Foliar photochemical processes and carbon metabolism under favourable and adverse winter conditions in a Mediterranean mixed forest, Catalonia (Spain)

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Abstract:

Evergreen trees in the Mediterranean region must cope with a wide range of environmental stresses from summer drought to winter cold. The mildness of Mediterranean winters can periodically lead to favourable environmental conditions above the threshold for a positive carbon balance, benefitting evergreen woody species more than deciduous ones. The comparatively lower solar energy input in winter decreases the foliar light saturation point. This leads to a higher susceptibility to photoinhibitory stress especially when chilly (<12 °C) or freezing temperatures (<0°C) coincide with clear skies and relatively high solar irradiances. Nonetheless, the advantage of evergreen species that are able to photosynthesize all year round where a significant fraction can be attributed to winter months, compensates for the lower carbon uptake during spring and summer in comparison to deciduous species. We investigated the ecophysiological behaviour of three co-occurring mature evergreen tree species (\textit{Quercus ilex} L., \textit{Pinus halepensis} Mill., and \textit{Arbutus unedo} L.). Therefore, we collected twigs from the field during a period of mild winter conditions and after a sudden
cold period. After both periods, the state of the photosynthetic machinery was tested in the laboratory by estimating the foliar photosynthetic potential with CO₂ response curves in parallel with chlorophyll fluorescence measurements. The studied evergreen tree species benefited strongly from mild winter conditions by exhibiting extraordinarily high photosynthetic potentials. A sudden period of frost, however, negatively affected the photosynthetic apparatus, leading to significant decreases in key physiological parameters such as the maximum carboxylation velocity (V_{c,max}), the maximum photosynthetic electron transport rate (J_{max}), and the optimal fluorometric quantum yield of photosystem II (F_v/F_m). The responses of V_{c,max} and J_{max} were highly species-specific, where Q. ilex exhibited the highest and P. halepensis the lowest reductions. In contrast, the optimal fluorometric quantum yield of photosystem II (F_v/F_m) was significantly lower in A. unedo after the cold period. The leaf position played an important role in Q. ilex showing a stronger winter effect on sunlit leaves in comparison to shaded leaves. Our results generally agreed with the previous classifications of photoinhibition-tolerant (P. halepensis) and photoinhibition-avoiding (Q. ilex) species on the basis of their susceptibility to dynamic photoinhibition, whereas A. unedo was the least tolerant to photoinhibition, which was chronic in this species. Q. ilex and P. halepensis seem to follow contrasting photoprotective strategies. However, they seemed equally successful under the prevailing conditions exhibiting an adaptive advantage over A. unedo. These results show that our understanding of the dynamics of interspecific competition in Mediterranean ecosystems requires consideration of the physiological behaviour during winter which may have important implications for long-term carbon budgets and growth trends.

1 Introduction

Mediterranean-type ecosystems are widely associated with broadleaved evergreen sclerophyllous shrubs and trees, the classic vegetation types in climates where hot and dry summers alternate with cool and wet winters (Aschmann, 1973; Blumler, 1991; Orshan, 1983; Specht, 1969). In summer, water is undoubtedly the most important factor limiting growth and survival in the Mediterranean region, whereas spring and autumn provide better growing conditions (Gracia et al., 1999; Orshan, 1983; Sabaté and Gracia, 2011). In winter, the low temperatures and solar radiation limit the amount of energy available for the vegetation, although soil-water contents and water-pressure deficits are favourable. This highly dynamic seasonality of favourable and unfavourable conditions produces a rich diversity of plants in
these regions (Cowling et al., 1996). In turn, this features a highly diverse range of traits and
taxa that has produced multiple survival strategies which help to explain the abundance and
distribution of species (Matesanz and Valladares, 2013). Nonetheless, the predicted
reductions in annual precipitation, increases in mean temperature, and increases in the
variability and occurrence of extreme droughts and heat waves in arid and semi-arid regions
are likely to affect species abundance and distribution (Friend, 2010; IPCC, 2013; Somot et
al., 2008). The battle for survival and dominance in plant communities facing these novel
changes in their environments evokes great uncertainties and worries in the scientific
community concerning the adaptive ability, distribution shifts, or, at worst local extinction of
species especially in Mediterranean type ecosystems (Matesanz and Valladares, 2013;
Peñuelas et al., 2013).

In this context, a pivotal role devolves on the winter period in Mediterranean type- climates as
mild winter temperatures can suddenly provide potential periods of growth and recovery from
stressful summer drought periods, above all for evergreen trees. Thus, the success in the
future dynamics of competition and novel environmental conditions will not only depend
upon the tolerance to withstand abiotic stresses, but also on their effectiveness to benefit
rapidly from periods when environmental conditions may be favourable such as in winter.
The effective acclimation of the photosynthetic apparatus during winter was hereby in the
focus of interest for this study. This acclimation is particularly essential for evergreen tree
species in order to compensate for their lower photosynthetic rates during the growth period,
relative to deciduous species. Plants have evolved diverse adaptive mechanisms to cope with
the consequences of stress and to acclimate to low temperatures (Blumler, 1991; Öquist and
Huner, 2003).

Hereby, mixed forests provide us with an ideal test-bed for investigating the different
ecophysiological strategies and their sensitivities to abiotic stresses, because all tree species
have to contend equally with the yearly variability of environmental conditions. Nevertheless,
most ecophysiological studies have been conducted in spring and summer, and winter has
been surprisingly overlooked despite its importance for our understanding of the dominance
of certain vegetation-types and of the responses of vegetation to stress, seasonality, and
species composition (Oliveira and Peñuelas, 2004; Orshan, 1983; Tretiach et al., 1997). Even
though efforts have recently been made to elucidate the behaviour of sclerophyllous
ecosystems under variable winter conditions (e.g. García-Plazaola et al., 1999, 1997;
Kyparissis et al., 2000; Levizou et al., 2004; Martínez-Ferri et al., 2004; Oliveira and Peñuelas, 2004, 2000), the physiological behaviour of co-occurring species of evergreen trees in the Mediterranean region, including leaf gas exchange (GE) and chlorophyll fluorescence (CF) methods, have been insufficiently studied for understanding the dynamics of photoinhibitory stress and interspecific competition. Therefore, in our study we used an ample set of parameters from GE & CF measurements in order to provide a snapshot in the plant’s physiology and in order to characterize in detail the effects on the photosynthetic light and carbon reactions during winter (Flexas et al., 2008; Guidi and Calatayud, 2014). This study was conducted on three species of evergreen trees (Quercus ilex L., Pinus halepensis Mill., Arbutus unedo L.) in northern Catalonia near Barcelona, Spain.

Our aims were to i) investigate the foliar physiology of these three species under mild winter conditions, ii) analyse the effect of sudden changes from favourable to unfavourable conditions on photochemical and non-photochemical processes associated with electron transport, CO₂ fixation, and heat dissipation, iii) determine if leaves exhibit distinct locational (sunlit or shaded) responses to winter stress, and iv) identify the species-specific strategies when coping with stress, induced by low temperatures and frost. These topics are of particular interest due to the recent report of an increased dominance of angiosperm trees and the negative impacts on pines over extensive areas of the Iberian Peninsula (Carnicer et al., 2013). Therefore, we must improve our understanding of the interactions among co-occurring tree species competing for scarce resources and trying to survive and tolerate novel environmental conditions to be able to predict ecosystem responses to global climate change.

2 Material and Methods

2.1 Field site

Our experiment was conducted at the field station of Can Balasc in Collserola Natural Park, a coastal massif (8500 ha) in the hinterlands of Barcelona, northeastern Spain (41° 25’ N, 2° 04’ E, 270 m a.s.l.). The forest stand at the study site has an area of 0.7 ha and is on a northeast-facing slope. The climate is characterised by typical Mediterranean seasonal summer droughts and warm temperatures, with a mean August temperature of 22.8 °C. The proximity to the Mediterranean Sea provides mild winters where frosts and snow are rare, as reflected in the mean January temperature of 7.9 °C. Mean annual precipitation and
temperature are 723 mm and 15.1 °C (1951-2010), respectively (Ninyerola et al., 2000). The soils have predominantly developed above lithological strata of shales and granite (Sanchez-Humanes and Espelta, 2011). Sensors for measuring air temperature (HMP45C, Vaisala Oyj, Finland) and solar radiation (SP1110 Skye Instruments Ltd., Powys, UK) were installed at a height of 3 m, in a clearing ca. 1 km from the plot.

