Distribution of branched glycerol dialkyl glycerol tetraethers in surface soils of Qinghai–Tibetan Plateau: implications of brGDGTs-based proxies in cold and dry regions

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Abstract. The methylation index of branched tetraethers (MBT) and cyclization ratio of branched tetraethers (CBT) based on the distribution of branched glycerol dialkyl glycerol tetraethers (brGDGTs) are useful proxies for the reconstruction of mean annual air temperature (MAT) and soil pH. Recently, a series of 6-methyl brGDGTs were identified which were previously co-eluted with 5-methyl brGDGTs. However, little is known about the distribution of 6-methyl brGDGTs in Qinghai-Tibet Plateau (QTP), a critical region of the global climate system. Here, we investigated 30 surface soils covering a large area of QTP, among which 6-methyl brGDGTs were the most abundant components (average 53±17% of total brGDGTs). The fractional abundance of 6-methyl brGDGTs showed a good correlation with soil pH, while the global MBT'sME calibration overestimates MAT in cold regions like QTP. We therefore propose a MBT5/6 index including both 5- and 6-methyl brGDGTs, presenting a strong correlation with MAT in QTP: MAT = −20.14 + 39.51 × MBT5/6 \( (n = 27, r^2 = 0.82; \text{RMSE} = 1.3 \, ^\circ\text{C}) \). Another index, namely IBT based on carbon skeleton isomerism of 5-methyl to 6-methyl brGDGTs, is dependent on soil pH: pH = 6.77 − 1.56 × IBT \( (n = 27; \ r^2 = \)
0.74, RMSE = 0.32). Our study suggests that changing the position of methyl group of brGDGTs may be another mechanism for some soil bacteria to adapt ambient pH change besides well-known cyclization.

1. Introduction

The Qinghai–Tibetan Plateau (QTP), with an area of over 2.5 million km² and an average elevation of over 4000 meters above sea level (a.s.l.), is the world highest and largest mountain plateau. The uplift of the QTP since early Cenozoic profoundly influences regional and global climates such as the evolution of Asian monsoon which affects lives of over two billion people (An et al., 2001; Li, 1991; Lin et al., 2008; Wang et al., 2008). A number of studies have showed that the QTP is a highly sensitive area for global climate change (e.g., Kang et al., 2010; Liu & Chen, 2000; Qiu, 2008; Yao et al., 2007). The record of 97 meteorological stations located over 2000 meters a.s.l. in China reveals that winter temperature rise is 0.32 °C per decade in the QTP since 1950s, approximately three times the global warming rate (Liu & Chen, 2000). However, the history of instrumental measurement is too short to fully record the evolution of the QTP climate. The reconstruction of the QTP temperature beyond instrumental measurement is challenging because few quantitative proxies are available. Microfossil assemblages based on pollen, diatom or chironomid are commonly used paleothermometers, but they are also influenced by precipitation, salinity, nutrient or other environmental factors (e.g., Keatley et al., 2009; Meriläinen et al., 2000; Seppä & Birks, 2001). The δ¹⁸O value of ice core in the QTP shows a good correlation with northern hemisphere temperature (Thompson et al., 1997; Yao et al., 2002). Unfortunately, ice core with a long term, continuous record is lacking in most QTP.

Over the past decades, two molecular proxies have been developed for estimation of continental temperature. The first one, namely UK’37, is based on the distribution of haptophyte-derived long-chain alkenones. This proxy was originally proposed for paleoceanography (Brassell et al., 1986; Prahl et al., 1988), but was found applicable for reconstruction of lake surface temperature (e.g., Liu et al., 2006; Zink et al., 2001). A
major limitation of UK’37 is that long-chain alkenones are not always present in lakes, although they were reported in some QTP lakes (e.g., Chu et al., 2005; Liu et al., 2011; Liu et al., 2006). In addition, salinity influences the compositions of long-chain alkenones in lakes (Liu et al., 2011). Besides UK’37, the methylation index of branched tetraethers (MBT) and cyclization ratio of branched tetraethers (CBT) can be also used to infer past continental temperature based on the distribution of branched glycerol dialkyl glycerol tetraethers (brGDGTs) (Weijers et al., 2007b):

\[
MBT = \frac{I_a + I_b + I_c}{I_{a} + I_b + I_c + I_{IIa} + I_{IIb} + I_{IIc} + I_{IIIa} + I_{IIIb} + I_{IIIc} + I_{IIa'} + I_{IIb'} + I_{IIc'} + I_{IIIa'} + I_{IIIb'} + I_{IIIc'}}
\]

(1)

\[
CBT = -\log \frac{I_{IIa} + I_{IIa'}}{I_a + I_{IIa}}
\]

(2)

where roman numbers denote relative abundance of compounds in Fig. 1. It should be pointed out that the Eq. 1 and 2 are rewritten from original definitions because the peaks previously identified as pure 5-methyl brGDGTs (Weijers et al., 2007b) are actually mixtures of 5-methyl and 6-methyl isomers (De Jonge et al., 2013).

