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Nitrate and dissolved nitrous oxide in groundwater within cropped fields and riparian buffers

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Received: 15 September 2008 - Accepted: 17 September 2008 - Published: 13 January 2009

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Published by Copernicus Publications on behalf of the European Geosciences Union.

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Abstract

Transport and fate of dissolved nitrous oxide (N₂O) in groundwater and its significance to nitrogen dynamics within agro-ecosystems are poorly known in spite of significant potential of N₂O to global warming and ozone depletion. Increasing denitrification in riparian buffers may trade a reduction in nitrate (NO₃⁻) transport to surface waters

- for increased N₂O emissions resulting from denitrification-produced N₂O dissolved in groundwater being emitted into the air when groundwater flows into a stream or a river. This study quantifies the transport and fate of NO₃⁻ and dissolved N₂O moving from crop fields through riparian buffers, assesses whether groundwater exported from crop
- fields and riparian buffers is a significant source of dissolved N₂O emissions, and evaluates the Intergovernmental Panel on Climate Change (IPCC) methodology to estimate dissolved N₂O emission. We measured concentrations of NO₃⁻; chloride (Cl⁻); pH; dissolved N₂O, dissolved oxygen (DO), and organic carbon (DOC) in groundwater under a multi-species riparian buffer, a cool-season grass filter, and adjacent crop fields located
- ¹⁵ in the Bear Creek watershed in central Iowa, USA. In both the multi-species riparian buffer and the cool-season grass filter, concentrations of dissolved N₂O in the groundwater did not change as it passed through the sites, even when the concentrations of groundwater NO₃⁻ were decreased by 50% and 59%, respectively, over the same periods. The fraction of N lost to leaching and runoff (0.05) and the modified N₂O emission
- $_{20}$ factor, [ratio of dissolved N_2O flux to N input (0.00002)] determined for the cropped fields indicate that the current IPCC methodology overestimates dissolved N_2O flux in the sites. A low ratio between dissolved N_2O flux and soil N_2O emission (0.0003) was estimated in the cropped fields. These results suggest that the riparian buffers established adjacent to crop fields for water quality functions (enhanced denitrification)
- $_{25}$ decreased NO_3^- and were not a source of dissolved N₂O. Also, the flux of dissolved N₂O from the cropped field was negligible in comparison to soil N₂O emission in the crop fields.

1 Introduction

Nitrous oxide is a potent greenhouse gas (Wang et al., 1976) with a global warming potential that is 298 times that of carbon dioxide (CO_2) and 25 times that of methane (CH_4) over a 100-year time horizon (Forster et al., 2007). In groundwater under agricul-

- ⁵ tural fields receiving N applications, or in riparian zones receiving groundwater or runoff water, excessive NO₃⁻ may be transformed to N₂O through the process of denitrification (Mosier et al., 1998; Nevison, 2000; IPCC, 2006). Denitrification is recognized as a major mechanism for decreasing NO₃⁻ in riparian buffers (e.g. Groffman and Hanson, 1997; Watts et al., 2000). Denitrification is controlled by the availability of oxygen (O₂),
- ¹⁰ NO_3^- , carbon (C), and the population of dentrifying organisms (e.g. Hill et al., 2000; Hill and Cardaci, 2004). Riparian buffers, which provide a C-rich environment, can increase denitrification directly by enhancing the availability of C to denitrifiers, and indirectly, through increasing the consumption of O_2 by heterotrophic microbes (e.g. Groffman, 1994; Hill, 1996). Since numerous studies have recognized that NO_3^- concentrations
- ¹⁵ in groundwater decrease as a result of increased denitrification in the riparian buffers (e.g. Groffman and Hanson, 1997; Watts et al., 2000), it has been hypothesized that the increased denitrification may be trading a decrease in NO₃⁻ transport to surface waters for increased N₂O emissions (Groffman et al., 1998 and 2000), that is, trading water pollution for atmospheric pollution. In contrast, because riparian buffers effi-
- ciently decrease NO₃⁻, a source of N₂O emissions, riparian buffers could provide an opportunity to decrease dissolved N₂O emissions if we can develop reliable strategies for decreasing N₂O production during denitrification (Groffman, 2000). Studies supporting this proposition include Blicher-Mathiesen and Hoffman (1999), who reported that denitrification in a riparian soil can act as a sink for dissolved N₂O in the inflowing
- groundwater as well as for N₂O produced internally. However, very few studies have addressed these issues and the data that can be utilized to evaluate these possibilities are extremely limited. Clearly, there is a need to evaluate processes influencing production and consumption of dissolved N₂O in different riparian buffers and to assess

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the potential to decrease the emissions (Groffman et al., 2000).

The resulting N₂O dissolves in groundwater, which flows into streams, rivers, and estuaries and is ultimately emitted into the atmosphere (Mosier et al., 1998; Nevison, 2000; IPCC, 2006). The Intergovernmental Panel on Climate Change (IPCC,

- $_{\rm 5}$ 2006) defines emission of dissolved N₂O as indirect N₂O emissions, in contrast to direct N₂O emissions from N sources such as fertilizers and crop residues in managed soils. The IPCC (1997, 2006) estimates indirect N₂O emissions from N leaching in agro-ecosystems multiplying N inputs by fraction of all N lost to leaching and runoff (Frac_{LEACH-(H)}) and emission factor for N₂O emissions from N leaching and runoff
- ¹⁰ (EF₅). The ÉF₅ consists of emission factors for groundwater (EF_{5g}), rivers (EF_{5r}), and estuaries (EF_{5e}). Owing to complexity of N cycle in agro-ecosystems and the limitation of available data, $\operatorname{Frac}_{\operatorname{LEACH}-(H)}$ and EF₅ have significant uncertainties. The default value of $\operatorname{Frac}_{\operatorname{LEACH}-(H)}$ currently used by IPCC (2006) is 0.3, and an uncertainty range 0.1–0.8 was reported by Seitzinger and Kroeze (1998); however, lower values
- ¹⁵ of 0.15-0.2 have been substituted for the default by several countries (Nevison, 2000). Emission factors for groundwater, EF_{5g} are derived from the ratio between dissolved N₂O and NO₃⁻ concentrations (IPCC 1997, 2006). Since N₂O emission factor is typically defined by the ratio between N₂O emission and N input (IPCC 2006), Weymann et al. (2008) proposed the ratio between dissolved N₂O and initial NO₃⁻ concentrations
- to reflect the N input and show the emission factor determined by the new concept was smaller than the emission factor determined by current EF_{5g} concept. Since N₂O concentration in groundwater does not necessarily reflect actual indirect N₂O emission (Höll et al., 2005) and spatial and temporal heterogeneity of N₂O concentration is high (Weymann et al., 2008), the emission factor proposed by Weymann et al. (2008) still
- ²⁵ has uncertainties for estimating indirect N₂O emission. Beyond these technical issues, it has been questioned whether indirect N₂O emission is a significant pathway in the N cycle (Davidson and Swank 1990; Ueda et al., 1991; Harrison and Matson, 2003; Reay et al., 2004; Höll et al., 2005; Well at al., 2005). Clearly, studies are needed to improve Frac_{LEACH-(H)} and dissolved N₂O emission factor, and evaluate the contribution

of dissolved N₂O in the N cycle.

The overarching objective of this study was to quantify dissolved N₂O in groundwater moving from row-cropped fields through riparian buffers of two vegetation types and to relate these patterns to observed patterns of groundwater NO₃⁻ concentration. Spe-

⁵ cific objectives were to quantify transport and fate of NO₃⁻ and dissolved N₂O in crop fields and riparian buffers, assess whether groundwater exported from crop fields and riparian buffers is a significant source of dissolved N₂O, and evaluate the validity of Frac_{LEACH-(H)} and the dissolved N₂O emission factor.