2.2 Stand history and composition of tree species

The history of Collserola Natural Park is typical for the area, being characterised by intensive exploitation for charcoal in Quercus- coppice forests and for agricultural purposes such as olive production until the 20th century. The abandonment of these practices at the beginning of the 20th century led to forest succession and restoration with the early successional and fast growing Aleppo Pine (P. halepensis Mill.). As in wide parts of the Mediterranean basin, this tree species was favoured by forest management for its rapid growth rates and timber yields (Maestre and Cortina, 2004). The cessation of forest practices in the early 1950s led to a second wave of succession characterised by extensive regeneration of the evergreen Holm Oak (Q. ilex L.) and the deciduous Pubescent Oak (Q. pubescens Willd.). As a result, many mixed forest stands in Collserola are currently characterised by two-layered canopies consisting of a dense layer from Quercus species surmounted by shelter trees of P. halepensis. The forest stand at our experimental site has reached the next and final stage of forest succession, where the dense Quercus canopy is out-competing the early successional P. halepensis, simply by suppressing the growth of the light demanding pine seedlings and saplings. This final stage of succession is typical of many pine-oak forest-type sites in the Iberia Peninsula. P. halepensis. is dependent mainly on fire disturbances for natural regeneration (Zavala et al., 2000). Interestingly, the diversity of tree species is enriched by the scattered occurrence of Strawberry trees (A. unedo) in the forest canopy being usually more characterised as a shrubby species widely abundant in the macchia ecosystems of the Iberian peninsula (Beyschlag et al., 1986; Reichstein et al., 2002). Its existence adds an ecological value to the forest due to its flowering and fruiting behaviour attracting insects and birds. It raises questions about its performance as a mature tree within the interspecific competition of this mixed forest. The forest diversity also encompasses a dense understory mainly consisting of Pistacia lentiscus L., Erica arborea L., Phillyrea latifolia L., Rhamnus alaternus L., Cistus spp, Crataegus monogyna Jacq., Bupleurum fruticosum L., and other less abundant species. The stand at our study site has reached a highly diverse stage of forest succession and has
provided us with a rare set of some of the most important Mediterranean tree species growing together naturally.

2.3 Sampling

The sampling of the mild winter period took place between 09.01.-19.01.12 (DOY 9-19). The frosty/chilly period lasted from 19.01.-04.02.12 (DOY 21-35). The sampling period after the frosty/chilly period took place between 14.02.-24.02.12 (DOY 45-55). We obtained sunlit leaves for GE-analyses by sampling five twigs with a pruning pull from the outer part of the upper third of the crown, and shaded leaves by sampling five twigs from the inner part of the crown, optimally at similar heights. In the second field campaign after the frost occurrence, however, we were constrained to sample shaded leaves only from Q. ilex due to limitation in labour and equipment. The shaded leaves of P. halepensis and A. unedo could only be sampled in the first, but not in the second field campaign. The twigs were immediately re-cut under water in buckets in the field and transported to the laboratory retained in plastic bags to minimise transpiration. Five replicates of each species were collected for the analysis of GE. The twigs were pre-conditioned in the laboratory at a room temperature of 24-28 °C in dim light for 1-3 d and freshly cut the following morning before the measurement of GE (Niinemets et al., 1999, 2005). We intended to avoid the problems we had faced in the field, such as the limited ability of the instruments to reach the standard operating temperature of 25 °C, which was hampered by low ambient temperatures or unpredictable plant responses such as closed stomata or patchy stomatal conductance (Mott and Buckley, 1998, 2000). The pre-conditioned twigs instead had a stable Ci and sufficiently high gs, which are required for conducting a noise-free CO2-response curve. The method of cutting twigs rehydrated stressed leaves at optimum conditions and allowed us to analyse their long-term acclimation to the environmental conditions from which they were derived. This method has been used in other studies (Epron and Dreyer, 1992; Haldimann and Feller, 2004; Laisk et al., 2002; Niinemets et al., 1999, 2005), and we confirmed that the leaves remained fresh and functional for several days controlled by gs and fluorescent signals (data not shown). Our ambient values of the GE- and CF-derived parameters accordingly represented the “ambient capacity” of pre-conditioned leaves under near-optimal ambient environmental conditions of CO2 concentrations and saturating light and at a room temperature of 20-25 °C (Reich et al., 1998).
2.4 GE and CF analyses

GE and CF were measured with a Li-Cor LI-6400XT Portable Photosynthesis System equipped with a LI-6400-40 Leaf Chamber Fluorometer (Li-Cor, Inc., Lincoln, NE, USA). Response curves for foliar net assimilation versus CO₂ concentration were recorded from five apparently healthy leaves per tree species and leaf position. CF was measured in parallel. *A. unedo* leaves were sufficiently large to cover the leaf cuvette (2 cm²), whereas sunlit leaves of *Q. ilex* were in some cases too small, and the area of the leaves had to be adjusted after the measurements. For the leaves of *P. halepensis*, we positioned a layer of needles (appr. 10-15) on the leaf cuvette, avoiding gaps and overlays. The putty-like adhesive ‘Blu-tack’ (Bostik SA, La Plaine St Denis, France) was also used to seal the gaskets and to keep the needles in position.

2.4.1 Preparation and acclimation

Prior to recording the response curves, the temperature of the clamped leaves (TLeaf) was adjusted to 25 °C, and the flow of ambient CO₂ in the leaf chamber (Cₐ) was set to 400 μmol CO₂ m⁻² s⁻¹ (controlled with a CO₂ mixer). The leaves were dark-adapted for 15-20 min before the measurements, and the data were logged when the GE-derived parameters such as stomatal conductance (gₛ), stomatal internal CO₂ concentration (Cᵢ) and mitochondrial respiration in darkness (Rₘ) had stabilised. For our purposes, dark-adaption did not necessarily mean strict prolonged darkness but referred to a sufficiently low level of ambient background light that did not cause an accumulation of reduced photosystem II (PSII) acceptors, which could be detected as an increase in fluorescence. The leaves were also pre-darkened with special leaf clips or a dark cloth to save time. The chamber light was then turned on at a saturating quantum flux density of 1000 μmol photons m⁻² s⁻¹ (20% blue LED, 80% red LED). The relatively high percentage of blue light stimulated the stomata to open (Farquhar and Sharkey, 1982; Kang et al., 2009; Niinemets et al., 2005). The relative humidity was maintained at 50% (±10%), and the air flow was maintained at 500 μmol s⁻¹. The above conditions were maintained for approximately 20-30 min until the net rate of carbon assimilation (Aₙₑₜ), gₛ, and Cᵢ of the leaf stabilised.

The GE-derived parameters Aₙₑₜ, gₛ, and Cᵢ likely require less time to stabilize, especially in healthy and unstressed leaves, but this minimum time range was necessary for the CF-derived parameters to ensure accurate measurement of the efficiency of harvesting light energy by
maximal oxidation and therefore open PSII reaction centres under ambient conditions of CO$_2$
and saturating light, which can be monitored by observing the stability of steady-state
fluorescence ($F_s$). If this stability is not achieved, the effective quantum yield of PSII ($\Phi_{PSII}$)
and subsequent calculations of important parameters such as the rate of electron transport
based on the CF measurement ($J_{CF}$) could be underestimated. After all parameters had
stabilised, the steady-state GE-derived parameters and several CF-derived parameters in the
light-adapted state were recorded simultaneously. $F_s$ followed shortly afterwards by the
maximum fluorescence yield in the light-adapted state ($F_{m'}$) were logged by the emission of a
pulse of white light at 10000 mmol m$^{-2}$ s$^{-1}$ to close all PSII reaction centres, followed by a so-
called ‘dark pulse’ for measuring the minimal fluorescence ($F_o'$) of a light-adapted leaf that
has been momentarily darkened. The measurement of CO$_2$ began after the completion of the
preparation and acclimation, which required approximately 30 min in unstressed leaves and
up to 2 h in stressed leaves.

2.4.2 CO$_2$ experiments

The CO$_2$-response curves were recorded at a $T_{Leaf}$ of 25 °C and a quantum flux density of
1000 $\mu$mol photons m$^{-2}$s$^{-1}$. The values of $C_a$ used to generate the response curves were 400
→ 300 → 200 → 150 → 100 → 50 → 400 → 400 → 600 → 800 → 1200 → 2000 (in $\mu$mol
CO$_2$ m$^{-2}$ s$^{-1}$). The minimum and maximum times for stabilising $A_{net}$, $g_s$, and $C_i$ for each log
were set to 4 and 6 min, respectively.