So far, only two species of Acidobacteria were identified to produce brGDGTs (Sinninghe Damsté et al., 2011), but the ubiquitous occurrence of brGDGTs in soils/peats, lakes and marginal seas suggest that other biological sources are likely (Schouten et al., 2013 and references therein). By analyzing globally distributed soils, Weijers et al. (2007b) found that the MBT is controlled by mean annual air temperature (MAT) and to less extent by soil pH, whereas CBT only relates to soil pH. Such relationship was corroborated by the subsequent study of Peterse et al. (2012) who proposed a simplified format of MBT (or MBT') based on seven quantifiable brGDGTs.

Since the advent, the MBT(MBT')-CBT paleotemperature proxy has been increasingly used for lakes (e.g., D'Anjou et al., 2013; Loomis et al., 2012; Sun et al., 2011), paleosol-loess sequences (e.g., Peterse et al., 2011; Zech et al., 2012), peat (Ballantyne et al., 2010) and marginal seas (e.g., Bendle et al., 2010; Weijers et al., 2007a; Zell et al., 2014). However, a relatively large scatter in global MBT/CBT–MAT calibrations (about 5 °C for root mean square error; RMSE) suggests that other factors besides temperature may influence brGDGTs-based indices (Peterse et al., 2012; Weijers et al., 2007b). In arid and semiarid areas such as western United States where
precipitation is the ecological limiting factor, mean annual precipitation (MAP) rather than MAT is the most important factor that affects brGDGT compositions (Dirghangi et al., 2013; Menges et al., 2014). The updated global calibration of MBT’-CBT indices (Peterse et al., 2012) also shows a weak correlation with MAT for those soil samples from arid regions (MAP < 500 mm). Some studies suggest that regional calibrations are needed to improve accuracy of the GDGTs-based proxy (e.g., Loomis et al., 2012; Pearson et al., 2011; Shanahan et al., 2013; Zink et al., 2010).

Another factor to cause the relatively large scatter of the MBT/CBT-MAT calibration is analytical error. By applying advanced analytical techniques, De Jonge et al. (2013) identified a series of novel 6-methyl brGDGTs which previously co-eluted with 5-methyl GDGTs that were used to calculate the brGDGTs proxies. The successful separation of 5- and 6-methyl brGDGTs resulted in a set of new brGDGT proxies, which were used to recalibrate traditionally defined MBT-CBT indexes (De Jonge et al., 2014):

\[
MBT'_{5-ME} = \frac{I_a + I_b + I_c}{I_a + I_b + I_c + II_a + II_b + II_c + III_a}
\]

MAT = -8.57 + 31.45 × MBT'_{5-ME} \quad (3)

\( n = 222, r^2 = 0.66; RMSE = 4.8 \, ^\circ C, P < 0.001 \)

\[
CBT_{5-ME} = -\log \frac{I_b + II_b}{I_a + II_a}
\]

\( n = 221, r^2 = 0.60; RMSE = 0.84, P < 0.001 \)

\[
pH = 7.84 - 1.73 \times CBT_{5-ME}
\]

\( n = 221, r^2 = 0.85; RMSE = 0.52, P < 0.001 \)