2 Material and methods

10 2.1 Study site

The study was conducted on two sites within the Bear Creek watershed, Story County and Hamilton County, Iowa, United States of America (42° 11′ N, 93°30′ W). Bear Creek (total length 56 km) is a third order stream with typical discharges of 0.3 to $1.4 \text{ m}^3 \text{ s}^{-1}$. The watershed drains 6810 ha of farmland, with nearly 90% of these acres in maize-

- soybean rotation. The study area was once a tallgrass prairie ecosystem containing wet prairie marshes and pothole wetlands in topographically low areas and forests along higher order streams. An ongoing objective of the Bear Creek watershed project has been to establish riparian buffers along the upper portions of the watershed as willing landowners and cost-share opportunities are identified (Schultz et al., 2004).
- This has provided a variety of sites of different streamside vegetation and buffer age to utilize in assessing the spatial and temporal variability of riparian buffers in reducing nonpoint source pollution. This study was conducted in two riparian buffers established in 1990 on opposite sides of Bear Creek (Fig. 1). One site is an established cool-season grass filter (length 35 m×width 20 m) along the north side of the creek. The
- ²⁵ dominant grass species in this cool-season grass filter are smooth brome (*Bromus inermis* Leysser), timothy (*Phleum pretense* L.), and Kentucky bluegrass (*Poa pratensis*

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L.). The other site is a multi-species riparian buffer (length 35 m×width 20 m) which consists of a forested buffer and a warm-season grass filter along the south side of the creek. Hybrid poplars (*Populus X euroamericana' Eugenei*), ninebark (*Physocarpus opulifolius* (L.) Maxim), and redosier dogwood (*Cornus sericea* L.) were planted in the

- ⁵ forest buffer. Switchgrass (*Panicum virgatum L*.), a native warm-season grass, was planted as a grass filter adjacent to the crop field. Details of the riparian buffer design, placement, and plant species are given in Schultz et al. (1995). The upslope crop fields are farmed in an annual maize-soybean rotation. Maize (*Zea mays* L.) usually was planted in early May and harvested at the end of October. The soybean crop (*Glycine*
- max (L.) Merr.) was planted in mid-May and harvested in mid-September. The study sites are on Coland soil (fine-loamy, mixed, mesic Cumulic Haplaquoll) which is well drained to poorly drained and formed from till or local alluvium and colluvium derived from till (DeWitt, 1984). The sites are underlined by alluvium of the DeForest Formation, which consists of a sand aquifer 2 m thick overlain by 1.5 m of loam (Spear, 2003).
- At each site, 12 monitoring wells were installed in three transects from the crop field edge to the creek along proposed groundwater flow paths, and a stilling well was installed to record the surface water elevation of the creek (Simpkins et al., 2002) (Fig. 1). At each site, 3 monitoring wells at the crop field edge of the buffers (cool-season grass filter: R1, R9, R39; multi-species riparian buffer: R16, R20, R24) and 3 monitoring
- wells (cool-season grass filter: R8, R12, R40; multi-species riparian buffer: R13, R17, R21) and a stilling well (cool-season grass filter: SWRN; multi-species riparian buffer: SWRS) at the creek edge of the buffers were used (Fig. 1).

2.2 Groundwater sampling and monitoring

Groundwater sampling and monitoring was conducted monthly in monitoring wells and stilling wells from November 2005 to April 2008 (Fig. 1). To determine water table elevation prior to sampling, hydraulic head was measured with an electronic water level tape. For measurement of NO₃⁻ and Cl⁻, groundwater was collected in polyethylene bottles using a peristaltic pump. For measurement of DOC, groundwater was collected in glass bottles. Samples for NO₃⁻ and DOC were acidified with 20 μ L of concentrated H₂SO₄. Dissolved oxygen was determined in the field using a portable photometer (Oxygen 2 SAM and Vacu-vials, CHEMetrics, Virginia, USA) with a detection limit of 0.1 mg L⁻¹, and pH was measured in the field using a portable pH meter (pH tester 2,

- ⁵ Eutech Instruments, Singapore) with a detection limit of 0.1 pH . Groundwater samples for measuring dissolved N₂O were obtained inline by filling a 10 ml syringe connected to a peristaltic pump and injecting the sample into 20 ml evacuated glass vials containing 0.3 mL 80% ZnCl₂ for preserving dissolved N₂O (Blicher-Mathiesen and Hoffman, 1999). Samples were packed in ice in the field and refrigerated (4°C) in the laboratory.
- ¹⁰ Additional data for this study included monthly groundwater samples collected from 1997 to 1999 in the same monitoring and stilling wells at each site (Spear, 2003).

2.3 Chemical analysis

Samples for NO_3^- were analyzed utilizing UV- second derivative spectroscopy (Crumpton et al., 1992) with a detection limit of 0.1 mg L⁻¹. Chloride samples were analyzed

- ¹⁵ with an ion specific electrode (Orion 9617BNWP, Thermo Scientific, Massachusetts, USA) with a detection limit of 0.1 mg L⁻¹. Dissolved organic carbon samples were filtered through a 0.45 μ m filter and analyzed by persulfate oxidation on a carbon analyzer (Phoenix 8000, Tekmar-DohrmannTM, Ohio, USA) with a detection limit of 0.1 mg L⁻¹. Vials storing groundwater samples of dissolved N₂O were warmed to
- ²⁰ room temperature (21–22°C), shaken, and brought to atmospheric pressure with He. A gas chromatograph (Model GC17A, Shimadzu, Kyoto, Japan) equipped with a ⁶³Ni electron capture detector and a stainless steel column (0.3175 cm diameter×74.54 cm long) with Porapak Q (80–100 mesh) was used to analyze headspace gas concentrations (Parkin and Kaspar, 2006). Dissolved gas concentrations were determined using
- the Bunsen coefficient relationship (Tiedje, 1994) and estimated detection limit was $0.6 \,\mu g \,L^{-1}$ (Spear, 2003).

2.4 Nitrate and dissolved N₂ O flux

Cumulative annual flux of NO₃-N and dissolved N₂O-N in groundwater at the crop field edge of the buffers was estimated using NO₃-N and dissolved N₂O-N concentrations measured in the cool-season grass filter (R1, R9, R39) and the multi-species riparian

- ⁵ buffer (R16, R20, R24) once a month in 2006–2007. Cumulative annual flux of NO_3 -N and dissolved N₂O-N in groundwater at the creek edge of the buffers was estimated using NO₃-N and dissolved N₂O-N concentrations measured in the cool-season grass filter (R8, R12, R40) and the multi-species riparian buffer (R13, R17, R21) once a month in 2006–2007.
- Daily mass flux was estimated using average linear velocity, effective porosity, measured concentrations and cross sectional area of the aquifer adjacent to Bear Creek (Fetter, 1999) (Eq. 1). The cross sectional area was determined by creating a hypothetical rectangle (35 m wide×2 m height) representing the aquifer underlying the riparian buffers adjacent to Bear Creek. Monthly mass flux was estimated by multiplying the
- daily mass flux with days of the month (Eq. 2), and annual mass flux was sum of all monthly mass flux (Eq. 3).

$$F_{x, \text{day}i} = v_x \times n_e \times \text{Conc} \times \text{cross sectional area of aquifer}$$
 (1)

$$F_{x, \text{month } i} = F_{x, \text{day } i} \times \text{days of month } i$$
 (2)

$$F_{x, \text{ year}} = \sum_{i=1}^{12} F_{x, \text{ month } i}$$
(3)

20 where,

 $F_{x, dav i}$ is estimated daily mass flux (g d⁻¹ m⁻²) in month *i*,

 v_x is average linear velocity (m d⁻¹): cool-season grass filter 0.23 and multispecies riparian buffer 0.13 from Spear (2003), n_e is effective porosity (unitless): 0.15 from Spear (2003),

Conc is concentration, $g m^{-3}$ or $mg L^{-1}$,

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Cross sectional area of aquifer (m²) is hypothetical rectangle representing aquifer,

 $F_{x, \text{month } i}$ is estimated monthly mass flux (g month⁻¹ m⁻²) in month *i*,

 $F_{x, \text{ vear}}$ is estimated annual mass flux (g y⁻¹ m⁻²) in a year.

To estimate total flux of NO₃-N and dissolved N₂O-N from all cropped fields (both sides of the creek) within the Bear Creek watershed, the above procedure (Eqs. 1–3) was used with the monthly measured NO₃-N and dissolved N₂O-N concentrations in groundwater at the crop field edge of the buffers. The concentrations measured in the

cool-season grass filter (R1, R9, R39) and the multi-species riparian buffer (R16, R20, R24) represented the concentrations in the aquifers on each side of the creek (Eq. 1). Two hypothetical rectangles (56 473 m wide×2 m height) representing the aquifers on each side of the creek were applied (Eq. 1).