2.4.3 Light experiments

Light-response curves (A/PPFD) were generated at a $C_a$ of 400 $\mu$mol CO$_2$ m$^{-2}$ s$^{-1}$ by
automatically applying changes in the photosynthetically active radiation with the LI-6400XT
light source. To obtain precise responses at the low range of the light gradient for estimating
the daily mitochondrial respiration by the Kok effect (Kok, 1948), we used the following
PPFD sequence: 2500 → 2000 → 1500 → 1000 → 800 → 600 → 500 → 400 → 300 → 200
→ 150 → 125 → 100 → 75 → 50 → 40 → 30 → 20 → 10 → 5 → 0 (in $\mu$mol photons m$^{-2}$ s$^{-1}$).
The minimum and maximum times between each light level for the generation of the
A/PPFD curves were set to 1 and 2 min, respectively. The gradient from high to low light
during an A/PPFD curve led to a drop in $T_{Leaf}$ as the light decreased. The rapid changes in the
light levels prevented the adjustment of $T_{Leaf}$ while guaranteeing stable air and water fluxes
and avoiding noisy measurements of $C_i$ and $g_s$. We thus decided to maintain a stable Peltier-
block temperature ($T_{\text{block}}$) in the leaf cuvette. Hence, $T_{\text{block}}$ was first adjusted so that $T_{\text{leaf}}$ was 25 °C at the beginning of the A/PPFD curve and then kept stable throughout the experiment. $T_{\text{Leaf}}$ had dropped by approximately 1-3 °C by the completion of the A/PPFD curve.

The calculation of the parameters $F_v/F_m$, NPQ, qp, and temperature functions, in supplementary material.

2.5 Calculation of CF-derived parameters

The maximum efficiency of PSII was calculated by:

$$\frac{F_v}{F_m} = \frac{(F_m - F_o)}{F_m}$$

where $F_v$ is the variable fluorescence of a dark-adapted sample, $F_m$ is the maximal fluorescence measured after a saturating light pulse, and $F_o$ is the minimal fluorescence measured under darkness. These parameters were obtained from dark-adapted leaves with closed PSII reaction centres as described in the previous sections. The $F_v/F_m$ ratio describes the fraction of photochemically absorbed photons under dark conditions. Typical values range between 0.75 and 0.85, depending on age, health, and preconditioning. The $F_v/F_m$ ratio provides information about the maximum or optimum quantum yield and serves as indicator of stress in the photosystems (Buschmann, 2007). Ratios below 0.80 are indicative of induced photoprotection and sustained energy dissipation (Maxwell and Johnson, 2000; Verhoeven, 2014), whereas leaves with ratios below 0.74 are considered to be below the recovery threshold (Björkman and Demmig, 1987). The effective quantum yield of PSII was estimated by:

$$\Phi_{\text{PSII}} = \frac{(F_m' - F_s)}{F_m'}$$

where $F_s$ is the steady-state fluorescence in a fully light-adapted sample, and $F_m'$ is the maximal fluorescence yield after a pulse of high light. The $\Phi_{\text{PSII}}$ is the counterpart of the optimum quantum yield and represents the fraction of photochemically absorbed photons in a light-adapted leaf (Maxwell and Johnson, 2000).

$$J_{CF} = \varepsilon * \Phi_{\text{PSII}} * \alpha_L$$

where $\varepsilon$ is a scaling factor for the partitioning of intercepted light between photosystems I and II. We assumed that light was equally distributed between both photosystems ($\varepsilon = 0.5$)
(Bernacchi et al., 2002; Niinemets et al., 2005). $\alpha_L$ is the foliar absorbance determined in separate measurements of foliar reflectance and transmittance. The following values of $\alpha_L$ were determined: 0.932 for Q. ilex and 0.912 for P. halepensis, with no differences between sunlit and shaded leaves of these two species, and 0.935 for sunlit leaves of A. unedo, and 0.917 for shaded leaves of A. unedo. For the determination of these leaf absorptances ($\alpha_L$), foliar reflectance and transmittance were measured at midday in August 2012 using a spectroradiometer UniSpec Spectral Analysis System (PP Systems, Haverhill, MA, USA). The value of $J_{CF}$ at a CO$_2$ concentration of 400 $\mu$mol CO$_2$ m$^{-2}$ s$^{-1}$ and a PPFD of 1000 $\mu$mol photons m$^{-2}$ s$^{-1}$ was termed ambient photosynthetic electron transport ($J_{amb}$). Its relationship with the net assimilation rate ($J_{amb}/A_{net}$) was used for the analyses of alternative electron sinks beside carbon metabolism.

### 2.6 Estimation of light respiration and calculation of the effective quantum yield of CO$_2$ ($\Phi_{CO_2}$)

In the literature, the term $R_d$ was sometimes used for dark respiration (Farquhar et al., 1980; Turnbull et al., 2003), but also for day respiration (Flexas et al., 2012; Yin et al., 2011). We will use $R_d$ to represent mitochondrial respiration during the day or under lighted conditions and $R_n$ to represent mitochondrial respiration at night or under dark-adapted conditions. We estimated $R_n$ during the day after darkening the leaf for at least 30 min. $R_d$ was estimated from the light-response curves with the combined GE and CF measurements proposed by Yin et al. (2009), named the CF method. This method amended the Kok method (Kok 1948) by substituting the $A/PPFD$ relationship with $A/PPFD \times \Phi_{PSII}$ (Yin et al., 2009). See reference for details.

The effective quantum yield of CO$_2$ ($\Phi_{CO_2}$, unitless) can be calculated using the estimated $\alpha_L$, $R_d$, together with $A_{net}$ and PPFD as follows

$$\Phi_{CO_2} = \frac{(A_{net} - R_d)}{PPFD \times \alpha_L}$$

### 2.7 The Farquhar, von Caemmerer, and Berry (1980) photosynthesis model (FvCB)

The FvCB photosynthesis model was employed on the assumption that foliar carbon assimilation was limited either by Rubisco activity ($A_c$) or by ribulose-1,5-bisphosphate (RuBP) regeneration ($A_j$) and was driven by light, temperature, and CO$_2$. The model was
further complemented with a third limitation: the photosynthetic rate limited by triose-phosphate use \( (A_p) \) (Sharkey, 1985). \( A_{net} \) can then be determined by the minimum of these three potential rates from an A/Cc curve:

\[
A_{net} = \min\{A_c, A_j, A_p, \}
\]  

where

\[
A_c = V_{c,\text{max}} \left[ \frac{C_c - \Gamma^*}{C_c + K_c (1 + \frac{O}{K_O})} \right] - R_d
\]  

where \( V_{c,\text{max}} \) represents the maximum rate of Rubisco carboxylation, \( K_c \) is the Michaelis-Menten constant of Rubisco for CO\(_2\), \( O \) is the partial pressure of O\(_2\) at Rubisco, and \( K_O \) is the Michaelis-Menten constant of Rubisco for O\(_2\) (Table C1, see Appendix C) and \( C_c \) determined with the variable J method (Equ. A7 and A8, see Appendix A). The equation representing photosynthesis limited by RuBP regeneration is:

\[
A_j = J \left[ \frac{C_c - \Gamma^*}{4C_c + 8\Gamma^*} \right] - R_d
\]  

where \( J \) is the rate of electron transport. The denominator of the above equation represents the stoichiometry of the number of electrons required to regenerate ATP and NADP; we have used four for \( C_c \) and eight for \( \Gamma^* \) (Flexas et al., 2012). \( J \) becomes \( J_{\text{max}} \) under light and CO\(_2\) saturation when the maximum possible rate of electron transport is theoretically achieved.

The photosynthetic rate limited by triose-phosphate use is estimated by:

\[
A_p = \frac{3TPU \cdot C_c}{\Gamma^* \left[ C_c - \left( \frac{1 + 3\alpha_{TPU}}{2} \right) \right]} - R_d
\]  

where TPU is the rate of triose-phosphate use at saturating CO\(_2\) concentrations, and \( \alpha_{TPU} \) is the proportion of glycerate not returned to the chloroplasts. This equation fits the A/Cc curve plateau at high concentrations of CO\(_2\) when a further increase in \( C_c \) no longer increases \( A_{net} \) or, in some cases, decreases \( A_{net} \).

These three estimated parameters \( (V_{c,\text{max}}, J_{\text{max}}, \text{and TPU}) \) define the biochemical capacity to drive the photosynthetic assimilation of CO\(_2\) but are defined here as the photosynthetic potential (Niinemets et al., 2006). The term photosynthetic capacity is here dismissed, despite its frequent use in the literature, to avoid confusion with studies that have used this term for
the maximum rate of assimilation under saturating light conditions (e.g. Bertolli and Souza, 2013).