For the QTP, several studies have reported GDGTs in lakes, mountains, hot springs and paleo-soils (e.g., Günther et al., 2014; He et al., 2012; Liu et al., 2013; Wang et al., 2012; Wu et al., 2013; Xie et al., 2012). Wang et al. (2012) analyzed GDGTs in surface sediments of the Lake Qinghai and surrounding soils, showing that brGDGTs-inferred MAT and soil pH were consistent with measured values. In contrast, Wu et al. (2013) found that brGDGTs-derived MAT was higher than instrumentally measured MAT in Kusai Lake sediments from the QTP. Based on the distributions of GDGTs in surface
sediments of the QTP lakes, Günther et al. (2014) developed the local calibration of MBT'-CBT ($r^2 = 0.59$; RMSE = 1.2 °C). However, there are only 9 lake sediments included in Günther et al. (2014). For the application of MBT-CBT indices in lakes, brGDGTs in lake sediments must be exclusively derived from inputs of surrounding soils. However, in-situ production of brGDGTs occurs in various lakes (e.g., Blaga et al., 2009; Blaga et al., 2010; Fietz et al., 2012; Pearson et al., 2011; Sinninghe Damsté et al., 2009; Tierney & Russell, 2009). Furthermore, the 6-methyl brGDGTs were not reported in any QTP studies, which may explain the relatively low $r^2$ value of the MBT/CBT-MAT calibration (e.g., Günther et al., 2014). Given these facts, a direct investigation of soils with improved chromatography is needed to understand environmental influences on the brGDGT distributions in the QTP.

Here, we analyzed all 5- and 6-methyl brGDGTs in 30 surface soils from the QTP. Our main objectives are to (1) determine the relative abundance and distribution of 5- and 6-methyl brGDGTs in the QTP soils; (2) evaluate the effect of recently identified 6-methyl brGDGTs on soil pH in the QTP; and (3) test whether global brGDGTs-MAT calibration is applicable in the QTP and thereby understand the influence of temperature, precipitation and soil pH on 5- and 6-methyl brGDGTs in the QTP.

2. Materials and methods

2.1. Sampling

A total of 30 surface soil samples (0-10 cm) were collected during two fieldworks in 2011 and 2012, which cover a large area of the QTP (84.64°~101.20°E; 28.24°~37.45°N) (Fig. 2). Sampling sites are typical alpine meadow, alpine steppe or alpine meadow steppe. The extremely dry winter results in the lack of persistent snow cover in most sampling sites. The soil samples were air-dried and passed through a 2 mm mesh to remove large gravels. Fine roots (if present) were picked up by steel tweezers. The detailed information on the sampling sites and environmental variables are listed in the supplementary material (Table S1).

2.2 Climate data
There are about 70 meteorological stations in the QTP, mainly distributed in the eastern part and northern border of the QTP. Thus, direct observation data on temperature and precipitation at our sampling sites are generally lacking. In this study, we use the WorldClim dataset (Hijmans et al., 2005) to interpolate annual, seasonal and monthly mean precipitation and temperature (Table S1). The local climate is dry and cold. The MAT of our sampling sites ranges from -5.5 to 7.6 °C with a vertical lapse rate of 0.487 °C/100 m to 0.699 °C/100 m (Cheng et al., 2012). The vertical lapse rate of air temperature decreases from north to south of the QTP. The mean annual precipitation (MAP) at different altitudes varies from ca. 85 mm to ca. 495 mm. The integrated maps are derived from monthly temperature and precipitation values gathered from thousands of weather stations around the world from 1950 to 2000 (47,554 locations for precipitation and 24,542 locations for temperature). The original point data was splines interpolated using latitude and longitude at a fine resolution, making it possible to obtain a reasonable estimation of climatic conditions at individual sites. The WorldClim GIS data used contain annual average of 6 climate variables at a 30 arc seconds resolution (~1 km resolution; http://www.worldclim.org/current.htm). Besides MAT and MAP, additional four climate variables were also used to evaluate the relationship between climate and 5- and 6-methyl brGDGTs indices, including Mean Temperature of Wettest Quarter (MWQT), Mean Temperature of Driest Quarter (MDQT), Mean Temperature of Warmest Quarter (MWQT’), Mean Temperature of Coldest Quarter (MCQT). A total of 30 sites from QTP cold and dry regions (Table S1) were extracted by 6 climate variables using Arcgis 9.3.

2.3 Soil pH and brGDGT analyses

For pH measurement, soils were mixed with deionized water in a ratio of 1/2.5 (g/ml). The soil pH values were determined by a pH meter with a precision of ±0.01 pH. The pH was reported as an average of three duplicate measurements for each sample with standard deviation of ±0.05.

The detailed procedure for lipid extraction was described by Wu et al. (2014). About 6 g dry soils were mixed with 600 ng C_{46} GDGT (internal standard) (Huguet et al., 2006)
and ultrasonically extracted with 20 ml dichloromethane (DCM)/methanol (3:1 v:v) for 15 min (3×). The combined extracts were concentrated to near dryness by a rotary evaporator and transferred to small vials. The concentrated extracts were completely dried under a mild stream of N₂ and re-dissolved in DCM. The total extracts were separated into two fractions by 5 ml hexane/DCM (9:1 v:v) and 5 ml DCM/methanol (1:1 v:v), respectively, on silica gel columns. The latter fraction containing brGDGTs was dissolved in 300 μl hexane/EtOAc (84:16,v/v).

