{ d}^{-1}$) in the southern South China Sea in November 2010. SST, surface seawater temperature (°C); SSS, surface seawater salinity; 25-T (S) seawater temperature (salinity) at 25 m in depth; Si, silicate (µmol L$^{-1}$); P, phosphate (µmol L$^{-1}$); N, nitrate plus nitrite (µmol L$^{-1}$); Chl $a$, chlorophyll $a$ (µg L$^{-1}$); $n$, the number of data points in the linear part; $R^2$, the determination coefficient of the regression of the linear part; the superscript “a” denotes a saturated feeding response; the superscript “b” denotes a saturated-increased feeding response. The significant level ($p$) of regressions when saturated-increased feeding responses occurred was in some occasions $> 0.05$ owing to the few points included in the regression.

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<td>0.013</td>
<td>0.084</td>
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</tr>
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<td>11 Nov</td>
<td>1996</td>
<td>29.22</td>
<td>31.70</td>
<td>29.41</td>
<td>33.11</td>
<td>20.2</td>
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<td>0.04</td>
<td>0.015</td>
<td>0.091</td>
<td>0.77</td>
<td>0.26</td>
<td>0.28</td>
<td>0.89</td>
<td>4</td>
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</tr>
<tr>
<td>KJ42</td>
<td>12 Nov</td>
<td>1460</td>
<td>29.62</td>
<td>31.46</td>
<td>29.32</td>
<td>32.66</td>
<td>7.3</td>
<td>0.36</td>
<td>0.02</td>
<td>0.017</td>
<td>0.088</td>
<td>0.39</td>
<td>0.13</td>
<td>0.50</td>
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<td>4</td>
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</tr>
<tr>
<td>KJ47</td>
<td>13 Nov</td>
<td>511</td>
<td>29.45</td>
<td>32.31</td>
<td>29.33</td>
<td>32.36</td>
<td>9.7</td>
<td>0.28</td>
<td>0.07</td>
<td>0.037</td>
<td>0.121</td>
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<td>0.46</td>
<td>0.60</td>
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</tr>
<tr>
<td>KJ50</td>
<td>14 Nov</td>
<td>1259</td>
<td>29.22</td>
<td>31.69</td>
<td>29.18</td>
<td>32.98</td>
<td>6.0</td>
<td>0.36</td>
<td>0.02</td>
<td>0.020</td>
<td>0.069</td>
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<td>0.13</td>
<td>0.43</td>
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</tr>
<tr>
<td>KJ53</td>
<td>14 Nov</td>
<td>145</td>
<td>29.62</td>
<td>32.06</td>
<td>28.92</td>
<td>33.20</td>
<td>3.8</td>
<td>0.36</td>
<td>0.07</td>
<td>0.017</td>
<td>0.050</td>
<td>-2.03</td>
<td>1.01</td>
<td>-2.06</td>
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</tr>
<tr>
<td>KJ65</td>
<td>16 Nov</td>
<td>2100</td>
<td>29.26</td>
<td>31.83</td>
<td>29.27</td>
<td>32.93</td>
<td>6.6</td>
<td>0.36</td>
<td>0.02</td>
<td>0.014</td>
<td>0.077</td>
<td>0.97</td>
<td>0.33</td>
<td>0.28</td>
<td>0.52</td>
<td>6</td>
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</tr>
<tr>
<td>KJ73</td>
<td>18 Nov</td>
<td>1672</td>
<td>29.42</td>
<td>31.72</td>
<td>29.50</td>
<td>32.84</td>
<td>4.4</td>
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<td>0.07</td>
<td>0.017</td>
<td>0.077</td>
<td>0.87</td>
<td>0.34</td>
<td>0.57</td>
<td>0.94</td>
<td>4</td>
<td>0.030</td>
</tr>
</tbody>
</table>
Table 3. Comparison of phytoplankton growth and mortality rates between two size fractions in the southern South China Sea in Nov 2010. $\mu_n$, phytoplankton growth rates with nutrient amendment (d$^{-1}$); $\mu$, phytoplankton growth rates without nutrient amendment (d$^{-1}$); $m$, microzooplankton grazing rates or phytoplankton mortality rates caused by microzooplankton grazing (d$^{-1}$); the superscript “a” denotes a saturated feeding response; the superscript “b” denotes a saturated-increased feeding response. The significant level ($p$) of regressions when saturated or saturated-increased feeding responses occurred was in some occasions > 0.05 owing to the few points included in the regression.