2.8 Curve fitting

The procedure for fitting the curves to estimate the photosynthetic parameters $V_{c,\text{max}}$, $J_{\text{max}}$, and TPU applied the least square fit method using the SOLVER estimator tool in Excel. In this procedure, the squared errors of the observed points on the A/C$_c$ curve and the modelled points of Eq.(s) 6, 7, and 8 were calculated and summed. Prior to the fitting procedure, the user must assess the limiting factors, i.e. which points are allocated to which Eq. (6 or 7 or 8). The initial slope of the A/C$_c$ curve is attributed to non-saturating CO$_2$ conditions when Rubisco activity limits $A_{\text{net}}$ (Eq. 6), while the slope of the curve is smoothed at higher CO$_2$ conditions (usually > 35 Pa), representing the limitation of the regeneration of ribulose-1,5-biphosphate (RuPb) (and hence light is a limiting factor) (Eq. 7). The transition zone (approximately at 25-35 Pa of $C_i$), however, is a grey zone where one point can be attributed to either one or another limitation. These points can also introduce noise in the estimations in cases of doubt and are best discarded. Moreover, unusual points with evidence of an error during the measurements were not included in the curve-fitting procedure. At very high CO$_2$ concentrations, the A/C$_c$ curve plateaus or even decreases slightly. In this case, these points can be attributed to the limitation of triose-phosphate use (Eq. 8). The CO$_2$ response curves, however, rarely exhibit such a plateau or decrease at high CO$_2$ concentrations when working on a $C_c$ rather than a $C_i$ basis, so TPU could seldom be estimated in our study. Finally, when attributing all observed points to one or another limitation, we could then estimate the values of $V_{c,\text{max}}$ and $J_{\text{max}}$ (and possibly TPU) with the SOLVER Excel tool, which iteratively changes the three parameters to minimise the sum of squares of deviation from the observation.

2.9 Correction for diffusion leakage

Large gradients between the ambient air and the CO2 concentrations inside the chamber are created during the generation of a carbon-response curve. This leakage is particularly important at the high and low ends of the carbon-response curve when a large CO$_2$-concentration gradient exists between the leaf chamber and the surrounding ambient concentration. Based on the findings by Flexas et al. (2007a), we corrected Anet by subtracting the diffusion leakage for each step of the A/Cc curve obtained from separate response curves with leaves thermally killed in hot water.
2.10 Statistical Analyses

All statistical analyses were performed using the R software package, version 3.0.2 (http://www.r-project.org/). Differences in the parameters between the mild and cold winters were determined with Student’s t-tests ($P \leq 0.05$). Shapiro-Wilk tests of normality tested for normality of the data. Data were normalised at $P \leq 0.1$. One-factorial analyses of variance (ANOVAs) with tree species as the main factor tested for differences between tree species of the parameters in the sampling periods. Significant differences were determined at $P \leq 0.05$ with Tukey’s HSD tests. Regression analyses were conducted to study the relationship between $J_{\text{max}}$ and $V_{c,\text{max}}$ and between $J_{\text{amb}}$ and $A_{\text{net}}$. Analyses of covariance (ANCOVAs) tested for differences in slopes and intercepts.

3 Results

3.1 Environmental Variables

Collserola Natural Park experienced extremely mild winter conditions in November and December 2011 and January 2012, when average minimum temperatures (10.4 °C in November, 5 °C in December, and 3.4 °C in January) remained above 0 °C and no frosts occurred. Average maximum temperatures were 16.3 °C in November, 12.2 °C in December, and 11.4 °C in January. All species had considerable shoot growth of up to 15 cm during this mild period. Sudden low temperatures, however, led to frost on six consecutive days and a minimum average temperature of -2.3 °C (Day of the year (DOY) 21-26) followed by eight days of cool temperatures averaging +2.6 °C (DOY 27-35) (Figure 1). The average radiation during first field campaign (DOY 9-19) was 46 and during the period of frost 58 W m$^{-2}$.

3.2 Photosynthetic potentials

Of the three photosynthetic parameters describing the photosynthetic potential, $V_{c,\text{max}}$ and $J_{\text{max}}$, and TPU, only the first two could be satisfactorily estimated from the A/C$_{c}$-response curves. The leaves were only occasionally limited by TPU (6 out of 42), despite the excessive CO$_{2}$ concentrations in the higher section of the CO$_{2}$-response curve. TPU was therefore discarded from further analysis. $V_{c,\text{max}}$ and $J_{\text{max}}$ were highest in $Q$. ilex but more importantly also decreased most strongly after the period of frost by nearly 50% ($P \leq 0.05$; Fig. 2). The photosynthetic potential of $P$. halepensis was affected the least, reflected by moderate decreases in $V_{c,\text{max}}$ and $J_{\text{max}}$ (16% and 19%), which were not significant. $V_{c,\text{max}}$ and $J_{\text{max}}$ were
lowest in *A. unedo* during the mild winter period and decreased by approximately 33% after the period of frost. This decrease, however, was not significant due to a large standard error.

### 3.3 GE-derived parameters under ambient conditions

The period of frost had a strong effect on several GE-derived parameters in *Q. ilex* leaves. The cold temperatures decreased $R_n$ in *Q. ilex* leaves, but the effect was much weaker than for $R_d$ and was not significant (Fig. 3). These parameters responded very weakly to the cold and frost in the leaves of *A. unedo* and *P. halepensis*. $A_{net}$ and $\Phi_{CO2}$ were also reduced in *Q. ilex* leaves by approximately 50%. This was significant for the $A_{net}$ (Fig. 4A) and low significant for $\Phi_{CO2}$ (Fig. 4B). Further differences were only significant for $\Phi_{CO2}$ in *P. halepensis* leaves being reduced by 12% ($P \leq 0.05$). The CO₂ conductance was more strongly reduced in $g_m$ than in $g_s$ for *Q. ilex* and *A. unedo* leaves which was only significant for the former whereas these parameters seemed unaffected in *P. halepensis* leaves (Fig. 5A and 5B). As a consequence, we observed a tendency of a $C_i$- increase in parallel with a $C_c$- decrease in *Q. ilex* and *A. unedo* leaves due to a lower CO₂ uptake in carbon metabolism, but not in *P. halepensis* (Fig. 6A and 6B). The differences observed were not significant ($P \leq 0.05$).

### 3.4 CF-derived parameters under ambient conditions

The GE-derived parameters enabled us to study the immediate responses, but several CF-derived parameters allowed us to determine in more depth the physiological changes in parts of the light-harvesting apparatus, namely PSII. $F_v/F_m$ estimates the maximum quantum yield of PSII and serves as a stress indicator (Fig. 7B). *A. unedo* leaves were most strongly affected by the period of frost, followed by *Q. ilex* leaves, whereas *P. halepensis* leaves were only marginally affected. The changes were not statistically significant in the latter two species ($P \leq 0.05$). $\Phi_{PSII}$ tended to decrease in all species but most strongly in *Q. ilex* leaves (42%), however insignificantly (Fig. 7A). NPQ responded very differently in the three species. NPQ did not change much between the two sampling periods in the leaves of *P. halepensis* (6%) but decreased significantly by 25% (0.05 ≤ $P$ ≤ 0.1) in *A. unedo* leaves and tended to increase in *Q. ilex* leaves by 31% ($P \geq 0.05$), however insignificantly (Fig. 8).
3.5 Relationships of foliar photosynthetic variables

The covariance of several relationships of the foliar photosynthetic variables were analysed in an ANCOVA to test for differences in the slopes and intercepts in these relationships. The ANCOVA for the relationship between $V_{c,\text{max}}$ and $J_{\text{max}}$ in *Q. ilex* leaves indicated a highly significant ($P \leq 0.01$) reduction in the slope and also intercept showing a similar strong effect on $J_{\text{max}}$ than on $V_{c,\text{max}}$ due to the change in weather (Fig. 9A and Table 2). In *P. halepensis*, the slope was significantly and the intercept marginal significantly reduced (Fig. 9B and Table 2). This shows a comparatively stronger effect on $V_{c,\text{max}}$ than on $J_{\text{max}}$ by the cold period. The sunlit leaves of *A. unedo* and the shaded leaves of *Q. ilex* did not show any significant changes in the relationship of $V_{c,\text{max}}$ and $J_{\text{max}}$ (Fig. 9A, 9C and Table 2). The relationship between the rate of electron transport at ambient conditions derived from CF and the CO$_2$ assimilation at ambient CO$_2$ concentrations ($J_{\text{amb}}/A_{\text{net}}$) was similar in all tree species (Fig. 10A,B,C and Table 2). The slopes were higher in response to the stress imposed by the low temperatures but were not significant. When all species were combined the change of the slope was marginally significant, indicating a possible increased alternative electron sink other than carbon metabolism (Table 2).