The GDGTs were analyzed on an Agilent 1200 High Performance Liquid Chromatography-atmospheric pressure chemical ionization-triple quadruple mass spectrometry (HPLC-APCI-MS²) system (Yang et al., 2015). The injection volume was 10 μl. The separation of 5- and 6-methyl brGDGTs was achieved with two silica columns in sequence (150 mm × 2.1 mm; 1.9 μm, Thermo Finnigan; USA) at a constant flow of 0.2 ml per min. The solvent gradient was: 84% A (hexane) and 16% B (EtOAc) for 5 min, then increasing the amount of B from 16% at 5 min to 18% at 65 min, and then to 100% B in 21 min. The column was flushed with 100% B for 4 min, and then back to 84/16 A/B to equilibrate it for 30 min. The APCI and MS conditions were: vaporizer pressure of $4.2 \times 10^5$ Pa, vaporizer temperature of 400 °C, drying gas flow of 6 L min⁻¹, temperature of 200 °C, capillary voltage of 3500 V, and corona current of 5 μA (3.2 kV). Peak integration was carried out using Agilent MassHunter. Samples were quantified based on comparisons of the respective protonated-ion peak areas of each GDGT to the internal standard in selected ion monitoring (SIM) mode. The protonated ions were m/z 1050, 1048, 1046, 1036, 1034, 1032, 1022, 1020, 1018 and 744 (C₄₆ GDGTs). Since we assume same response factors among different brGDGTs and C₄₆ GDGTs, our study can be only regarded as semi-quantification.

2.4 Statistical analyses

In order to assess the relationship of 5- and 6-methyl brGDGT distributions with environmental variables such as temperature, precipitation and soil pH, we performed redundancy analysis (RDA) (van den Wollenberg, 1977), a constrained form of the linear ordination method of principal components analysis (PCA). Species (fractional
abundance of 15 brGDGTs) were centered and standardized with zero average and unit variance before RDA. The significance of the explanatory variances within a 1% confidence interval was tested with 999 unrestricted Monte Carlo permutations. Subsequently, a series of partial RDAs (pRDA) were performed to constrain the unique and independent influence of individual environmental parameter alone, as well as compared to all other parameters. All statistical analyses were performed with the CANOCO version 4.5 software (Wageningen UR, USA).

3. Results and discussion

3.1 brGDGTs abundance in the QTP soils

All soil samples except for P790, P840 and P855 contain detectable amounts of brGDGTs. Consequently, 27 soils were used to calibrate brGDGTs’ indices in this study. With the application of two silica LC columns in tandem, 5-methyl and 6-methyl brGDGT isomers were successfully separated, increasing the number of detectable brGDGT compounds from 9 (Peterse et al., 2012; Weijers et al., 2006) to 15 (Fig. 1). There were three tetra-methylated brGDGTs (Ia, Ib and Ic), six penta-methylated brGDGTs (IIa, IIb, IIc, IIa’, IIb’, IIc’) and six hexa-methylated brGDGTs (IIIa, IIIb, IIIc, IIIa’, IIIb’, IIIc’). The mean fractional abundance of 5-methyl brGDGTs ($f_{5ME}$) and 6-methyl brGDGTs ($f_{6ME}$) was shown in Fig. 3. The 6-methyl brGDGTs accounted for average 53% of the total amount of brGDGTs, which were dominated by IIa’ and IIIa’.

Such composition of brGDGTs is different from that of the global soils (239 soils) that 5-methyl brGDGT (Ia and IIa) are usually the most abundant isomers and 6-methyl brGDGTs only comprise on average 24% of the total amounts of brGDGTs (De Jonge et al., 2014), suggesting that the brGDGT-producing bacteria may change their membrane lipids to adapt environmental conditions. So far, two species of Acidobacteria are only identified biological sources for brGDGTs, but they only produce tetra-methylated brGDGTs (Sinninghe Damsté et al., 2011). In our study, the majority of the QTP soils are weak alkaline (6.2~8.4 pH unit), which may favor thriving of non-Acidobacteria and thereby lead to the higher proportion of 6-methyl brGDGTs.
3.2 Environmental control on brGDGT distributions in QTP