<table>
<thead>
<tr>
<th>Size fraction</th>
<th>Station</th>
<th>$\mu_n$</th>
<th>$m$</th>
<th>$\mu$</th>
<th>$R^2$</th>
<th>n</th>
<th>$p$</th>
<th>$\mu/\mu_n$</th>
<th>$m/\mu$</th>
</tr>
</thead>
<tbody>
<tr>
<td>&lt; 3 µm</td>
<td>KJ28</td>
<td>0.57</td>
<td>0.15</td>
<td>0.39</td>
<td>0.57</td>
<td>8</td>
<td>0.050</td>
<td>0.69</td>
<td>0.37</td>
</tr>
<tr>
<td></td>
<td>KJ32</td>
<td>1.29</td>
<td>0.16</td>
<td>0.78</td>
<td>0.23</td>
<td>8</td>
<td>0.231</td>
<td>0.60</td>
<td>0.21</td>
</tr>
<tr>
<td></td>
<td>KJ35</td>
<td>1.30</td>
<td>0.50</td>
<td>1.01</td>
<td>0.65</td>
<td>6</td>
<td>0.052</td>
<td>0.78</td>
<td>0.50</td>
</tr>
<tr>
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<td>KJ39</td>
<td>0.50</td>
<td>0.12</td>
<td>0.05</td>
<td>0.89</td>
<td>4$^a$</td>
<td>0.058</td>
<td>0.10</td>
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</tr>
<tr>
<td></td>
<td>KJ42</td>
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<td>0.05</td>
<td>0.33</td>
<td>0.04</td>
<td>4$^b$</td>
<td>0.791</td>
<td>1.24</td>
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</tr>
<tr>
<td></td>
<td>KJ47</td>
<td>1.27</td>
<td>0.10</td>
<td>0.18</td>
<td>0.00</td>
<td>4$^a$</td>
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<tr>
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<td>KJ50</td>
<td>0.73</td>
<td>0.01</td>
<td>0.32</td>
<td>0.96</td>
<td>4$^b$</td>
<td>0.020</td>
<td>0.43</td>
<td>0.03</td>
</tr>
<tr>
<td></td>
<td>KJ53</td>
<td>-2.64</td>
<td>0.89</td>
<td>-2.55</td>
<td>0.59</td>
<td>8</td>
<td>0.027</td>
<td>0.96</td>
<td>-0.35</td>
</tr>
<tr>
<td></td>
<td>KJ56</td>
<td>0.85</td>
<td>0.29</td>
<td>0.15</td>
<td>0.42</td>
<td>6</td>
<td>0.167</td>
<td>0.18</td>
<td>1.88</td>
</tr>
<tr>
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<td>KJ57</td>
<td>0.80</td>
<td>0.39</td>
<td>0.58</td>
<td>0.94</td>
<td>6$^a$</td>
<td>0.001</td>
<td>0.73</td>
<td>0.67</td>
</tr>
<tr>
<td>&gt; 3 µm</td>
<td>KJ28</td>
<td>1.20</td>
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<td>0.77</td>
<td>0.68</td>
<td>8</td>
<td>0.012</td>
<td>0.64</td>
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</tr>
<tr>
<td></td>
<td>KJ32</td>
<td>1.34</td>
<td>0.17</td>
<td>0.62</td>
<td>0.65</td>
<td>8</td>
<td>0.015</td>
<td>0.46</td>
<td>0.28</td>
</tr>
<tr>
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<td>KJ35</td>
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<td>0.09</td>
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</tr>
<tr>
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<td>KJ39</td>
<td>1.71</td>
<td>1.31</td>
<td>1.76</td>
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<td>4$^b$</td>
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<td>1.03</td>
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<tr>
<td></td>
<td>KJ42</td>
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<td>0.38</td>
<td>1.14</td>
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<td>4$^b$</td>
<td>0.520</td>
<td>1.28</td>
<td>0.34</td>
</tr>
<tr>
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<td>KJ47</td>
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<td>0.45</td>
<td>0.69</td>
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<td>8</td>
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<td>0.30</td>
<td>0.65</td>
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<tr>
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<td>KJ50</td>
<td>1.34</td>
<td>0.68</td>
<td>0.97</td>
<td>0.91</td>
<td>4$^a$</td>
<td>0.046</td>
<td>0.72</td>
<td>0.70</td>
</tr>
<tr>
<td></td>
<td>KJ53</td>
<td>-1.17</td>
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<td>-1.39</td>
<td>0.65</td>
<td>8</td>
<td>0.016</td>
<td>1.18</td>
<td>-0.79</td>
</tr>
<tr>
<td></td>
<td>KJ56</td>
<td>1.23</td>
<td>0.22</td>
<td>0.56</td>
<td>0.16</td>
<td>8</td>
<td>0.328</td>
<td>0.46</td>
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</tr>
<tr>
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<td>KJ73</td>
<td>1.03</td>
<td>0.35</td>
<td>0.77</td>
<td>0.92</td>
<td>4$^b$</td>
<td>0.041</td>
<td>0.75</td>
<td>0.46</td>
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</table>
Table 4. Pearson correlation analyses between dilution experimental parameters and environmental variables. $\mu$, phytoplankton growth rates without nutrient amendment; $\mu_n$, phytoplankton growth rates with nutrient amendment; $m$, microzooplankton grazing rates; SST, surface seawater temperature; SSS, surface seawater salinity; Si, silicate; P, phosphate; N, nitrate plus nitrite; Chl $a$, chlorophyll $a$.