3.6 Role of leaf position

Under mild conditions, the leaves of *Q. ilex* showed the most strongly pronounced differences in the leaf position (data of *P. halepensis* Mill. & *A. unedo* L. not shown). Leaves of *Q. ilex* growing under high irradiances had a more active carbon metabolism ($A_{\text{net}}, R_d, R_n, \Phi_{\text{CO}_2}$), photochemical efficiency ($\Phi_{\text{PSII}}$), and photosynthetic potential (high $J_{\text{max}}$ and $V_{c,\text{max}}$) in all tree species. As described in Material and Methods the effect of the leaf position after the sudden cold period was only studied for *Q. ilex*. After the sudden frost period, the photosynthetic potential was much higher in sunlit than in shaded leaves of *Q. ilex*, with both $J_{\text{max}}$ and $V_{c,\text{max}}$ being highly significant (Fig. 2 and Table 1). These differences disappeared after the cold period, because $J_{\text{max}}$ and $V_{c,\text{max}}$ in the shaded leaves remained unaffected by the frost. $F_v/F_m$ was generally higher in the shaded leaves, but not significantly ($P \leq 0.05$) (Fig. 8 and Table 1). The photosynthetic parameters under ambient conditions, such as $A_{\text{net}}, g_s, C_i, C_c$, and $g_m$, were not affected much by the leaf position (Fig. 4, 5, 6 and Table 1). Although not significant, the effects of the cold period on these parameters were stronger in the sunlit leaves. In comparison to these parameters, the leaf position had more pronounced effects on $R_n$ and $R_d$ (Fig. 3 and Table 1). The response of respiration to winter stress, however, differed
depending on the location of the leaves. \( R_n \) maintained the same balance between sunlit and shaded leaves before and after the cold period, but \( R_d \) decreased comparatively more in sunlit leaves due to the period of frost. This pattern was also reflected in \( \Phi_{CO2} \) (Fig. 4B and Table 1) and in the CF-derived parameters \( \Phi_{PSII} \) and NPQ, (Fig. 7A, 8 and Table 1) indicating a stronger effect on the photochemical machinery of sunlit leaves than on shaded leaves. Shaded leaves also exhibited a lower \( J_{amb}/A_{net} \) ratio, but the ratio increased equally in both leaf positions after the cold period, indicating a similar behaviour of dissipating energy by alternative electron sinks (Fig. 10A and Table 1).

4 Discussion

4.1 Winter in the Mediterranean region

Mediterranean-type ecosystems are exposed to stress from summer droughts but also from low temperatures in winter (Mitrakos, 1980). Less attention, however, has been paid to the degree and extent as well as the wide variation among years and regions of these stress periods, in response to which Mediterranean evergreen species have developed a dynamic photoprotective ability in order to withstand these stressors (Kyparissis et al., 2000; Martínez-Ferri et al., 2004). Despite the occurrence of lower temperatures than in spring conditions, in winter the photosynthetic potential recovered once the leaves became acclimated to the new conditions (Dolman et al., 2002; Hurry et al., 2000). This is important for the plants overall performance because the photosynthetic exploitation of favourable conditions in winter is crucial for achieving a positive carbon balance in Mediterranean evergreen tree species (García-Plazaola et al., 1999b; Martínez-Ferri et al., 2004). We showed how a long lasting comfortable winter period without frost lead to notably high photosynthetic potentials and carbon assimilation in winter being equal to or partly even exceeding spring values (Sperlich et al., unpublished data). As a result, increased winter temperatures influenced phenological responses, advanced winter cambium activation, spring bud burst and leaf unfolding which has been reported in an increasing number of studies (Peñuelas and Filella, 2001). These observations were also reflected in the high sap flow per tree \( (J_t) \), ranging for all tree species on average between 5 and 10 kg d\(^{-1}\) during the mild winter period (Sánchez et al., unpublished results). Whereas sudden frosts have often been attributed to higher altitudes of the Mediterranean region (Blumler, 1991; Tretiach et al., 1997), we showed that it can also be an important factor for plant growth and distribution in other areas such as the sub-humid Mediterranean climate of our study site (Garcia-Plazaola et al., 2003a). At night when frosts
are more likely to occur, we observed the lowest temperatures whereas at daytime the
temperatures were often above zero degrees. However, as we showed, not only cool daytime
but also cool nighttime temperatures or frosts can affect subsequent daytime photosynthesis
and induce photoprotective processes (see also Flexas et al., 1999). In our study, the sudden
occurring low temperatures affected strongly the photosynthetic apparatus, although the
responses were highly species specific. We will elucidate the physiological mechanism in the
following.

4.2 PSII – primary target of stress induced by low temperatures

Typically in winter there is an imbalance between light energy absorbed in photochemistry
and light energy used in metabolism. This is shown in our data by increased thermal energy
dissipation (NPQ) and reduced PSII efficiency ($\Phi_{PSII}$) in order to reduce the harmful effects of
excess energy reflecting an inactivation and damage of PSII reaction centres, more precisely,
the reaction-centre protein D1 (Aro et al., 1993; Demmig-Adams and Adams, 1992; Mulo et
al., 2012). More precise information about the underlying processes that have altered this
efficiency is provided by the $F_{v}/F_{m}$ ratio. Chronic changes occurring in the $F_{v}/F_{m}$ ratio can be
related to a cascade of processes which are induced to protect the photosynthetic apparatus
including i) re-organisation of the thylakoid membrane, ii) closure of reaction centres, iii)
and/or reduced antennal size (Ensminger et al., 2012; Huner et al., 1998; Maxwell and
Johnson, 2000; Verhoeven, 2014). The small changes in the $F_{v}/F_{m}$ ratio observed in the leaves
of $Q$. ilex and $P$. halepensis reflected photoprotective responses without any photodamage.
The significantly decline of $F_{v}/F_{m}$ in $A$. unedo, however, indicated strong chronic
photoinhibition and is an indication of severe photodamage (Martínez-Ferri et al., 2004). We
conclude that $A$. unedo suffered most notably from the low temperatures whereas $Q$. ilex and
$P$. halepensis were equipped with a good photoprotective capacity able to keep the
photosynthetic apparatus intact (Öquist and Huner, 2003). $Q$. ilex showed the most dynamic
responses, negating the harmful excitation stress by lowering the photochemical operating
efficiency ($\Phi_{PSII}$) and increasing the use of alternative thermal-energy pathways (NPQ). This
photoprotective capability represented by a higher NPQ is usually linked to the xanthophyll
cycle that responds to environmental factors such as temperature, water deficit, and nutrient
availability (Demmig-Adams and Adams, 1996; García-Plazaola et al., 1997). Inter-
conversions of the cycle and pool sizes occur following the need to dissipate excess excitation
energy in response to summer drought (García-Plazaola et al., 1997; Munné-Bosch and
Peñuelas, 2004), but also to winter stress (Corcuera et al., 2004; Garcia-Plazaola et al., 2003a; Kyparissis et al., 2000; Oliveira and Peñuelas, 2001). The implicit interpretation of being equipped with a high capacity of photoprotection when NPQ increases was recently questioned by Lambrev et al. (2012). This study reported that quenching and photoprotection were not necessarily linearly related and stated that several possibilities of photoprotective responses other than NPQ of CF existed, such as antennal detachment that could possibly vary with species and growth conditions. The highly dynamic and photoprotective capability of Q. ilex leaves, however, was also demonstrated by several other photosynthetic parameters such as $V_{c,\text{max}}, J_{\text{max}}, A_{\text{net}}, \Phi_{CO_2},$ and $R_d$, which confirmed this trend and were in accord with the findings by Corcuera et al. (2004). Despite reports of several mechanisms of resistance to drought stress in A. unedo, including increased levels of zeaxanthin that indicates an enhanced thermal dissipation of excess excitation energy in periods of summer stress (Munné-Bosch and Peñuelas, 2004), we found that A. unedo leaves had a lower capacity of photoprotection in response to induced over-excitation of the photosystems by winter stress.

### 4.3 High photosynthetic potentials and strong effects of low temperatures

$V_{c,\text{max}}$ and $J_{\text{max}}$ were strongly correlated (Wullschleger, 1993), being regulated in a coordinated manner above all in Q. ilex. Interestingly, the ANCOVAs indicated that $J_{\text{max}}$ decreased more strongly than did $V_{c,\text{max}}$. This is because the above described photoprotective adjustments lead to a lower energy-use efficiency in the reaction centres and consequently also to a downregulation of the photosynthetic electron transport $J_{\text{max}}$. The larger decrease of $J_{\text{max}}$ relative to $V_{c,\text{max}}$ indicated that low temperature stress became manifest first in a hampered pathway of photochemical energy, because PSII complexes are primarily affected by light-induced damage (Maxwell and Johnson, 2000; Taz and Zeiger, 2010; Vass, 2012). Hence, the limitations of the photosynthetic rate by RuBP regeneration are stronger affected by frost and cold induced stress than those by RuBP carboxylation. The relative amounts of photosynthetic proteins can probably explain the differences observed in the $J_{\text{max}}/V_{c,\text{max}}$ ratio (Hikosaka et al., 1999; Onoda et al., 2005).