A number of studies have demonstrated that temperature, precipitation and pH are the most important factors that affect the brGDGT distributions in soils (e.g., De Jonge et al., 2014; Dirghangi et al., 2013; Peterse et al., 2012; Weijers et al., 2006; Weijers et al., 2007b; Yang et al., 2015; Yang et al., 2014). In order to evaluate the contribution of these parameters to 5- and 6-methyl brGDGT distributions in the QTP, a RDA was performed (Fig. 4). The first component explains 65.2% of the variance, mainly reflecting the variation in soil pH and to less extent MAP. Soil pH presents strong positive relationships with fractional abundance of brGDGTs IIIa’, IIb’, IIb, and negative relationships with that of IIIa, IIa, Ia. The second component of the RDA plot explains 6.1% of total variance, mainly reflecting the variation in MAT and MAP. The brGDGT-IIIa’, IIIa, IIa, IIb, IIC show negative relationships with MAT (in the lower part of RDA), whereas brGDGT-IIa, Ia, Ib and Ic present positive relationships with MAT (in the upper part of RDA). These results support a physiological mechanism that soil bacteria change the number of methyl branches of brGDGTs with temperature in order to maintain acceptable fluidity of their membranes (Weijers et al., 2007b).

Our RDA result shows that MAT and pH have a significant independent effect on the brGDGT distribution in the QTP soils, however, no significant correlation was observed between MAP and brGDGTs (p > 0.05; Table 1). Soil pH explaining up to 60.1% of the total variables is the largest contributor to the variance, followed by MAT (up to 16.4%) and MAP (up to 10.8%). The predominant influence of soil pH on brGDGT distributions was also observed in global soil dataset (De Jonge et al., 2014; Peterse et al., 2012) and Chinese soils (Yang et al., 2015; Yang et al., 2014). In order to estimate the independent, marginal effect of MAT, MAP and pH, partial RDA (pRDA) was performed. The explained variance of pH still remains high (39.9%), indicating that brGDGT distributions are indeed linked to soil pH, whereas MAT contribute to a smaller amount (10.6%) of the variance (Table 2). Similar to the result of RDA, pRDA also showed minor contribution of MAP (2.0%) to brGDGT distributions. The comparison between RDA and pRDA suggests a decreasing contribution of these three environmental variables (pH, MAT, MAP) when they are considered as a unique
contribution (Table 2). Thus, there is a “synergistic effect” (an “antagonistic action”) when MAP and pH (MAT and pH) are considered as covariables, resulting in a positive joint effect of 20.4% for total contribution of pH+MAT+MAP to brGDGT distributions in the QTP soils.

3.3 Evaluation of brGDGT-based proxies in the QTP

Since the identification of 6-methyl brGDGTs (De Jonge et al., 2013), a set of new brGDGT indices such as MBT'$_{5\text{ME}}$ and CBT$_{5\text{ME}}$ have been proposed in order to reduce uncertainty of reconstructed MAT and soil pH (De Jonge et al., 2014; Weijers et al., 2007a; Yang et al., 2015). However, even with application of the MBT$_{5\text{ME}}$-MAT recalibration and the multiple regression, relatively large scatter still exists for those samples from cold regions (De Jonge et al., 2014). Therefore, further calibrations of brGDGT-derived proxies are needed.

3.3.1 MAT calibration in cold and dry regions of the QTP

Consistent with the finding of De Jonge et al. (2014), our result shows that CBT$_{5\text{ME}}$ no longer contributes significantly to MAT after the exclusion of 6-methyl brGDGTs ($p = 0.51; n = 27$). Therefore, we use MBT'$_{5\text{ME}}$ only to calibrate MAT. Considering that limited samples from cold regions were included in previous studies (Peterse et al., 2012; Weijers et al., 2007b), we added our QTP data into the global soil dataset (De Jonge et al., 2014), resulting in a new calibration of MBT'$_{5\text{ME}}$-MAT:

$$\text{MAT} = -10.07 + 33.50 \times \text{MBT'}_{5\text{ME}}$$

($n = 249, r^2 = 0.70; \text{RMSE} = 4.7^\circ\text{C}, P < 0.001$) (9)

The correlation coefficient of Eq. 9 ($r^2 = 0.70$) is slightly higher than the previous global calibration ($r^2 = 0.66; \text{Eq. 4}$), while its RMSE (4.7 $^\circ\text{C}$) is similar to the previous calibration (4.8 $^\circ\text{C}; \text{Eq. 4}$) (De Jonge et al., 2014). Furthermore, the comparison of our estimated MAT and actual MAT ($\Delta\text{MAT} = \text{MAT}_{\text{est}} - \text{MAT}_{\text{act}}$) showed an apparent overestimation (average 2.8 $^\circ\text{C}; \text{Fig. 5}$). Therefore, the simple extension of dataset is not successful in improving accuracy of the MBT'$_{5\text{ME}}$-MAT proxy at the global scale.