3 Determining $Frac_{LEACH-(H)}$, EF_{5g} , and ratios of dissolved N₂O flux to N inputs (EI-EF_{5q}) and dissolved N₂O flux to soil N₂O emission (N₂O_(D)-N/N₂O_{direct}-N)

In this study, the Frac_{LEACH-(H)} was determined by the ratio of N inputs to runoff and leaching N in all crop fields (corn fields: 3404.95 ha, soybeans fields: 3404.95 ha) within the Bear Creek watershed. Nitrogen inputs included the annual amount of synthetic fertilizer N applied to crop fields (FSN) and N inputs from crop residue (FCR) and they were determined in Kim et al. (2009). Estimated NO₃-N flux in groundwater discharged from all cropped fields was used as the amount of leaching N, and runoff N was estimated using the runoff rate (5.23 kg N ha⁻¹ y⁻¹) determined in Kim et al. (2009). In this study, the N₂O emission factor, EF_{5g} was determined by the mean of the ratio

of dissolved N_2O concentration to NO_3 -N concentration in groundwater discharged

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from the crop fields, the multi-species riparian buffer, and the cool-season grass filter in 2006–2007. A modified emission factor, $EI-EF_{5g}$, was determined based on the ratio between dissolved N₂O flux (N₂O_(D)-N) in groundwater discharged from all cropped fields and N inputs (FSN+FCR) in all crop fields within the watershed.

Soil N₂O emission (N₂O_{direct}-N) in all cropped fields within the watershed was estimated using the measured soil N₂O emission rate (corn field: 7.2 kg N ha⁻¹ yr⁻¹, soybeans field: 16.8 kg N ha⁻¹ yr⁻¹) in the sites in 2006–2007 (Kim et al., 2009) and the ratio of dissolved N₂O flux to soil N₂O emission (N₂O_(D)-N/N₂O_{direct}-N) was determined using the data.

10 3.1 Statistical analysis

The Shapiro-Wilk normality test was performed to determine the normal distribution of the data. A two-sample t-test was used to evaluate differences in concentrations of NO_3^- ; Cl⁻; pH; and dissolved N₂O, DO, and DOC in groundwater at the crop field edge of the buffers and groundwater at the creek edge of the buffers. When the standard ¹⁵ assumption of normality and equal variance were violated, the Mann-Whitney rank sum test was used. One-way ANOVA was used to evaluate the difference in groundwater tables and creek water stage. Differences were considered significant at the *P*<0.05 level. GLM was utilized to determine correlations between groundwater water quality parameters and dissolved N₂O. Statistical analyses were conducted by SAS ver

²⁰ 8.1 (SAS institute, 1999).

4 Results

4.1 Groundwater and creek elevations

In the cool-season grass filter, the groundwater elevation at the crop field edge of the buffer and the groundwater elevation at the creek edge of the buffer were significantly different (P<0.0001) from each other in both 1997–1999 and 2005–2008. The groundwater elevation at the crop field edge of the buffer (318.16±0.03 m a.s.l., n=69) was significantly higher than at the stream edge of buffer (317.43±0.02 m a.s.l., n=74) over the entire period (Tukey's Studentized Range Test), indicating general

- ⁵ groundwater flow from the crop fields to Bear Creek. However, the groundwater elevation at the creek edge of the buffer (317.43 ± 0.02 m a.s.l., n=74) and creek elevation (317.35 ± 0.04 m a.s.l., n=66) were not significantly different (Tukey's Studentized Range Test) during the entire period. In summer 1998, early spring 2006, and summer 2007, the groundwater elevation adjacent to the creek and the creek elevation
- were very similar. In December 2007, the groundwater elevation adjacent to creek was lower than the creek elevation, indicating the possibility for movement of stream water into the riparian aquifer.

Within the multi-species riparian buffer, the groundwater elevation within all wells and Bear Creek elevation were significantly different in both the 1997–1998 and 2005–2008

- periods (*P*<0.0001), again indicating general groundwater flow from the crop fields to Bear Creek under the buffer. In contrast the cool-season grass filter, the groundwater elevation at the creek edge of the buffer (317.60±0.03 m a.s.l., *n*=73) was significantly higher than the creek elevation (317.34±0.04 m a.s.l., *n*=58) (Tukey's Studentized Range Test) during the entire period, indicating that there was no movement of the creek water into the riparian aquifer.
 - 4.2 Nitrate concentration, flux, and reduction rate

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In the cool-season grass filter, NO₃-N concentrations in groundwater adjacent to the crop field showed a repeated seasonal trend with the highest concentrations in winter and lowest in summer (Fig. 2). However, NO₃-N concentrations in groundwater adjacent to the creek did not show any seasonal trend (Fig. 2). Average NO₃-N concentration was 9.5 mg L^{-1} in groundwater wells adjacent to crop fields and 4.9 mg L^{-1} in wells adjacent to creek, during 1997–1999 (Fig. 3), and 9 and 3.3 mg L^{-1} , respectively, during 2005–2008 (Fig. 4). In this cool-season grass filter site, NO₃-N concentrations

in groundwater adjacent to crop fields were significantly higher than those adjacent to the creek during both 1997–1999 (Mann-Whitney rank sum test P<0.0001) and 2005–2008 (Mann-Whitney rank sum test P<0.0001). The average NO₃-N concentration in groundwater within the cool-season grass filter decreased by 48% in 1997–1999 and

- 59% in 2005–2008 when comparing wells nearest the creek with those nearest the crop field. In January 2006–December 2007, NO₃-N flux in groundwater from the crop field to the cool-season grass filter was 14.2 kg N and NO₃-N flux from the cool-season grass filter to the creek was 5.1 kg N (Fig. 4). This indicates that 9.1 kg N was removed from the groundwater as it flowed from the crop field through the cool-season
- ¹⁰ grass filter. This equates to a removal rate of 130 kg groundwater NO₃-N ha⁻¹ in the cool-season grass filter (9.1 kg N loss in length 35 m×width 20 m) during the period of January 2006 to December 2007.

In the multi-species riparian buffer, NO_3 -N concentrations in groundwater adjacent to the crop field showed a repeated seasonal trend with the highest concentrations in winter and lowest in summer (Fig. 2). However, NO_3 -N concentration in groundwater adjacent but the highest concentration in groundwater adjacent but the highest concentration in groundwater adjacent but the highest concentration is groundwater adjacent but the highest concent but the highest

- ter adjacent to the creek did not show any seasonal trend (Fig. 2). Average NO₃-N concentrations were 4.9 mg L^{-1} in groundwater wells adjacent to the crop field and 5.0 mg L^{-1} in wells adjacent to the creek, respectively, during 1997–1999 (Fig. 3), and $4.0 \text{ and } 2.0 \text{ mg L}^{-1}$, respectively, during 2005–2008 (Fig. 4). The differences in concentration of the creek is the creek of the creek in the creek is the creek
- trations during 1997–1999 were not significant (Mann-Whitney rank sum test P=0.91) (Fig. 3) but, within this same buffer, average NO₃-N concentration in groundwater decreased by 49.5% in 2005–2008 across the riparian buffer (Mann-Whitney rank sum test P<0.0001) (Fig. 4). In January 2006–December 2007, NO₃-N flux in groundwater from the crop field to the multi-species riparian buffer was 4.4 kg N and groundwater
- $_{25}$ NO₃-N flux from the multi-species riparian buffer to the creek was 2.1 kg N (Fig. 4). This indicates the NO₃-N flux was 2.3 kg N (52.2%) lower in groundwater nearest the creek compared to near the crop field edge and the resulting groundwater NO₃-N removal was 33.1 kg N ha⁻¹ (2.3 kg N loss in length 35 m×width 20 m) in the multi-species riparian buffer in January 2006–December 2007.

4.3 Chloride concentration and the ratio of nitrate to chloride

Average Cl⁻ concentrations in groundwater ranged between 13.2 and 13.4 mg L^{-1} within the cool-season grass filter during 1997–1999 and between 20.6 and 20.9 mg L⁻¹ within the multi-species riparian buffer during the same period (Fig. 3).

- ⁵ During 2005–2008, average Cl⁻ concentrations in groundwater ranged between 18.2 and 20.6 mg L⁻¹ within grass filters and between 18.2 and 20.8 mg L⁻¹ within the multispecies riparian buffer (Fig. 4). None of these differences in Cl⁻ concentrations were significant. In the cool-season grass filter, the average NO₃⁻/Cl⁻ ratio within groundwater adjacent to crop fields was significantly higher than adjacent to the creek in both
- ¹⁰ 1997–1999 (Mann-Whitney rank sum test P < 0.0001) and 2005–2008 (Mann-Whitney rank sum test P < 0.0001) (Figs. 3, 4 and 5). Within groundwater under the multispecies riparian buffer, there was no significant difference in the average NO₃⁻/Cl⁻ ratio of groundwater adjacent to crop fields and adjacent to the creek in 1997–1999 (Mann-Whitney rank sum test P = 0.41) (Figs. 3 and 5). However, within this same system,
- the average NO_3^-/Cl^- ratio within groundwater adjacent to crop fields was significantly higher than that adjacent to the creek in 2005–2008 (Mann-Whitney rank sum test P < 0.0001) (Figs. 4 and 5).