The physiological responses were highly species-specific. Q. ilex leaves responded with significant decreases (approximately 50%) in their photosynthetic potentials (both $V_{c,\text{max}}$ and $J_{\text{max}}$). In contrast, $V_{c,\text{max}}$ and $J_{\text{max}}$ decreased in P. halepensis leaves by only 16 and 19%, respectively, and in A. unedo leaves by approximately 30% (for both parameters).
4.4 Inhibition of carbohydrate metabolism

As demonstrated above, adjustments to the frost event took place via the energy flow in the antennal systems and a downregulation of photosynthetic electron transport as well as regulatory mechanisms including the inhibition of Rubisco activity, but also via stomatal and mesophyllic diffusion behaviour (Ensminger et al., 2012; Gratani et al., 2000; Taz and Zeiger, 2010). Interestingly, the mesophyllic diffusion resistance was stronger pronounced as a response to low temperatures, especially in Q. ilex reducing the CO₂ available for fixation in the chloroplasts. This underlines the recently growing awareness in the scientific community about the important role of $g_m$ as an additional regulating parameter as response to stress, above all in sclerophyllic species (Flexas et al., 2008; Niinemets et al., 2011). In general, our results demonstrated that the efficiency of carbon use in the photosynthetic metabolism and foliar respiratory responses were highly species dependant (Zaragoza-Castells et al. 2007, 2008). For instance, P. halepensis and Q. ilex leaves depicted extraordinarily high values of $A_{net}$, $R_d$, $R_n$, and $\Phi_{CO2}$ in the mild winter period, but only Q. ilex exhibited a significant downregulation after the frost event. The downregulation of photosynthesis, the most efficient process to get rid of excess energy, suggests alternative energy pathways such as photorespiration. We did not measure photorespiration directly, but we could infer some of its characteristics by studying the relationship between $J_{amb}$ and $A_{net}$. All tree species had a relatively higher proportion of electron flux during the period that can be explained by utilization in the carbon metabolism. This has been mainly attributed to photorespiration, but also to the Mehler reaction that protects plants from photodamage in bright light (Allen and Ort, 2001; D’Ambrosio et al., 2006; Flexas et al., 1998, 1999; Fryer et al., 1998; Huner et al., 1998).

4.5 Leaf position specific responses to abiotic stress in winter

It is well known that leaves growing under high irradiances have a more active carbon metabolism ($A_{net}$, $R_d$, $R_n$, and $\Phi_{CO2}$), photochemical efficiency ($\Phi_{PSII}$), and photosynthetic potential (high $J_{max}$ and $V_{c, max}$) (Taz and Zeiger, 2010). Hereby, Q. ilex showed the most strongly pronounced differences between sunlit and shaded leaves. Plants develop leaves with a highly specialised anatomy and morphology for the absorption of the prevailing light in their local environments resulting generally in smaller but also thicker sunlit leaves (Kull and Niinemets, 1993; Terashima and Hikosaka, 1995). Nevertheless, the higher carbon metabolism and photochemical activity of sunlit leaves decreased strongly, partly below the...
level of shaded leaves, whereas shaded leaves showed little sign of any downregulation but maintained a relatively stable effective quantum yield of CO₂ assimilation in both periods. Furthermore, the photosystems showed no sign of photodamage and generally maintained a higher maximum efficiency than did sunlit leaves. We concluded that foliar-level physiology during winter was better protected in the shaded crown of *Q. ilex* unexposed to the dramatic changes in radiation in the outer canopy, confirming the results by Valladares et al. (2008).

We also concluded that *Q. ilex* is a highly dynamic species able to rapidly change its metabolism on the antioxidant and photoprotective level in dependence to its leaf position (García-Plazaola et al., 1997, 1999a; Martínez-Ferri et al., 2004). We show that the foliar plasticity in morphology and anatomy of *Q. ilex* (Bussotti et al., 2002; Valladares et al., 2000) can also be attributed to its biochemical metabolism. We stress that the solar environment of the leaves is a crucial factor when assessing tree performance, especially when comparing tree species in a competitive context.

### 4.6 Ecological context

*Q. ilex* had the most drastic photoprotective response to frost and cool temperatures, whereas *P. halepensis* exhibited a homeostatic behaviour with a very active carbon assimilatory and respiratory metabolism in both periods. *A. unedo* was intermediate, with large decreases in the parameters of carbon metabolism but also a high variability in its response to frost. *A. unedo* also had the lowest photoprotective capability, which might be explained by previous characterisations to be semi-deciduous to drought being at the borderline to evergreen sclerophyllous species (Gratani and Ghia, 2002a, 2002b). Moreover, *A. unedo* occurs naturally most commonly as a shrub and is less frequently found in the forest canopy of mixed forests growing up to 8-10 m tall as in our study site (Beyschlag et al., 1986; Reichstein et al., 2002). Investments in leaves are thus lower and leaf longevity shorter. Leaves of *A. unedo* are more rapidly replaced relative to more sclerophyllic leaves such as those of *Q. ilex*. We postulated that *A. unedo*, considered a relict of the humid-subtropical Tertiary tree flora, was more sensitive to winter stress, which is consistent with its presence mostly in the western Mediterranean basin and its frequent occurrence in coastal zones where humidity and temperature are the main factors determining its geographical distribution (Gratani and Ghia, 2002a and references therein). Our results suggested that *Q. ilex* could greatly benefit from favourable winter conditions exhibiting a high photosynthetic potential and carbon metabolism. Angiosperms are known to make efficiently use of favourable winter
periods to recover depleted carbon reserves and embolism induced loss of hydraulic capacity (Carnicer et al., 2013 and references therein). When these relatively favourable conditions changed, *Q. ilex* quickly re-adjusted the photosynthetic machinery to the prevailing conditions, as indicated by the largest decreases in photosynthetic potential and carbon metabolism. Some researchers have proposed the lutein-epoxy cycle in photoprotection of *Quercus* as a mechanism to maintain sustained energy dissipation (Garcia-Plazaola et al., 2003b), which could help to account for the higher tolerance to low temperatures in *Q. ilex* relative to other co-occurring Mediterranean trees or shrubs (Ogaya and Peñuelas, 2003, 2007). *P. halepensis* did not suffer a pronounced chronic photoinhibition, confirming the results by Martínez-Ferri *et al.* (2004). Despite a pronounced downregulation of photosynthetic electron transport and an increase in alternative electron sinks, the light-saturated ambient photosynthesis and stomatal conductance remained surprisingly high and constant. *P. halepensis* thus exhibited a successful refinement of photosynthetic electron flow and possibly a successful repair of protein D1 in the PSII reaction centre. The strong downregulation in *Q. ilex* and the homogenous response of *P. halepensis* were possibly due to distinct, previously described strategies. *Q. ilex* has been characterised as a photoinhibition-avoiding species and *P. halepensis* as a photoinhibition-tolerant species (Martinez-Ferri *et al.*, 2000). We have extended this categorisation for *A. unedo*, a less photoinhibition-tolerant tree species, which favoured carbon metabolic processes at the cost of chronic photoinhibition and photodamage. This strategy is similar to those in other semi-deciduous shrubs (Oliveira and Peñuelas, 2001; Oliveira and Peñuelas, 2004). The physiological responses of *Q. ilex*, a slowly growing late-successional species, to environmental stressors are highly plastic (Zavala *et al.*, 2000) due to its vegetative activity in a wide range of temperatures and high stomatal control in stressful conditions (Gratani *et al.*, 2000; Savé *et al.*, 1999), high plasticity index and resprouting dynamics (Espelta *et al.*, 1999; Gratani *et al.*, 2000), deep rooting system and large carbohydrate pools (Canadell and Lopez-Soria, 1998; Canadell *et al.*, 1999), and high adaptive variability in foliar phenomorphology (Sabaté *et al.*, 1999). Our findings showed the intra-crown variability in *Q. ilex*, where shaded leaves were widely unaffected by the inhibitory cold stress (Oliveira and Peñuelas, 2001). The ability of *Q. ilex* to perform rapid metabolic changes in the antioxidant and photoprotective mechanisms could be of adaptive importance (García-Plazaola *et al.*, 1999a). In contrast, *P. halepensis* is a fast growing conifer that quickly occupies open spaces after disturbances such as fires (Zavala *et al.*, 2000). *P. halepensis*, as do all pines, has a low ability to store carbohydrates and therefore follows a
strategy of water conservation and embolism avoidance (Meinzer et al., 2009). High rates of
photosynthesis and growth require high concentrations of carboxylation enzymes in the
carbon cycle that have high maintenance costs (Valladares and Niinemets, 2008), perhaps
accounting for the high respiration rates found in *P. halepensis* leaves. Moreover, differences
among the species are also likely to be the result of distinct foliar morphologies and crown
architectures. Pine trees are characterised by a relatively low exposure of foliar surface area to
direct sunlight due to the cylindrical shape and steep angles of their needles but at the same
time are able to exploit a wider range of incident light angles than broadleaved trees. Despite
reported flexible adjustments in the orientation of the leaves in several Mediterranean
broadleaved sclerophyllous species (Oliveira and Peñuelas, 2000; Vaz et al., 2011; Werner et
al., 2002), needle leaves probably still confer some benefits to attain near-saturated
photosynthetic rates over a wider range of diurnal and seasonal variation in sun angles (Jordan
and Smith, 1993; Lusk et al., 2003), while at the same time showing a high tolerance to
photoinhibition. This might account for the good performance of *P. halepensis* under mild
winter conditions with moderate abiotic stresses such as in our study. However, under more
severe and re-occurring frost events, *P. halepensis* might reach the threshold of its tolerance
and severe frost damage can occur. This explains also its absence in mountain regions with
more severe winters where *Q. ilex* becomes more competitive. Despite following distinct
physiological strategies, both *Q. ilex* and *P. halepensis* seem to cope equally well with the
winter conditions they were exposed to whereas the foliar photosynthetic systems of *A. unedo*
were more sensitive to sudden frost impacts. Thus, *A. unedo* might have been in a competitive
disadvantage for the following growing season.