Alternatively, we conducted a regional calibration of MBT'$_{5\text{ME}}$ versus MAT based
on 27 QTP soils, and a new equation of MBT'_{5ME}-MAT was expressed:

$$\text{MAT} = -10.82 + 28.36 \times \text{MBT'_{5ME}} \quad (n = 27, r^2 = 0.65; \text{RMSE}=1.8 \degree \text{C}, P<0.001) \quad (10)$$

The slope of Eq. 10 (28.36) is distinct difference from that of global surface soils

(33.50; Eq. 4). Meanwhile, its RMSE value (1.8 °C) is substantially smaller than that of

De Jonge et al. (2014) (4.8 °C). This reduced uncertainty in reconstructed MAT is

attributed to smaller spatial heterogeneity of soils, similar vegetation types (e.g., alpine

meadow) and a narrower MAT range (-5.5~7.6°C) in the QTP. Usually, the regional

calibration has higher $r^2$ values than the global one due to its smaller size of dataset and

smaller spatial heterogeneity (Loomis et al., 2014; Yang et al., 2014). However, our

calibration for the QTP has a slightly lower $r^2$ value (0.65) than the global one (0.70; Eq.

9), suggests that the calibration based on MBT'_{5ME} alone is not superior to the traditional

MBT calibration. The RDA result reveals that similar to 5-methyl brGDGTs, 6-methyl

brGDGTs also significantly correlate with MAT (Fig. 4). Thus, we propose a new

brGDGT index (MBT_{5/6}) including 5-methyl brGDGTs used in the traditional definition

and two dominant 6-methyl brGDGTs (IIa' and IIIa’), expressed as:

$$\text{MBT}_{5/6} = \frac{Ia + Ib + Ic + IIa'}{Ia + Ib + Ic + IIa + IIb + IIC + IIIa + IIIa'} \quad (11)$$

Based on data of the QTP soils, the linear correlation of MAT and MBT_{5/6} was

established as (Fig. 6): MAT = $-20.14 + 39.51 \times \text{MBT}_{5/6}$

$$\quad (n = 27, r^2 = 0.82; \text{RMSE} = 1.3 \degree \text{C}, P < 0.001) \quad (12)$$

This calibration has substantially higher $r^2$ (i.e. 0.82) and lower RMSE (i.e. 1.3 °C)

than Eq. 9 (i.e., 0.70 for $r^2$ and 4.7 °C for RMSE) and Eq. 11 (i.e., 0.65 for $r^2$ and 1.8 °C

for RMSE), supporting that the inclusion of 5-methyl and 6-methyl brGDGTs is essential

for improved accuracy of MAT reconstruction. However, this result is different from the

finding from the Mount Shennongjia (central China) that 6-methyl brGDGTs are

regarded as the interference, leading to a larger scatter of the MBT'-MAT proxy (Yang

et al., 2015). Nevertheless, these differences highlight the importance of regional

calibrations of brGDGT proxies.

3.3.2 Effect of soil pH on position of methyl group(s) of brGDGTs
It is generally accepted that the proton permeability of the cell membrane plays a crucial role in maintaining pH gradient across the membrane of bacteria and archaea (Konings et al., 2002). Weijers et al. (2007b) observed a strong correlation between relative abundance of cyclopentane moieties of brGDGTs and soil pH, and hypothesized that, some soil bacteria can change the methyl groups of brGDGTs into cyclopentyl groups with ambient pH rise, which will loosen the packing of the membrane lipids, enabling more water molecules to get trapped.

Following the approach of De Jonge et al. (2014), we got the following correlation between soil pH and CBT’ which is a modified format of originally defined CBT (Weijers et al., 2007b):

\[ \text{pH} = 7.01 + 1.49 \times \text{CBT}' \quad (n = 27, r^2 = 0.78, \text{RMSE} = 0.30 \text{ pH unit}) \]  

The Eq. 12 has slightly lower \( r^2 \) and substantially lower RMSE compared with the global calibration of pH-CBT’ \((n = 221, r^2 = 0.85, \text{RMSE} = 0.52)\) (De Jonge et al., 2014), suggesting that both global and regional calibrations are applicable for soil pH reconstruction.