4.4 Dissolved N₂O

Dissolved N₂O-N concentration in groundwater under both riparian buffers showed a repeated seasonal trend, with the concentrations highest in winter and lowest in summer (Fig. 6). Average dissolved N₂O-N concentrations in groundwater ranged between 6.8 and 7.8 μ g L⁻¹ within the cool-season grass filter during 1997–1999 and between 6.0 and 6.1 μ g L⁻¹ within the multi-species riparian buffer during the same period (Fig. 3). During 2005–2008, average dissolved N₂O-N concentrations in ground-

water ranged between 11.6 and $14.4 \,\mu g L^{-1}$ within the cool-season grass filters and between 9.0 and 9.1 $\mu g L^{-1}$ within the multi-species riparian buffer (Fig. 4). Within groundwater under the cool-season grass filter, there was no significant difference in

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dissolved N₂O-N concentration in wells adjacent to the crop fields and adjacent to the creek during both 1997–1999 (Mann-Whitney rank sum test P=0.49) and 2005–2008 (Mann-Whitney rank sum test P=0.29). In January 2006–December 2007, dissolved N₂O-N flux was 19.7 g N in groundwater adjacent to the crop field and 20.0 g N in the cool-season grass filter near the creek (Fig. 4).

This pattern was repeated in groundwater under the multi-species riparian buffer, with no significant difference in dissolved N₂O-N concentrations in groundwater adjacent to crop fields and the creek during either 1997–1999 (Mann-Whitney rank sum test P=0.96) or 2005–2008 (Mann-Whitney rank sum test P=0.93). In January 2006–

December 2007, dissolved N₂O-N flux was 8.3 g N in groundwater adjacent to the crop field and 7.7 g N in the multi-species riparian buffer near the creek (Fig. 4).

4.5 Dissolved oxygen, dissolved organic carbon, pH, and water temperature

Average dissolved oxygen concentration in groundwater under the grass filter adjacent to crop fields $(5.0\pm0.3 \text{ mg L}^{-1})$ was significantly higher than adjacent to the creek

¹⁵ (2.6±0.3 mg L⁻¹) in 1997–1999 (two sample t-test P < 0.0001) (Figs. 3 and 4). However, there was no significant difference within this same system in DO concentration in groundwater adjacent to crop fields and adjacent to the creek (2.7–3.3 mg L⁻¹) in 2005– 2008 (two sample t-test P=0.34). Within the multi-species riparian buffer, there was no significant difference in DO concentration in groundwater adjacent to crop fields and

adjacent to the creek either 1997–1999 (2.8–3.4 mg L⁻¹) (two sample t-test *P*=0.29) or 2005–2008 (2.7–3.3 mg L⁻¹) (two sample t-test *P*=0.24). In both buffer vegetation types, the average dissolved organic carbon (DOC) concentration (0.6–1.1 mg L⁻¹) within the groundwater was not significantly different adjacent

to crop fields and adjacent to the creek during either 1997–1999 (two sample t-test P>0.1) or 2005–2008 (two sample t-test P>0.1) (Figs. 3 and 4). Similarly, there was no significant difference in groundwater temperature under either buffer type within wells adjacent to crop fields and adjacent to the creek during either 1997–1999 (two sample t-test P>0.1) or 2005–2008 (two sample t-test P>0.1).

Within the grass filter, pH in groundwater adjacent to crop fields (7.5) was significantly higher than adjacent to the creek in 1997–1999 (7.3) (two sample t-test P=0.03); however, there was no significant differences in 2005–2008 (7.4–7.5) (two sample t-test P=0.30). Within the multi-species riparian buffer, there was no significant difference

⁵ in pH in groundwater adjacent to crop fields and adjacent to the creek during either 1997–1999 (7.5) (two sample t-test P=0.70) or 2005–2008 (7.4) (two sample t-test P=0.62).

4.6 Relation between dissolved N₂O concentrations and water characteristics

There was a significant negative relationship between water temperature and dissolved N₂O concentration in groundwater adjacent to both crop fields and the creek within the grass filter (Pearson coefficient r=-0.31, P=0.003) and the multi-species riparian buffer (Pearson coefficient r=-0.39, P=0.006, in respect). There was also a significant relationship between DO and dissolved N₂O concentration in groundwater adjacent to the creek within the multi-species riparian buffer (Pearson coefficient r=0.33, P=0.006, in respect).

- P=0.048). Nitrate concentration and pH did not show a significant correlation with dissolved N₂O concentrations in either the groundwater adjacent to crop fields or the creek (all *P*>0.05) (Fig. 7a and b).
 - 4.7 Frac_{LEACH-(*H*)}, EF_{5g}, EI-EF_{5g}, and N₂O_(*D*)-N/N₂O_{direct}-N in the crop fields and riparian buffers
- Estimated N leaching (TL) (estimated NO₃-N flux) from all crop fields within the Bear Creek watershed was 30 084 kg N in 2006–2007 (Table 1). Run-off N (TR) from all crop fields within the Bear Creek watershed was 71 231 kg N (North of Bear Creek: 35 615 kg N, South of Bear Creek: 35 615 kg N) in 2006–2007 (Table 2). Annual synthetic fertilizer N applications to crop fields (FSN) were estimated at 908 440 kg N and
- N inputs from crop residue (FCR) were estimated at 1 186 965 kg N in 2006–2007 (Table 2). Using these data, the ratio of runoff and leaching N (TR+TL, 101 315 kg N) to

N inputs ($F_{SN}+F_{CR}$, 2095406.2 kg N), Frac_{LEACH-(H)}, was determined to be 0.05 (Table 2).

EF5g (mean ratio of dissolved N₂O concentration to NO₃-N concentration) in groundwater discharged from crop fields in this study (n=99) was 0.0022 (95% C.I. 0.0013–

0.0031) (Fig. 7A). EF_{5g} in groundwater discharged from riparian buffers in this study (n=101) was 0.0041 (95% C.I. 0.0028–0.0054) (Fig. 7B).

Estimated dissolved N₂O flux (N₂O_(D)-N) in all cropped fields within the Bear Creek watershed was 45.2 kg N in 2006–2007 (Table 1). Nitrogen inputs ($F_{SN}+F_{CR}$) were 2 095 406 kg N in all cropped fields within the Bear Creek watershed in 2006–2007

- ¹⁰ (Table 2). Using these data, the EI-EF_{5g} (ratio of dissolved N₂O flux to N inputs, N₂O_(D)-N/ $F_{SN}+F_{CR}$) was determined to be 0.00002 in all cropped fields within the Bear Creek watershed (Table 2). EI-EF_{5g} within riparian buffers in this study was zero since there was no significant change in dissolved N₂O flux in groundwater under either riparian buffer (Fig. 4).
- ¹⁵ The estimated soil N₂O emission (N₂O_{direct}-N) in all cropped fields within the Bear Creek watershed was 163 437 kg N (corn fields: 49 031 kg N, soybeans fields: 114 406 kg N) in 2006–2007 (Table 2). Using estimated dissolved N₂O flux (N₂O_(D)-N) and soil N₂O emission (N₂O_{direct}-N), N₂O_(D)-N/N₂O_{direct}-N (ratio of dissolved N₂O flux to soil N₂O emission) was determined to be 0.0003 (Table 2). N₂O_(D)-N/N₂O_{direct}-N in
- 20 groundwater discharged from riparian buffers to soil N₂O emission in riparian buffers was zero since there was no significant change in dissolved N₂O within groundwater under either riparian buffer (Fig. 4).