Overall, we conclude that the photosynthetic exploitation of relatively favorable winter
conditions might be crucial for evergreen Mediterranean tree species for achieving a positive
annual carbon balance. The winter period might give important insights helping to explain the
dynamics of Mediterranean forest communities when withstanding increased novel
environmental conditions projected in multiple climate change scenarios and benefitting from
periods of potential recovery and growth in winter.

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References


Hurry, V., Strand, a, Furbank, R. and Stitt, M.: The role of inorganic phosphate in the development of freezing tolerance and the acclimatization of photosynthesis to low


Fig. 2

Panel A: Bar graph showing $V_{c_{\text{max}}}$ (µmol CO$_2$ m$^{-2}$ s$^{-1}$) for different conditions.

Panel B: Bar graph showing $J_{\text{max}}$ (µmol electron m$^{-2}$ s$^{-1}$) for different conditions.
Fig. 3

(A) $R_n$ ($\mu$mol CO$_2$ m$^{-2}$ s$^{-1}$)

(B) $R_d$ ($\mu$mol CO$_2$ m$^{-2}$ s$^{-1}$)

Legend:
- Q. ilex (sun)
- Q. ilex (shade)
- P. halepensis
- A. unedo
Fig. 4

A) $A_{\text{net}}$ (µmol CO$_2$ m$^{-2}$ s$^{-1}$)

- Q. flex (sun)
- Q. flex (shade)
- P. halepensis
- A. uneo

B) $Q_{\text{CO}_2}$

- mild
- frost

Values indicate percentage change compared to the control.
Fig. 5
Fig. 6

The figure illustrates the variation in carbon dioxide uptake (C) in different conditions. A) shows the C uptake in mild and frost conditions, with a 22% increase and a 2% decrease, respectively. B) presents the C uptake in different species under mild and frost conditions, with varying increases and decreases in uptake percentages.
Fig. 8
Fig. 9

Comparison of $J_{\text{max}}$ (µmol electron m$^{-2}$s$^{-1}$) and $V_{C_{\text{max}}}$ (µmol CO$_2$ m$^{-2}$s$^{-1}$) for different conditions:

A) [Plot Description]
B) [Plot Description]
C) [Plot Description]
Fig. 10

![Figure 10](image-url)
Tables

Table 1: *P* values of Student’s *t*-tests for the differences between sunlit and shaded leaves of *Q. ilex*.

<table>
<thead>
<tr>
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<th>Both periods</th>
<th>Mild period</th>
<th>Frost period</th>
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<td><em>V</em>&lt;sub&gt;c,max&lt;/sub&gt;</td>
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<tr>
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### Table 2. Regression coefficients and results from ANCOVA analyses of the $J_{\text{amb}}/A_{\text{net}}$ and $J_{\text{max}}/V_{c,\text{max}}$ relationships.

#### Regression analyses of $J_{\text{max}}$ & $V_{c,\text{max}}$

<table>
<thead>
<tr>
<th>tree species</th>
<th>Q. ilex</th>
<th>Q. ilex</th>
<th>P. halepensis</th>
<th>A. unedo</th>
<th>all species</th>
</tr>
</thead>
<tbody>
<tr>
<td>leaf position</td>
<td>sunlit</td>
<td>shaded</td>
<td>sunlit</td>
<td>sunlit</td>
<td>sunlit</td>
</tr>
<tr>
<td>mild</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>reg. line</td>
<td>$y = 0.81 x + 41.6$</td>
<td>$y = 1.2 x + 6.1$</td>
<td>$y = 115.9 x + 148.8$</td>
<td>$y = 0.954 x + 31.5$</td>
<td>$y = 50.2 x + 0.77$</td>
</tr>
<tr>
<td>$R^2$</td>
<td>0.97</td>
<td>0.48</td>
<td>0.32</td>
<td>0.95</td>
<td>0.94</td>
</tr>
<tr>
<td>$P$</td>
<td>2E-04</td>
<td></td>
<td></td>
<td>0.193</td>
<td></td>
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<tr>
<td>frost</td>
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<td></td>
<td></td>
</tr>
<tr>
<td>reg. line</td>
<td>$y = 0.94 x + 3.6$</td>
<td>$y = 1.89 x - 9.19$</td>
<td>$y = 971 x + 9.9$</td>
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<td></td>
</tr>
<tr>
<td>$R^2$</td>
<td>0.89</td>
<td>0.035</td>
<td>0.53</td>
<td>0.91</td>
<td></td>
</tr>
<tr>
<td>$P$</td>
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<td></td>
<td>0.29</td>
<td></td>
</tr>
<tr>
<td>p (slope)</td>
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<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>p (intercept)</td>
<td>0.3</td>
<td>0.022</td>
<td>0.28</td>
<td>0.008</td>
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</tr>
</tbody>
</table>

#### Regression analyses of $J_{\text{amb}}$ & $A_{\text{net}}$

<table>
<thead>
<tr>
<th>tree species</th>
<th>Q. ilex</th>
<th>Q. ilex</th>
<th>P. halepensis</th>
<th>A. unedo</th>
<th>all species</th>
</tr>
</thead>
<tbody>
<tr>
<td>leaf position</td>
<td>sunlit</td>
<td>shaded</td>
<td>sunlit</td>
<td>sunlit</td>
<td>sunlit</td>
</tr>
<tr>
<td>mild</td>
<td></td>
<td></td>
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<td></td>
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</tr>
<tr>
<td>reg. line</td>
<td>$y = 10.8 + 56.1$</td>
<td>$y = 3.1 x + 39$</td>
<td>$y = 9.22 x + 58.3$</td>
<td>$y = 9.7 x + 54.9$</td>
<td>$y = 10.9 x + 51.9$</td>
</tr>
<tr>
<td>$R^2$</td>
<td>0.76</td>
<td>0.91</td>
<td>0.51</td>
<td>0.96</td>
<td>0.84</td>
</tr>
<tr>
<td>$P$</td>
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<td></td>
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<td>0.005</td>
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</tr>
<tr>
<td>frost</td>
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<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>reg. line</td>
<td>$y = 15.4 x + 21.1$</td>
<td>$y = 2.7 x + 46.8$</td>
<td>$y = 11.9 x + 31.9$</td>
<td>$y = 14.6 x + 14.2$</td>
<td>$y = 13.5 x + 22.3$</td>
</tr>
<tr>
<td>$R^2$</td>
<td>0.73</td>
<td>-0.13</td>
<td>0.52</td>
<td>0.46</td>
<td>0.76</td>
</tr>
<tr>
<td>$P$</td>
<td>0.093</td>
<td></td>
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<td>0.200</td>
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</tr>
<tr>
<td>p (slope)</td>
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<td>0.72</td>
<td>0.59</td>
<td>0.322</td>
<td>0.098</td>
</tr>
<tr>
<td>p (intercept)</td>
<td>0.51</td>
<td>0.45</td>
<td>0.31</td>
<td>0.29</td>
<td>0.071</td>
</tr>
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</table>
Figure 1. Maximum and minimum temperatures on the primary y-axes (in red squares and circles, respectively) and radiation (in yellow crosses) on the secondary y-axes are presented for the mild and frost winter period for the day of the year (DOY) in January and February 2012.

Figure 2. Bar plot of the effect of a sudden period of frost following a mild winter period in 2012 on A) the maximum velocity of carboxylation ($V_{c,max}$) and B) the maximum rate of electron transport ($J_{max}$) in sunlit leaves of Q. ilex (light green bar), in shaded leaves of Q. ilex (dark green bar), P. halepensis (beige bar), and A. unedo (blue bar). The error bars represent the standard error, and the percentages indicate the change between periods where significance is indicated with an asterisk ($P \leq 0.05$) and marginal significance with an asterisk in brackets ($0.05 \geq P \leq 0.1$).