We noted that some non-cyclopentyl brGDGTs such as Ia, IIa and IIIa show negative correlations with soil pH, while other brGDGTs show positive correlations with soil pH in the RDA (Fig. 4). Based on these facts, we put all positively correlated brGDGTs on the numerator and all negatively correlated brGDGTs on the denominator to build a new CBT index (or CBT’’):

\[ \text{CBT}'' = \log \frac{Ib+Ic+Ie+IIb+IIc+IIe'+IIa'+IIb'+IIe'+IIIa'+IIb'+IIe'}{Ia+IIa+IIa} \]  

A linear correlation between soil pH and CBT’’ was established based on 27 QTP soils:

\[ \text{pH} = 6.93 + 1.49 \times \text{CBT}'' \quad (n = 27, r^2 = 0.80, \text{RMSE} = 0.29 \text{ pH unit}) \]  

The similar \( r^2 \) and RMSE between Eq. 13 and 15 was attributed to minor amounts of brGDGTs Ib, IIb, IIc, IIIb and IIIc (average 8% of total brGDGTs; Fig. 3) which were excluded from the CBT’ index but included in our CBT’’ index.

The fractional abundance of 6-methyl brGDGTs showed strong positive correlations with soil pH in both the QTP \((r^2 = 0.74; \text{Fig. 8})\) and global soil dataset \((0.41 < r^2 < 0.72; \text{De Jonge et al., 2014})\). This is apparent contrast with the previous assumption
that non-cyclopentyl moieties (such as IIa’ and IIIa’) negatively correlate with soil pH. Unlike 6-methyl brGDGTs, some 5-methyl brGDGTs did not show positive correlations with soil pH (de Jonge et al., 2014). Thus, we hypothesize that besides cyclization, the position of methyl group(s) of brGDGTs also influences cell membrane fluidity. In order to test this hypothesis, we define a new index about carbon skeleton Isomerism of Branched Tetraethers (or IBT) as the abundant ratio of non-cyclopentyl 6-methyl to 5-methyl brGDGTs:

$$IBT = -\log \frac{IIa' + IIa}{IIa + IIa'}$$

We performed a linear regression of IBT versus soil pH based on 27 QTP soils (Fig. 8), yielding an equation as:

$$pH = 6.77 - 1.56 \times IBT \quad (n = 27; \ r^2 = 0.74, RMSE = 0.32)$$

(16)

Meanwhile, the linear correlation of CBT_{5ME} and soil pH was also established:

$$pH = 7.98 - 1.12 \times CBT_{5ME} \quad (n = 27; \ r^2 = 0.66, RMSE = 0.37)$$

(17)

For the regional calibration, the IBT index has higher $r^2$ and lower RMSE than traditionally defined CBT_{5ME} index, supporting that the carbon skeleton isomerism of brGDGTs (i.e., changing the position of methyl group) is indeed a physiological mechanism of brGDGTs-producing bacteria to adapt soil pH change.

3.3.3 Seasonality of brGDGTs proxies in the QTP

The QTP is under strong influence of Asian Monsoon, characterized by warm/humid summer (June to August) and dry/cold winter (December to February) (An et al., 2001; Qiu, 2008). In order to examine if there is a seasonal bias on brGDGT distributions, we analyze the correlation coefficients of 5- and 6-methyl brGDGTs proxies (i.e., MBT'_{5ME}, MBT_{5/6}, CBT_{5ME} and IBT) versus annual and seasonal air temperature (Table S2). Overall, there is no apparent seasonal bias for MBT'_{5ME} and MBT_{5/6}. This is likely attributed to significant correlation between seasonal temperature and MAT in the QTP ($r^2 > 0.80, p < 0.0001$). In addition, no significant correlation was observed between the CBT indices/IBT and MAT/seasonal temperature (-0.3 < $r$ < 0.3; Table S2), suggesting minor influence of air temperature on these indices.
Our results are consistent with that of Weijers et al. (2011) who found no significant seasonal bias in MBT-CBT indices in mid-latitude soils. Therefore, the reconstruction of MAT based on the 5- and 6-methyl brGDGTs proxies is doable in the QTP.