<table>
<thead>
<tr>
<th></th>
<th>$\mu$</th>
<th>$m$</th>
<th>SST</th>
<th>SSS</th>
<th>Si</th>
<th>P</th>
<th>N</th>
<th>Chl $a$</th>
<th>$m/\mu$</th>
<th>$\mu/\mu_n$</th>
<th>$\mu - m$</th>
</tr>
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<tbody>
<tr>
<td>$\mu$</td>
<td>1</td>
<td>0.762$^a$</td>
<td>0.425$^b$</td>
<td>0.547$^a$</td>
<td>$-0.348$</td>
<td>$-0.570^a$</td>
<td>0.369</td>
<td>$-0.646^a$</td>
<td>$-0.356$</td>
<td>0.496$^b$</td>
<td>0.827$^a$</td>
</tr>
<tr>
<td>$m$</td>
<td>1</td>
<td>0.617$^a$</td>
<td>0.477$^b$</td>
<td>$-0.206$</td>
<td>$-0.536^a$</td>
<td>0.539$^a$</td>
<td>$-0.660^a$</td>
<td>0.252</td>
<td>0.163</td>
<td>0.266</td>
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<tr>
<td>SST</td>
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<td>0.225</td>
<td>$-0.289$</td>
<td>$-0.426^b$</td>
<td>0.450$^b$</td>
<td>$-0.542^a$</td>
<td>0.093</td>
<td>0.332</td>
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</tr>
<tr>
<td>SSS</td>
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<td>$-0.893^a$</td>
<td>0.649$^a$</td>
<td>$-0.519^b$</td>
<td>$-0.061$</td>
<td>0.293</td>
<td>0.396</td>
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</tr>
<tr>
<td>Si</td>
<td>1</td>
<td>0.484$^b$</td>
<td>$-0.433^b$</td>
<td>0.304</td>
<td>0.239</td>
<td>$-0.406$</td>
<td>$-0.336$</td>
<td></td>
<td></td>
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<tr>
<td>P</td>
<td>1</td>
<td>$-0.721^a$</td>
<td>0.690$^a$</td>
<td>0.002</td>
<td>$-0.357$</td>
<td>$-0.383$</td>
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<tr>
<td>N</td>
<td>1</td>
<td>$-0.541^a$</td>
<td>0.246</td>
<td>0.221</td>
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</tr>
<tr>
<td>Chla</td>
<td>1</td>
<td>$-0.007$</td>
<td>$-0.404$</td>
<td>$-0.389$</td>
<td></td>
<td></td>
<td></td>
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<td></td>
</tr>
<tr>
<td>$m/\mu$</td>
<td>1</td>
<td>$-0.597^a$</td>
<td>$-0.748^a$</td>
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<td></td>
<td></td>
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<tr>
<td>$\mu/\mu_n$</td>
<td>1</td>
<td>0.596$^a$</td>
<td>1</td>
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<td></td>
<td></td>
<td></td>
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</tr>
</tbody>
</table>

$^a$ Significant correlation at the level of 0.01.