5 Discussion

5.1 Transport and fate of nitrate

²⁵ Nitrate concentration in groundwater was significantly decreased under the coolseason grass filter in both 1997–1999 and 2005–2008 and under the multi-species

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riparian buffer in 2005–2008. Processes that may decrease NO₃⁻ concentration in groundwater include dilution of groundwater, uptake by vegetation, and denitrification. Andress (1999), using an isotopic method, found denitrification occurring at this coolseason grass filter site. Our data showed a decrease in the NO₃⁻/Cl⁻ ratio in both sites,

- ⁵ with a significant decrease in NO₃⁻ concentration and an insignificant change in the Cl⁻ concentration. These results suggest that dilution from a converging or diverging flow path was not a major factor contributing to the decrease in groundwater NO₃⁻ concentration (e.g. Vidon and Hill, 2004; Davis et al., 2007). Uptake of NO₃⁻ by vegetation was not investigated in this study but is known to occur in riparian buffers (e.g. Clément et
- al., 2003; Dhondt et al., 2003; Hefting et al., 2005).
 In our studies, there was no significant NO₃⁻ decrease observed during 1997–1999 under the multi-species riparian buffer. Andress (1999) and Simpkins et al. (2002) demonstrated that this site, then a 7-year-old buffer, has a sand aquifer which might decrease groundwater residence time and reduce the potential for N loss, allowing
- ¹⁵ transport of NO₃⁻ to the creek. Several studies have documented the importance of hydrogeologic setting, specifically the direction of groundwater flow and the position of the water table in thin sand aquifers underlying the buffers, in determining buffer N removal efficiency (Puckett, 2004). To the point of this study however, the multi-species riparian buffer has been shown to be a site of significant groundwater NO₃⁻ removal as
- the groundwater moves from cropped fields to the creek. The age of the buffer could also be a potential contributing factor for the difference found in N removal efficiency.
 - 5.2 Transport and fate of dissolved N₂O

The dissolved N_2O concentration in groundwater was not significantly changed during travel under either the cool-season grass filter or the multi-species riparian buffer in 1997–1999 or 2005–2008. The dissolved N_2O concentrations in both sites (6–

²⁵ in 1997–1999 or 2005–2008. The dissolved N₂O concentrations in both sites (6– $14 \mu g L^{-1}$) were similar to those (0– $6.3 \mu g N L^{-1}$) reported by Davidson and Firestone (1988), Davidson and Swank (1990), Papen and Butterbach-Bahl (1999),

Blicher-Mathiesen and Hoffmann (1999), Höll et al. (2005), and Davis et al. (2007) and less than those reported by Weller et al. (1994) (17.2 μ g N L⁻¹) and Well et al. (2001) (10.2–53.2 μ g N L⁻¹). Weller et al. (1994) estimated 0.35 kg N ha⁻¹ of annual N₂O loss in soil emission and 0.04 kg N ha⁻¹ in groundwater (<1% of the intercepted N) and

- $_{5}$ concluded that N₂O production in the riparian buffer is neither an important fate of N removed from cropland discharges nor an important source of atmospheric N₂O pollution. Blicher-Mathiesen and Hoffman (1999) reported that denitrification in a riparian soil can act as a sink for dissolved N₂O in the inflowing groundwater as well as for N₂O produced in the riparian sediment. Davis et al. (2007) reported that both dissolved
- N₂O and NO₃⁻ were significantly lower in the riparian area than in the adjacent cropping system. Clough et al. (2007) reported significant consumption of ¹⁵N₂O injected into groundwater in an upland-marsh transition zone of a salt marsh and a forested alluvial riparian zone. These studies commonly concluded that dissolved N₂O is decreased in riparian areas.
- Our results regarding NO_3^- decrease without increasing dissolved N_2O in the coolseason grass filter or the multi-species riparian buffer can be explained three different ways. First, it may be that denitrification completed the reduction of NO_3^- to N_2 without producing N_2O (Blicher-Mathiesen and Hoffman, 1999). In the groundwater, very low concentrations of DO (<2 ppm) were often observed and the anaerobic microsites
- ²⁰ might support completion of denitrification (e.g. Desimone and Howes, 1996; Spalding and Parrott, 1994; Starr and Gillham, 1993). This possibility is supported by the significant relationship we found between DO and dissolved N₂O. Second, produced N₂O in groundwater can be released into unsaturated soil above the groundwater table. In this study, the estimated NO₃⁻ losses in groundwater in the cool-season grass
- filter and multi-species riparian buffer was $130.0 \text{ kg N ha}^{-1}$ and $33.1 \text{ kg N ha}^{-1}$, respectively, and N₂O emission measured on the soil surface was $5.8 \text{ kg N}_2\text{O-N ha}^{-1}$ in the sites through 2006 to 2007 (Kim et al., 2009). The ratio of N₂O emission measured on the soil surface to NO₃⁻¹ loss in the groundwater in the cool-season grass filter and multi-species riparian buffer was 0.04 and 0.17, respectively. Since the N₂O emis-

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sion measured on the soil surface includes the N₂O produced in the unsaturated soil layer, the results suggest that release of N₂O produced in groundwater into unsaturated soil above the groundwater table to be an insignificant pathway of NO₃⁻ losses. This is consistent with the finding of Deurer et al. (2008) who estimated that upward

- $_5$ fluxes from the exchange zone into the unsaturated zone ranged between 0.0009 to $0.3\,kg\,N_2O\,ha^{-1}\,y^{-1}$ and the yearly downward fluxes into the exchange zone had about the same order of magnitude. Third, vegetation and microbial communities within the riparian buffers can assimilate and immobilize NO_3^- resulting in NO_3^- decrease without increasing dissolved N_2O in the groundwater. Since this study did not investigate
- NO₃⁻ losses by these pathways, we cannot exclude the possibility. Overall, it is suggested that the cool-season grass filter or the multi-species riparian buffer should be considered insignificant sources of dissolved N₂O flux.
 - 5.3 Determined $Frac_{LEACH-(H)}$, EF_{5g} , EI- EF_{5g} and $N_2O_{(D)}$ - N/N_2O_{direct} -N and their implication
- ¹⁵ In this study, $Frac_{LEACH-(H)}$, N lost to leaching and runoff, was determined to be 0.05 a value is 6-fold lower than the current IPCC's default value (0.3). Thoms et al. (2005) suggested a value of 0.07 (0.03–0.1) as an appropriate $Frac_{LEACH-(H)}$ for New Zealand conditions. Our $Frac_{LEACH-(H)}$ is similar to the value suggested for New Zealand (Thomas et al., 2005).
- Our results determine EF_{5g} (emission factor for N₂O emissions from N leaching and runoff) of groundwater leached from the crop fields at 0.0022 (95% C.I. 0.0013– 0.0031). The Intergovernmental Panel for Climate Change (2006) reported that the previously used EF_{5g} for groundwater leached from crop fields (0.015) (IPCC, 1997) was too high and they modified EF_{5g} to 0.0025 based on several studies (Hiscock et
- ²⁵ al., 2003; Reay et al., 2004; Sawamoto et al., 2005). Our EF_{5g} is similar to the new IPCC EF_{5g}. However, since 1) the N₂O emission factor is typically defined by the ratio of N₂O emission and N input (IPCC, 2006; Weymann et al., 2008), 2) N₂O-N concentration in groundwater does not necessarily reflect actual dissolved N₂O emission (Höll

et al., 2005), and 3) insignificant correlation between NO₃-N and dissolved N₂O-N concentrations was observed in this study (Fig. 7), it is suggested that EI-EF_{5g} is suitable for the emission factor of dissolved N₂O emission in the sites. The EI-EF_{5g} (0.00002) in the crop fields is 35-fold less than the value of Frac_{LEACH-(H)} (0.3)×EF_{5g} (0.0025)

and this indicates the current IPCC methodology using $Frac_{LEACH-(H)}$ and EF_{5g} substantially overestimates dissolved N₂O flux at this site. It is suggested that $EI-EF_{5g}$ be determined in various regions and environments and these values be evaluated as improvements to current IPCC methodology.

In this study, $N_2O_{(D)}$ -N/ N_2O_{direct} -N (the ratio of dissolved N_2O flux to soil N_2O emission) in the crop fields was 0.0003 indicating that the mass of dissolved N_2O leaving the crop field was negligible in comparison to soil N_2O emission. This is consistent with the findings of Davidson and Swank (1990), Ueda et al. (1991), Harrison and Matson (2003), Reay et al. (2004), Höll et al. (2005), and Well et al. (2005) who suggested that indirect N_2O emission is an insignificant pathway in the N cycle. However, in this

study, it was recognized that a monthly sampling interval of dissolved N₂O measurements may not be frequent enough to observe increases of dissolved N₂O-N concentration and peaks of dissolved N₂O emission affected by dry-wet events (Höll et al., 2005) and that estimating NO₃-N and dissolved N₂O-N flux of the whole watershed using results from measurements in two sites embodies significant uncertainties.

