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Supplement of

Organic matter quality of deep permafrost carbon – a study from Arctic Siberia

J. Strauss et al.

Correspondence to: J. Strauss (jens.strauss@awi.de)
1 Supplementary methods

1.1 Grain-size analyses

To disaggregate the sample and to measure only clastic grains, organic components were removed by adding 35 % hydrogen peroxide three times a week to the samples. The samples were continuously shaken (Innova 2300, New Brunswick) for 6 weeks. The organic-free samples were ‘washed’ to neutral pH values by repeated decantation after centrifugation (Heraeus Cryofuge 8500i, Thermo Scientific). Samples were dispersed in 1% ammonia solution and 1 g dispersing agent (Na₄P₂O₇ × 10H₂O) and shaken (RS12 Rotoshake, Gerhardt) for at least 5 hours. Finally, each sample was split by a rotary cone sample divider (Laborette 27, Fritsch) and two sub-samples of each main sample were analyzed using a laser particle sizer (LS 200, Beckmann-Coulter). Grain sizes between 0.375 and 1000 µm were determined (Fig. 2, S1). Because no grain fractions >1000 µm occurred, there was no need for fraction reintegration. Grain-size calculations were done after Folk and Ward (1957) using Gradistat v8 (Blott and Pye, 2001).

1.2 Elemental composition

To determine the total elemental carbon and total nitrogen (TN) content, the samples were freeze-dried (sublimator 3-4-5, Zirbus Technology) and homogenized by grinding (Pulverisette 5 planetary mill, Fritsch) before being measured by a carbon-nitrogen-sulphur analyzer (Vario EL III, Elementar). Two 5 mg sub-samples per sample were taken and encapsulated in two zinc capsules. For ensuring complete oxidation of the sample during measurements ~10 mg of tungsten-(VI)-oxide was added. Background signals were detected by measuring a blank capsule at the beginning and calibration standards after each 20 samples. The quantified measurement accuracy is <± 0.1 wt %. TOC wt% was measured with a TOC analyzer (Vario Max C, Elementar). Therefore, depending on the previously measured TC content, up to 100 mg were weighed in a crucible and analyzed. The C/N ratio was calculated as the quotient of TOC wt% and TN values. The volumetric TOC content (TOC kg/m³) was calculated according to Strauss et al. (2013).

1.3 Bulk density and volumetric carbon content

Bulk density (BD) measurements were performed in two steps; (1) in the field lab by determining the volume of frozen samples with the Archimedes Principle (quantifying the water displaced in a water-filled glass beaker using a balance (FCB 8K0.1, Kern; accuracy ±0.1 g), and (2) in the freeze-dried laboratory samples; BD was calculated using equation 1. Estimating the BD is required to convert the measured-weight-based TOC wt% content per sample to a volume-based value. Thus, the TOC kg/m³ was calculated according to equation 2.
1.4 Carbon isotope studies

Stable TOC carbon isotopes were determined with a Finnigan MAT Delta-S mass spectrometer combined with a FLASH elemental analyzer and a CONFLO III gas mixing system. Before analysis, samples were treated with hydrochloric acid (1.3 molar) and heated at 97°C for three hours to remove the carbonate. After that, the samples were washed to a neutral pH value, dried, and weighed in silver capsules. The sub-sample mass was dependent upon TOC wt% content of each sample. The δ\(^{13}\)C value was determined by the ratio of \(^{13}\)C to \(^{12}\)C atoms expressed in per mille (‰). In order to quantify the deviations of the \(^{13}\)C/\(^{12}\)C ratios, the international standard Vienna Pee Dee Belemnite (VPDB) is used.

Ages were determined by radiocarbon dating of selected macroscopic plant remains. The present measurements were performed at the Poznań Radiocarbon Laboratory, Poland, using compact carbon accelerator mass spectrometry (AMS) (Goslar et al., 2004). The presented radiocarbon ages are uncalibrated ages; Table 1 includes calibrated ages as well. Radiocarbon ages are given in year before present (a BP).

2 Supplementary figures

![Grain-size distribution of a) Yedoma and b) thermokarst profiles.](image)

**Figure S1.** Grain-size distribution of a) Yedoma and b) thermokarst profiles. The different profiles are visualized in different shades of grey.
**Figure S2.** Histogram of the n-alkanes. The histograms are sorted stratigraphically from left (old) to right (young). The histograms of the Yedoma deposits are visualized with a blue background.
Figure S3. Histogram of the $n$-fatty acids. The histograms are sorted stratigraphically from left (old) to right (young). The histograms of the Yedoma deposits are visualized with a blue background.

Figure S4. Rough schematic of $n$-alkane chain length in different organisms, after Killops and Killops (2009).
Figure S5. TOC variations from different Yedoma studies sorted from easternmost (Itkillik River, Alaskan North Slope) to westernmost (Mamontov Klyk, western Laptev Sea) studies (Schirrmeister et al., 2008a; Schirrmeister et al., 2008b; Schirrmeister et al., 2011; Strauss et al., 2013), including the present study from Buor Khaya Peninsula. The means are illustrated by the values; the range is shown by the bars.

**Supplementary References**