$^b$ Significant correlation at the level of 0.05.
### Table 5. Comparisons of environmental variables, and phytoplankton growth and microzooplankton grazing between May–June 2009 and November 2010 in the southern South China Sea.

<table>
<thead>
<tr>
<th>Season</th>
<th>SST (°C)</th>
<th>SSS (‰)</th>
<th>Si (µmol L⁻¹)</th>
<th>P (µmol L⁻¹)</th>
<th>N (µmol L⁻¹)</th>
<th>Chl a (µg L⁻¹)</th>
<th>µ/µₙ</th>
<th>m/µ</th>
<th>µ/µₙ</th>
</tr>
</thead>
<tbody>
<tr>
<td>Summer</td>
<td>29.64 ± 0.60</td>
<td>32.82 ± 0.16</td>
<td>3.8 ± 2.6</td>
<td>0.04 ± 0.03</td>
<td>0.90 ± 0.52</td>
<td>0.066 ± 0.022</td>
<td>0.80 ± 0.25</td>
<td>0.46 ± 0.20</td>
<td>0.92 ± 0.32</td>
</tr>
<tr>
<td>Winter</td>
<td>29.27 ± 0.25</td>
<td>31.92 ± 0.30</td>
<td>7.9 ± 4.7</td>
<td>0.35 ± 0.03</td>
<td>0.04 ± 0.02</td>
<td>0.104 ± 0.024</td>
<td>0.54 ± 0.30</td>
<td>0.27 ± 0.13</td>
<td>0.54 ± 0.22</td>
</tr>
</tbody>
</table>

The mean ratio values of µ/µₙ and m/µ were compared based on the Mann–Whitney test, while the other mean values were compared based on the independent t test.

| p     | < 0.01 | < 0.01 | < 0.01 | < 0.01 | < 0.01 | < 0.01 | < 0.01 | < 0.01 | > 0.1 |

SST, surface seawater temperature (°C); SSS, surface seawater salinity; Si, silicate (µmol L⁻¹); P, phosphate (µmol L⁻¹); N, nitrate plus nitrite (µmol L⁻¹); Chl a, chlorophyll a (µg L⁻¹); µ, phytoplankton growth rates without nutrient amendment (d⁻¹); µₙ, phytoplankton growth rates with nutrient amendment (d⁻¹); m, microzooplankton grazing rates (d⁻¹); p, the significant level.
Figure 1. Stations for dilution experiments in the southern South China Sea during May to June 2009 and November 2010. The open circle indicates experimental stations during May to June 2009; the cross indicates experimental stations during November 2010.
Figure 2. Examples of dilution experiment plots of net phytoplankton growth rate as a function of dilution factor. (a) Linear feeding responses in the dilution experiment at KJ69; (b) saturated feeding responses in the dilution experiment at KJ28; (c) saturated-increasing feeding responses in the dilution experiment at KJ65; (d–f) negative phytoplankton growth rates at KJ53, (d) for total phytoplankton, (e) for the size fraction > 3 µm and (f) for the size fraction < 3 µm; the black circle indicates experimental bottles enriched with addition nutrients; the open circle indicates experimental bottles without nutrient amendment.
Figure 3. Relative preference index (RPI) for size-fractionated phytoplankton. > 3 µm, size fraction larger than 3 µm; < 3 µm, size fraction smaller than 3 µm; the asterisk indicates experiments conducted in May 2009, others were conducted in November 2010. RPI > 1 indicates positive selection and vice versa.
Figure 4. Correlation between phytoplankton growth and microzooplankton grazing rates. (a) May to June 2009; (b) November 2010; > 3 μm, size fraction larger than 3 μm; < 3 μm, size fraction smaller than 3 μm.
Figure 5. Monthly rainfall in the southern South China Sea estimated on the basis of data from the Tropical Rainfall Measuring Mission. (a) May 2009; (b) June 2009; (c) October 2010; (d) November 2010.