20 6 Conclusions

Monitoring of groundwater under a cool-season grass filter, a multi-species riparian buffer, and adjacent crop fields during 1997–1999 and 2005–2008 indicated that the concentration of dissolved N₂O was not significantly changed, even when the concentration of groundwater NO_3^- were decreased by 49.5% under the multi-species ripar-

ian buffers and 58.8% under the cool-season grass filter, over the same time periods. The decrease in the NO₃⁻/Cl⁻ ratio in groundwater under riparian buffers with significant NO₃⁻ concentration decrease provides evidence that dilution from a converging or diverging flow path was not a major factor contributing to the decreased NO₃⁻ concentration in groundwater. Our results indicated that determined Frac_{LEACH-(H)} (0.05) is 6-fold less than the current IPCC's default value (0.3) and the N₂O emission factor (EF_{5g}) based on the ratio between dissolved N₂O-N and NO₃-N concentrations was

- ⁵ not suitable in the site. The modified emission factor (EI-EF_{5g}) based on the ratio between dissolved N₂O flux and N input (0.00002) was determined in the crop fields and indicates the current IPCC methodology overestimates dissolved N₂O flux in the site. A low ratio between dissolved N₂O flux and soil N₂O emission (0.0003) was observed in the crop fields. Based on these results, we suggest that the riparian buffers established
- adjacent to crop fields to decrease NO_3^- did not increase dissolved N_2O in groundwater and dissolved N_2O flux from the crop fields was negligible in comparison to soil N_2O emission.

Acknowledgements. The authors wish to thank Leigh Ann Long, Otis Smith and numerous students for technical assistants in the field and lab. We are also grateful to Dr. William Simpkins
 for providing technical supports and valuable comments on this project. Financial support was provided US EPA, USDA NRI, USDA SARE Graduate Student Grant Program, Leopold Center for Sustainable Agriculture, University of Iowa Center for Global and Regional Environmental Research, and University of Missouri Center for Agroforestry.

References

- Andress, R. J.: Fate and transport of nitrate in groundwater within a riparian buffer in the Bear Creek watershed, M.S. thesis, Iowa State University, Ames, IA, USA, 157 pp., 1999.
 Blicher-Mathiesen, G. and Hoffmann, C. C.: Denitrification as a sink for dissolved nitrous oxide in a freshwater riparian fen, J. Environ. Qual., 28, 257–262, 1999.
- Clément, J. C., Holmes, R. M., Peterson, B. J., and Pinay, G.: Isotopic investigation of denitrification in a riparian ecosystem in western france, J. Appl. Ecol., 40, 1035–1048, 2003.
- Clough, T. J., Addy, K., Kellogg, D. Q., Nowicki, B. L., Gold, A. J., and Groffman, P. M.: Dynamics of nitrous oxide in groundwater at the aquatic-terrestrial interface, Global Change Biol., 13, 1528–1537, 2007.

671

- Crumpton, W. G., Isenhart, T. M., and Mitchell, P. D.: Nitrate and organic n analyses with 2nd-derivative spectroscopy, Limnol. Oceanogr., 37, 907–913, 1992.
- Davidson, E. A. and Firestone, M. K.: Measurement of nitrous oxide dissolved in soil solution, Soil Sci. Soc. Am. J., 52, 1201–1203, 1988.
- Davidson, E. A. and Swank, W. T.: Nitrous oxide dissolved in soil solution: An insignificant pathway of nitrogen loss from a southeastern hardwood forest, Wat. Resour. Res., 26, 1687– 1690, 1990.

Davis, J. H., Griffith, S. M., Horwath, W. R., Steiner, J. J., and Myrold, D. D.: Mitigation of shallow groundwater nitrate in a poorly drained riparian area and adjacent cropland, J. Environ. Qual., 36, 628–637, 2007.

- Desimone, L. A. and Howes, B. L.: Denitrification and nitrogen transport in a coastal aquifer receiving wastewater discharge, Environ. Sci. Technol., 30, 1152–1162, 1996.
- Deurer, M., von der Heide, C., Bottcher, J., Duijnisveld, W. H. M., Weymann, D., and Well, R.: The dynamics of N₂O near the groundwater table and the transfer of N₂O into the unsaturated zone: A case study from a sandy aquifer in Germany, Catena, 72, 362–373, 2008.
- Dewitt, T. A.: Soil survey of Story County, Iowa, USDA Soil Conserv. Serv., Washington, D. C., USA, 149 pp., 1984.
- Dhondt, K., Boeckx, P., Van Cleemput, O., and Hofman, G.: Quantifying nitrate retention processes in a riparian buffer zone using the natural abundance of N-15 in NO₃, Rapid Commun. Mass Spectrom., 17, 2597–2604, 2003.
- Fetter, C. W.: Contaminant Hydrology, 2nd ed., Prentice Hall, New Jersey, USA, 500 pp., 1999.
 Forster, P., Ramaswamy, V., Artaxo, P., Berntsen, T., Betts, R., Fahey, D. W., Haywood, J., Lean, J., Lowe, D. C., Myhre, G., Nganga, J., Prinn, R., Raga, G., Schulz, M., and Van Dorland, R.: Changes in Atmospheric Constituents and in Radiative Forcing, in: Climate Change 2007:
- The Physical Science Basis, in: Contribution of Working Group I to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change, edited by: Solomon, S., Qin, D., Manning, M., Chen, Z., Marquis, M., Averyt, K. B., Tignor, M., and Miller, H. L., Cambridge University Press, Cambridge, United Kingdom and New York, NY, USA, 2007.
- Groffman, P. M. and Hanson, G. C.: Wetland denitrification: Influence of site quality and relationships with wetland delineation protocols, Soil Sci. Soc. Am. J., 61, 323–329, 1997.
- Groffman, P. M., Gold, A. J., and Addy, K.: Nitrous oxide production in riparian zones and its importance to national emission inventories, Chemosphere-Global Change Science, 2, 291–299, 2000.

- Groffman, P. M., Gold, A. J., and Jacinthe, P. A.: Nitrous oxide production in riparian zones and groundwater, Nutr. Cycling Agroecosyst., 52, 179–186, 1998.
- Groffman, P. M.: Denitrification in freshwater wetlands, Curr. Topics Wetland Biogeochem., 1, 15–35, 1994.
- ⁵ Harrison, J. and Matson, P.: Patterns and controls of nitrous oxide emissions from waters draining a subtropical agricultural valley, Global Biogeochem. Cy., 17, 1080, doi:10.1029/2002gb001991, 2003.
 - Hefting, M. M., Clement, J. C., Bienkowski, P., Dowrick, D., Guenat, C., Butturini, A., Topa, S., Pinay, G., and Verhoeven, J. T. A.: The role of vegetation and litter in the nitrogen dynamics
- of riparian buffer zones in Europe, Ecol. Eng., 24, 465–482, 2005.
 Hill, A. R. and Cardaci, M.: Denitrification and organic carbon availability in riparian wetland soils and subsurface sediments, Soil Sci. Soc. Am. J., 68, 320–325, 2004.

Hill, A. R.: Nitrate removal in stream riparian zones, J. Environ. Qual., 25, 743-755, 1996.

Hill, A. R., Devito, K. J., Campagnolo, S., and Sanmugadas, K.: Subsurface denitrification in

- a forest riparian zone: Interactions between hydrology and supplies of nitrate and organic carbon, Biogeochemistry, 51, 93–223, 2000.
 - Hiscock, K. M., Bateman, A. S., Muehlherr, I. H., Fukada, T., and Dennis, P. F.: Indirect emissions of nitrous oxide from regional aquifers in the United Kingdom, Environ. Sci. Technol., 37, 3507–3512, 2003.
- Höll, B. S., Jungkunst, H. F., Fiedler, S., and Stahr, K.: Indirect nitrous oxide emission from a nitrogen saturated spruce forest and general accuracy of the IPCC methodology, Atmos. Environ., 39, 5959–5970, 2005.
 - Intergovernmental Panel on Climate Change: 2006 IPCC Guidelines for National Greenhouse Gas Inventories, Prepared by the National Greenhouse Gas Inventories Programme, edited
- by: Eggleston, H. S., Buendia, L., Miwa, K., Ngara, T., and Tanabe, K., IGES, Japan, 2006. Intergovernmental Panel on Climate Change: Revised 1996 IPCC Guidelines for National Greenhouse Gas Inventories, Reference Manual, Vol. 3, IPCC WG I Technical Support Unit, Bracknell, UK, 1997.
 - Kim, D.-G., Isenhart, T. M., Parkin, T. B., Schultz, R. C., Loynachan, T. E., and Raich, J. W.: Nitrous oxide emissions from riparian forest buffers, warm-season and cool-season grass
 - filters, and crop fields, Biogeosciences Discuss., 6, 607-650, 2009,

http://www.biogeosciences-discuss.net/6/607/2009/.