Figure 3. Bar plot of the effect of a sudden period of frost following a mild winter period on A) nighttime respiration ($R_n$) and B) daytime respiration ($R_d$) in sunlit leaves of Q. ilex (light green bar), in shaded leaves of Q. ilex (dark green bar), P. halepensis (beige bar), and A. unedo (blue bar). The error bars represent the standard error, and the percentages indicate the change between periods where significance is indicated with an asterisk ($P \leq 0.05$) and marginal significance with an asterisk in brackets ($0.05 \geq P \leq 0.1$).

Figure 4. Bar plot of the effect of a sudden period of frost following a mild winter period on A) net assimilation ($A_{net}$) and B) the effective quantum yield of net CO$_2$ assimilation ($\Phi_{CO_2}$) in sunlit leaves of Q. ilex (light green bar), in shaded leaves of Q. ilex (dark green bar), P. halepensis (beige bar), and A. unedo (blue bar). The error bars represent the standard error, and the percentages indicate the change between periods where significance is indicated with an asterisk ($P \leq 0.05$) and marginal significance with an asterisk in brackets ($0.05 \geq P \leq 0.1$).

Figure 5. Bar plot of the effect of a sudden period of frost following a mild winter period on A) mesophyllic conductance ($g_m$) and B) stomatal conductance ($g_s$) in sunlit leaves of Q. ilex (light green bar), in shaded leaves of Q. ilex (dark green bar), P. halepensis (beige bar), and A. unedo (blue bar). The error bars represent the standard error, and the percentages indicate the change between periods where significance is indicated with an asterisk ($P \leq 0.05$) and marginal significance with an asterisk in brackets ($0.05 \geq P \leq 0.1$).

Figure 6. Bar plot of the effect of a sudden period of frost following a mild winter period on A) the stomatal internal CO$_2$ concentration ($C_i$) and B) the chloroplastic CO$_2$ concentration.
(Cₜ) in sunlit leaves of *Q. ilex* (light green bar), in shaded leaves of *Q. ilex* (dark green bar), *P. halepensis* (beige bar), and *A. unedo* (blue bar). The error bars represent the standard error, and the percentages indicate the change between periods where significance is indicated with an asterisk (*P*≤0.05) and marginal significance with an asterisk in brackets (0.05≥*P*≤0.1).

**Figure 7.** Bar plot of the effect of a sudden period of frost following a mild winter period on A) the effective quantum yield of photosystem II (Φₚₛᵢˡ) and B) the maximum efficiency of photosystem II (*Fᵥ/Fₘ*) in sunlit leaves of *Q. ilex* (light green bar), in shaded leaves of *Q. ilex* (dark green bar), *P. halepensis* (beige bar), and *A. unedo* (blue bar). The error bars represent the standard error, and the percentages indicate the change between periods where significance is indicated with an asterisk (*P*≤0.05) and marginal significance with an asterisk in brackets (0.05≥*P*≤0.1).

**Figure 8.** Bar plot of the effect of a sudden period of frost following a mild winter period on non-photochemical quenching (NPQ) in sunlit leaves of *Q. ilex* (light green bar), in shaded leaves of *Q. ilex* (dark green bar), *P. halepensis* (beige bar), and *A. unedo* (blue bar). The error bars represent the standard error, and the percentages indicate the change between periods where significance is indicated with an asterisk (*P*≤0.05) and marginal significance with an asterisk in brackets (0.05≥*P*≤0.1).

**Figure 9.** Relationship between the maximum velocity of carboxylation (*Vₖ,ₘₐₓ*) and the maximum rate of electron transport (*Jₖₘₐₓ*) in *Q. ilex* (A), *P. halepensis* (B), *A. unedo* (C), leaves. Leaves measured under mild conditions are indicated by green circles and cyan triangles in shaded and sunlit locations, respectively. Leaves measured after the period of frost are indicated by green diamonds and blue squares in shaded and sunlit locations, respectively.

**Figure 10.** Relationship between the rate electron transport from chlorophyllic fluorescence (*Jₐₘ₉*) and net assimilation (*Aₙₑₙ*) at ambient CO₂ concentrations and saturating light (*Aₙₑₙ*) in *Q. ilex* (A), *P. halepensis* (B), *A. unedo* (C), leaves. Leaves measured under mild conditions are indicated by green circles and cyan triangles in shaded and sunlit locations, respectively. Leaves measured after the period of frost are indicated by green diamonds and blue squares in shaded and sunlit locations, respectively.
Appendix A

Temperature functions

The effective Michaelis-Menten constants $K_c$ and $K_o$ and the photorespiratory compensation point, $\Gamma^*$, were taken from (Bernacchi et al., 2002) and are summarized in Table 3. The following generic temperature response functions were used to adjust these parameters to the prevailing $T_{Leaf}$ during the experiments

$$K_c = e^\left(\frac{\Delta Ha}{R \times (273.15 + T_{Leaf})}\right)$$  \hspace{1cm} (A1)

$$K_o = e^\left(\frac{\Delta Ha}{R \times (273.15 + T_{Leaf})}\right)$$  \hspace{1cm} (A2)

$$\Gamma^* = e^\left(\frac{\Delta Ha}{R \times (273.15 + T_{Leaf})}\right) \times \frac{O_2}{20.9}$$  \hspace{1cm} (A3)

where $R$ is a unitless gas constant (0.008314), $c$ is a scaling constant, $\Delta Ha$ represents the activation energy and $O_2$ is the oxygen concentration of the ambient air assumed to be 20.9 kPa.

CF- parameters

$$NPQ = \frac{(F_m - F_m'\prime)}{F_m\prime}$$  \hspace{1cm} (A4)

Photochemical quenching (qP) indicates the proportion of open PSII reaction centres and tends to be highest in low light when leaves use light most efficiently (Maxwell and Johnson, 2000). qP was estimated by:
\[ qP = \frac{F_{m'} - F_s}{F_{m'} - F_o'} \]  

(A5)

where \( F_o' \) is the minimum fluorescence in a light-adapted leaf after a pulse of darkness. The rate of photosynthetic electron transport, \( J_{CF} \), is calculated by (Genty et al., 1989):

**Estimation of mesophyll conductance**

The CO₂ pathway leads from the atmosphere to the intercellular air spaces through the stomata and from there diffuses through the air spaces of the mesophyll, cell walls, cytosol, and chloroplastic envelopes and finally reaches the sites of CO₂ fixation in the chloroplastic stroma where it is fixed by ribulose-1,5-bisphosphate carboxylase/oxygenase (Rubisco). In this study, we call this pathway the internal mesophyll diffusion conductance \( (g_m) \) and estimate it with the variable-\( J \) method by Harley et al. (1992):

\[ g_m = \frac{A_{net}}{C_i - \frac{[\Gamma^* J_{CF} + 8(A_{net} + R_d)]}{J_{CF} - 4(A_{net} + R_d)}} \]  

(A7)

where \( \Gamma^* \) is the CO₂ concentration at which the photorespiratory efflux of CO₂ equals the rate of photosynthetic uptake of CO₂ (Table 3). Similarly to \( g_s \), \( g_m \) is defined as a unitless molar fraction, rendering the units for conductance the same as those for photosynthesis. Nonetheless, the drawdown of CO₂ from the intercellular airspaces to the sites of carboxylation is thought to be dominated by the liquid phase of the chloroplast and is hence dependent on the partial pressure of the gas according to Henry's law (Harley et al., 1992). The units for conductance (mol m\(^{-2} \) s\(^{-1} \) bar\(^{-1} \)) are thus directly comparable to \( g_s \) when the atmospheric pressure is 1 bar. We assumed normal pressure (1.01325 bar) in our experiments that were conducted in Barcelona, which is close to sea level. The variable-\( J \) method accounts for the variation in \( g_m \) with \( C_i \) and provides more accurate estimates of photosynthetic parameters than do \( A/C_c \) curves that assume a constant \( g_m \), especially during episodes of water stress (Flexas et al., 2007). The chloroplastic CO₂ concentration can then be determined using

\[ C_c = C_i - \frac{A_{net}}{g_m} \]  

(A8)

where \( C_c \) is the chloroplastic CO₂ concentration.
Appendix C

Table C1. The scaling constant (c) and energies of activation (ΔHa) describing the temperature responses for Rubisco enzyme kinetic parameters $K_c$, $K_o$, and $\Gamma^*$. Taken from Bernacchi et al., (2002).

<table>
<thead>
<tr>
<th></th>
<th>25°C</th>
<th>c</th>
<th>ΔHa</th>
<th>unit</th>
</tr>
</thead>
<tbody>
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<td>80.99</td>
<td>Pa</td>
</tr>
<tr>
<td>$K_o$</td>
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<td>12.38</td>
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<td>kPa</td>
</tr>
<tr>
<td>$\Gamma^*$</td>
<td>3.74</td>
<td>11.19</td>
<td>24.46</td>
<td>Pa</td>
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</table>