4. Conclusions

By applying improved chromatography, we successfully separated 5- and 6-methyl brGDGTs in the surface soils from the Qinghai-Tibet Plateau (QTP), a cold and dry region. This is the first time to report 6-methyl brGDGTs in the QTP, providing an opportunity to optimize brGDGTs’ proxies in this critical region. Three conclusions were reached based on brGDGT data in 27 surface soils. Firstly, the 6-methyl brGDGTs are widely distributed in the QTP soils accounting for average 53% of total amounts of brGDGTs. Secondly, soil pH is the most important contributor to the variance of brGDGTs, followed by MAT, while MAP has no significant effect on brGDGTs’ distributions. Thirdly, two new indices including recently identified 6-methyl brGDGTs were proposed to estimate MAT and soil pH, respectively. The first one, namely MBT$_{5/6}$, is useful for the MAT reconstruction in cold and dry regions (like QTP) with an improved RMSE of 1.3 $^\circ$C. The second one, namely IBT, is allowed to estimate soil pH with an RMSE of 0.32. Our study demonstrates that besides previously reported cyclization, isomerization of 5-methyl to 6-methyl brGDGTs (expressed as IBT) is another strategy for brGDGTs-producing bacteria to adapt ambient pH change.

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**Fig. 1.** Molecular structures of 5- and 6-methyl branched GDGTs used in this study. The compounds that have one or two methyl groups at the ω6 or ω6 position are defined as 6-methyl brGDGTs, while the compounds that have one or two methyl groups at the ω5 or ω5 position are defined as 5-methyl brGDGTs.
Fig. 2. Locations of soil sampling sites (n = 30) in the QTP (Pink solid circles).
Fig. 3. Average ($n = 27$) fractional abundance of brGDGTs in surface soils of the QTP.
Fig. 4. RDA triplot showing the relationship between 5- and 6-methyl brGDGTs%, MAT, MAP and soil pH from the QTP. Numbers in the plot correspond to the soils in supplementary material (Table S1). The first and second axis explained 65.2% and 6.1% of the variance, respectively.
Fig. 5. A) Scatterplot of MBT'\textsubscript{5ME} with actual MAT; B: difference between estimated MAT and actual MAT (ΔMAT). Solid and empty circles represented soils in this study and global soils (de Jonge et al., 2014), respectively.
Fig. 6. A) Linear regression of MBT'\textsubscript{5ME} with actual MAT; B) difference between estimated MAT and actual MAT (\(\Delta\text{MAT}\)). Data are from this study for 27 surface soils of the QTP.
Fig. 7. Linear regression plot of MBT\textsubscript{5/6} versus MAT in the QTP.

\[ r^2 = 0.82 \]
\[ \text{RMSE} = 1.3°C \]
**Fig. 8.** Plots of fractional abundance of 6-methyl brGDGTs of the total amount of brGDGTs ($f_{6ME}$) versus soil pH in the QTP.
Fig. 9. Scatterplots of A) soil pH versus CBT_{SME} and B) soil pH versus IBT based on 27 soil samples in the QTP.
Table 1: Results of RDA and partial RDA (pRDA) showing the total and unique contributions of soil pH, MAT and MAP to the variance in brGDGT distributions in the QTP soils.

<table>
<thead>
<tr>
<th>Variables</th>
<th>Total contribution (%)</th>
<th>Unique contribution (%)</th>
<th>p Value</th>
<th>p Value</th>
<th>Max eigenvalues*</th>
<th>Eigenvalues</th>
</tr>
</thead>
<tbody>
<tr>
<td>pH</td>
<td>0.001</td>
<td>60.1</td>
<td>0.001</td>
<td>39.9</td>
<td>39.9</td>
<td></td>
</tr>
<tr>
<td>MAT</td>
<td>0.001</td>
<td>16.4</td>
<td>0.001</td>
<td>10.6</td>
<td>10.6</td>
<td></td>
</tr>
<tr>
<td>MAP</td>
<td>0.179</td>
<td>10.8</td>
<td>0.172</td>
<td>2.0</td>
<td>2.0</td>
<td></td>
</tr>
<tr>
<td>All variables</td>
<td>0.001</td>
<td>72.9</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Joint effects</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>20.4</td>
<td></td>
</tr>
</tbody>
</table>

*The first environmental variable which has been selected into the analysis has the maximum eigenvalues (explained variances), there are 6 sequences with different arrangement of pH, MAT and MAP. However, no matter which sequence has been selected for RDA, the total variables contribution is invariant.