20

25

Lee, K. H., Isenhart, T. M., and Schultz, R. C.: Sediment and nutrient removal in an established

673

multi-species riparian buffer, J. Soil Water Conserv., 58, 1-8, 2003.

- Mosier, A., Kroeze, C., Nevison, C., Oenema, O., Seitzinger, S., and van Cleemput, O.: Closing the global N₂O budget: Nitrous oxide emissions through the agricultural nitrogen cycle OECD/IPCC/IEA phase II development of IPCC guidelines for national greenhouse gas inventory methodology, Nutr. Cycling Agroecosyst., 52, 225–248, 1998.
- Nevison, C.: Review of the IPCC methodology for estimating nitrous oxide emissions associated with agricultural leaching and runoff, Chemosphere-Global Change Science, 2, 493– 500, 2000.
- Papen, H. and Butterbach-Bahl, K.: A 3-year continuous record of nitrogen trace gas fluxes from untreated and limed soil of a N-saturated spruce and beech forest ecosystem in Ger-
 - many 1. N₂O emissions, J. Geophys. Res., 104, 18487–18504, 1999.
 Parkin, T. B. and Kaspar, T. C.: Nitrous oxide emissions from corn-soybean systems in the Midwest, J. Environ. Qual., 35, 1496–1506, 2006.
- Puckett, L. J.: Hydrogeologic controls on the transport and fate of nitrate in ground water be-
- neath riparian buffer zones: Results from thirteen studies across the United States, Water Sci. Technol., 49, 47–53, 2004.
 - Reay, D. S., Edwards, A. C., and Smith, K. A.: Determinants of nitrous oxide emission from agricultural drainage waters, Water, Air and Soil Pollution: Focus, 4, 107–115, 2004.
- Reay, D. S., Smith, K. A., and Edwards, A. C.: Nitrous oxide in agricultural drainage waters following field fertilisation, Water, Air and Soil Pollution: Focus, 4, 437–451, 2004.
- SAS Institute Inc.: SAS/STAT User's Guide, Version 8, SAS Inst., Cary, NC, 3884 pp., USA, 1999.

Sawamoto, T., Nakajima, Y., Kasuya, M., Tsuruta, H., and Yagi, K.: Evaluation of emission factors for indirect N₂O emission due to nitrogen leaching in agro-ecosystems, Geophys. Res. Lett., 32, L03403, doi:03410.01029/02004GL021625, 2005.

- Schultz, R. C., Colletti, J. P., Isenhart, T. M., Simpkins, W. W., Mize, C. W., and Thompson, M. L.: Design and placement of a multispecies riparian buffer strip system, Agroforestry Systems, 29, 201–226, 1995.
- Schultz, R. C., Isenhart, T. M., Simpkins, W. W., and Colletti, J. P.: Riparian forest buffers in agroecosystems-lessons learned from the bear creek watershed, central Iowa, USA, Agroforestry Systems, 61, 35–50, 2004.
 - Seitzinger, S. P. and Kroeze, C.: Global distribution of nitrous oxide production and N inputs in freshwater and coastal marine ecosystems, Glob. Biogeochem. Cy., 12, 93–113, 1998.

- Simpkins, W. W., Wineland, T. R., Andress, R. J., Johnston, D. A., Caron, G. C., Isenhart, T. M., and Schultz, R. C.: Hydrogeological constraints on riparian buffers for reduction of diffuse pollution: Examples from the Bear Creek watershed in Iowa, USA, Water Sci. Technol., 45, 61–68, 2002.
- Spalding, R. F. and Parrott, J. D.: Shallow groundwater denitrification, Sci. Total Environ., 141, 17–25, 1994.
 - Spear, B. A.: Fate and transport of nitrate in groundwater through three multi-species riparian buffers along Bear Creek in central Iowa, M.S. thesis, Iowa State University, Ames, IA, USA, 172 pp., 2003.
- ¹⁰ Starr, R. C. and Gillham, R. W.: Denitrification and organic carbon availability in two aquifers, Ground Water, 31, 934–947, 1993.
 - Thomas, S. M., Ledgard, S. F., and Francis, G. S.: Improving estimates of nitrate leaching for quantifying New Zealand's indirect nitrous oxide emissions, Nutr. Cycling Agroecosyst., 73, 213–226, 2005.
- ¹⁵ Tiedje, J. M.: Denitrifiers, in: Methods of Soil Analysis, Part 2. Microbiological and Biochemical Properties, Soil Sci. Soc. Amer., Madison, Wisconsin, USA, 245–267, 1994.

Ueda, S., Ogura, N., and Wada, E.: Nitrogen stable isotope ratio of groundwater N₂O, Geophys. Res. Lett., 18, 1449-1452, 1991.

Vidon, P. and Hill, A. R.: Denitrification and patterns of electron donors and acceptors in eight riparian zones with contrasting hydrogeology, Biogeochemistry, 71, 259–283, 2004.

Wang, W. C., Yung, Y. L., Lacis, A. A., Mo, T., and Hansen, J. E.: Greenhouse effects due to man-made perturbations of trace gases, Science, 194, 685–690, 1976.

- Watts, S. H. and Seitzinger, S. P.: Denitrification rates in organic and mineral soils from riparian sites: A comparison of N₂ flux and acetylene inhibition methods, Soil Biol. Biochem., 32, 1383–1392, 2000.
- Well, R., Augustin, J., Davis, J., Griffith, S. M., Meyer, K., and Myrold, D. D.: Production and transport of denitrification gases in shallow ground water, Nutr. Cycling Agroecosyst., 60, 65–75, 2001.
- Well, R., Flessa, H., J aradat, F., Toyoda, S., and Yoshida, N.: Measurement
 of isotopomer signatures of N₂O in groundwater, J. Geophys. Res, 110, G02006, doi:02010.01029/02005JG000044, 2005.
 - Weller, D. E., Correl, D. L., and Jordan, T. E.: Denitrification in riparian forests receiving agricultural discharges, in: Global Wetlands: Old World and New, edited by: Mitsch, W. J., Elsevier,

675

Amsterdam, The Netherlands, 117–131, 1994.

- Weymann, D., Well, R., Flessa, H., von der Heide, C., Deurer, M., Meyer, K., Konrad, C., and Walther, W.: Groundwater N₂O emission factors of nitrate-contaminated aquifers as derived from denitrification progress and N2O accumulation, Biogeosciences, 5, 1215–1226, 2008, http://www.biogeosciences.pdf/5/1215/2008/
- 5 http://www.biogeosciences.net/5/1215/2008/.

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Table 1. Summarizing the procedures (Eqs. 1, 2, and 3) used to estimate total NO₃-N and dissolved N₂O-N flux in groundwater discharged from whole crop fields (North of Bear Creek and South of Bear Creek) in the Bear Creek watershed to Bear Creek in 2006–2007. Conc. is concentration, V_x is average linear velocity (North of Bear Creek: 0.23 and South of Bear Creek: 0.13 from Spear, 2003), n_e is effective porosity (0.15 from Spear, 2003), and $F_{x, day}$ and $F_{x, month}$ are estimated daily and monthly mass flux, respectively.

	Date	Conc.	V _x	ne	$F_{x, day}$	Aquifer	days	$F_{x, \text{month}}$	Total flux
	Date	mg L^{-1}	m day ⁻¹	no unit	g N day $^{-1}$ m $^{-2}$	m²		kg N month ⁻¹	kg N
NO ₃ -N, North of Bear Creek	Jan 2006 Dec 2007	11.7 6.7	0.23 0.23	0.15 0.15	0.4037 0.2312	112946 112946	31 31	1413.3 809.3	
	2006–2007								22 957.9
	Jan 2006 	3.5 	0.13	0.15	0.0689 	112946 	31 	241.2 	
NO ₃ -N, South of Bear Creek	Dec 2007 2006–2007	3.4	0.13	0.15	0.0657	112946	31	229.9	7126.1
NO3-N, Whole crop fields	2006-2007								30 084.0
N ₂ O-N, North of Bear Creek	Jan 2006 Dec 2007 2006–2007	0.0039 0.0179	0.23 0.23	0.15 0.15	0.000134 0.000618	112946 112946	31 31	0.5 2.2	31.8
N ₂ O-N, South of Bear Creek	Jan 2006 Dec 2007 2006–2007	0.0073 0.0106	0.13 0.13	0.15 0.15	0.000143 0.000206	112946 112946	31 31	0.5 0.7	13.4
N ₂ O-N, Whole crop fields	2006–2007								45.2

Table 2. Summarizing parameters used to calculate the ratios of N inputs to runoff and leaching N (Frac_{LEACH-(H)}), dissolved N₂O flux to N inputs (EI-EF_{5g}), and dissolved N₂O flux to soil N₂O emission (N₂O_(D)-N/N₂O_{direct}-N) in crop fields in Bear Creek watershed in 2006–2007. *F_{SN}* is annual amount of synthetic fertilizer N applied to crop fields and *F_{CR}* is N inputs from crop residue.

		Bear Creek watershed			
Factor	Unit	Corn fields	Soybean fields	Total	
Area ^a	ha	3404.95	3404.95	6809.9	
N Fertilizer application rate ^b	kg N ha ⁻¹ y ⁻¹	133.4	0	_	
FSN	kg N	908 440.7	0	908 440.7	
N residue rate ^c	kg N ha ⁻¹ y ⁻¹	92.2	82	_	
FCR	kg N	627 872.8	559 092.8	1 186 965.6	
FSN+FCR	kg N	1 536 313.4	559 092.8	2 095 406.2	
Runoff rate ^d	kg N ha ⁻¹ y ⁻¹	5.23	5.23	5.23	
Total runoff (TR)	kg N	35615.8	35615.8	71231.6	
Total leaching (TL) ^e	kg N	-	-	30 084.1	
TR+TL	kg N	-	-	101 315.6	
Frac ^f	kg N (kg N of N input) ⁻¹	_	-	0.05	
$N_2O_{(D)} - N^9$	kg N	_	_	45.2	
$EI-EF_{5g}^{h}$	kg N (kg N of N input) ⁻¹	_	_	0.00002	
N ₂ O _{direct} -N rate ⁱ	kg N ha ⁻¹ y ⁻¹	7.2	16.8	_	
N ₂ O _{direct} -N	kg N	49031.3	114 406.3	163 437.6	
N ₂ O _(D) -N/N ₂ O _{direct} -N	kg N (kg N) ⁻¹	-	-	0.0003	

^a Assumed 50% corn fields and 50% soybean fields. ^b Kim et al. (2009). ^c Kim et al. (2009). ^d Calculated from Lee et al. (2003). ^e Total NO₃-N flux in groundwater from crop fields in the Bear Creek watershed (Table 1).

 $^{\rm f}$ TR+TL/FSN+FCR. $^{\rm g}$ Total dissolved N₂O-N flux in groundwater from crop fields in the Bear Creek watershed (Table 1). $^{\rm h}$ Ratio of dissolved N₂O flux to N inputs, N₂O_(D)-N/FSN+FCR. $^{\rm i}$ Soil N₂O emission in the crop field in 2006–2007 (Kim et al., 2009).



Fig. 1. Map showing location of monitoring wells (•) and creek stilling wells (\blacksquare) in a coolseason grass filter (**A**) and a multi-species riparian buffer (**B**) in the Bear Creek watershed. In a cool-season grass filter (A), monitoring wells R8, R12, and R40 are adjacent to the creek and monitoring wells R39, R1, and R9 are adjacent a crop field. In a multi-species riparian buffer (B), monitoring wells R13, R17, and R21 are adjacent to the creek and monitoring wells R16, R20, and R24 are adjacent a crop field. Figures (A and B) are from Spear (2003).





Fig. 2. Seasonal variation of groundwater NO_3 -N concentration in groundwater under a coolseason grass filter and a multi-species riparian buffer in 1997–1999 (data from Spear, 2003) and 2005–2008.

	Crop field	Multi-species riparian buffer	Bear Creek	Cool-season grass filter	Crop field	
Groundwater flow direction	$\rightarrow \rightarrow$	\rightarrow \rightarrow		\leftarrow \leftarrow	← ←	Groundwater flow direction
Cl	20.6 (1.2)	20.9 (1.0)		13.4 (1.0)	13.2 (0.9)	Cl ⁻
NO ₃ -N	4.9 (0.5)	5 (0.4)		4.9 (2.4)*	9.5(0.7)*	NO ₃ -N
NO3 ⁻ /Cl ⁻	0.3 (0.1)	0.2 (0.0)		0.4(0.0)*	0.8(0.1)*	NO ₃ ⁻ /Cl ⁻
Dissolved N ₂ O-N	6.1(1.0)	6 (0.7)		6.8(0.8)	7.8(1.2)	Dissolved N ₂ O-N
DOC	1.1(0.1)	0.6 (0.4)		0.7(0.4)	0.9(0.4)	DOC
DO	3.4(0.5)	2.8(0.2)		2.6(0.3)	5(0.3)	DO
pН	7.5(0.0)	7.5(0.0)		7.3(0.0)	7.5(0.0)	рН

Fig. 3. Groundwater characteristics adjacent to crop fields and Bear Creek in a multi-species riparian buffer and a cool-season grass filter in 1997–1999 (data from Spear, 2003). Unit for Cl⁻, NO₃-N, DOC, and DO is mg L⁻¹ and unit of dissolved N₂O-N is μ g L⁻¹. The value inside parenthesis is standard error of the mean and an asterisk (*) indicates *P*<0.05. The number of measurements: Cl⁻(*n*=21–23), NO₃-N (*n*=26–29), NO₃⁻/Cl⁻(*n*=17–22), dissolved N₂O-N (*n*=26–27), DOC (*n*=3), DO (*n*=19-=21), and pH (*n*=3).

	Crop field	Multi-species riparian buffer	Bear Creek	Cool-season grass filter	Crop field	
Groundwater flow direction	$\rightarrow \rightarrow$	\rightarrow \rightarrow		\leftarrow \leftarrow	<i>← ←</i>	Groundwater flow direction
Cl	20.8 (1.2)	18.2 (0.6)		18.2 (0.6)	20.6 (1.2)	Cl
NO ₃ -N	4.0 (0.3)*	2.0 (0.2)*		3.3 (0.3)*	7.9 (0.5)*	NO ₃ -N
NO ₃ -N flux	4.4	2.1		5.1	14.2	NO ₃ -N flux
NO3 ⁻ /Cl ⁻	0.2 (0.1)*	0.1 (0.0)*		0.2 (0.0)*	0.4 (0.0)*	NO3 ⁻ /Cl ⁻
Dissolved N ₂ O-N	9.0 (1.1)	9.1 (1.3)		14.4 (2.2)	11.6 (1.5)	Dissolved N ₂ O-N
Dissolved N ₂ O-N flux	8.3	7.7		20.0	19.7	Dissolved N ₂ O-N flux
DOC	1.2 (0.1)	1.0 (0.1)		1.1 (0.1)	1.9 (0.1)	DOC
DO	3.3 (0.3)	2.7 (0.3)		2.7 (0.3)	3.3 (0.5)	DO
pH	7.4 (0.1)	7.4 (0.0)		7.4 (0.1)	7.5 (0.1)	pН

Fig. 4. Groundwater characteristics (in 2005–2008) and NO₃-N and dissolved N₂O-N fluxes (in January 2006–December 2007) adjacent to crop fields and Bear Creek in a multi-species riparian buffer and a cool-season grass filter in 2005–2008. Unit for Cl⁻, NO₃-N, DOC, and DO is mg L⁻¹, dissolved N₂O-N is μ g L⁻¹, NO₃-N flux is kg N (2006 and 2007 years)⁻¹, and dissolved N₂O-N flux is g N (2006 and 2007 years)⁻¹. The value inside parenthesis is standard error of the mean and an asterisk (*) indicates *P*<0.05. The number of measurements: Cl⁻(*n*=29), NO₃-N (*n*=29), NO₃⁻/Cl⁻(*n*=29), dissolved N₂O-N (*n*=25–26), DOC (*n*=8), DO (*n*=26–27), and pH (*n*=21).



Fig. 5. Seasonal variation of the NO_3^-/CI^- ratio in groundwater under a cool-season grass filter and a multi-species riparian buffer in 1997–1998 (data from Spear, 2003) and 2005–2008.



Fig. 6. Seasonal variation of dissolved N_2O -N concentration in groundwater under a coolseason grass filter and a multi-species riparian buffer in 1997–1999 (data from Spear, 2003) and 2005–2008.



Fig. 7. Relationship between NO₃-N and dissolved N₂O-N concentration in groundwater discharged from crop fields **(A)** and riparian buffers (a cool-season grass filter and a multi-species riparian buffer) **(B)** in 1997–1999 (data from Spear, 2003) and 2005–2008 in this study (•, n=99-101) and data from Davis et al. (2007) (\bigcirc , n=7). Default of EF_{5g} (IPCC, 2007) (--), and EF_{5g} of this study (**—**) and 95% confidence interval of the EF_{5g} of this study (**—**).