



despite their bulk nature. Correlation with oxygen index values and  $\delta^{13}\text{C}$  was weaker assumingly indicating carboxylation of the peat during drier periods and enrichment of isotopically lighter peat components during decomposition, respectively. FTIR, C/N ratio, Pyrolysis-GC-MS analyses and Rock Eval hydrogen indices appear to reflect mass loss and related changes in the molecular peat composition during mineralization best. Different to the other investigated proxies, Pyrolysis-GC-MS and FTIR analyses allow disentangling decomposition processes and vegetation changes. UV-ABS measurements of alkaline peat extracts show only weak correlation with other decomposition proxies as they mainly reflect the formation of humic acids through humification and to a lesser extend mass loss during mineralization.

## 1 Introduction

Many studies used ombrotrophic bogs as archives to reconstruct climate or environmental conditions of the past (Barber, 2006; Barber and Langdon, 2007). Changes of the physical-chemical conditions through time or changes in vegetation offer different proxies for past environmental conditions (Chambers et al., 2012; Charman et al., 2009 and references therein). Some of these proxies, such as pollen, dust, volcanic ash or trace elements and pollutants from anthropogenic sources, are related solely to atmospheric deposition and may therefore be considered as “external” proxies, reflecting changes on a spatial or regional scale. Such proxies are thought to be mostly unaffected by internal processes of peat formation, although an influence of decomposition on trace element records has been reported recently (Biester et al., 2003, 2004, 2012). In addition to these external proxies, a number of internal proxies exist which describe changes of the bog vegetation (Booth, 2010; Charman et al., 2006; McMullen et al., 2004) or of the water table position over time, such as non-pollen palynomorphs (Mighall et al., 2006) or Testate amoebae (Hendon et al., 2001; Woodland et al., 1998). The strongest factor controlling peat accumulation is the ratio between peat formation and decay of organic matter (OM). This is mainly dependent on the climatic or hydro-

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logical conditions, and resulting redox conditions and the availability of nutrients. As in ombrotrophic bogs nutrient supply is limited to atmospheric deposition and therefore generally low, the most important factor for peat decomposition is the position of the water table. Due to the critical role of the water table (Ise et al., 2008), a bog's profile is generally separated into two sections: (i) the permanently water logged catotelm, characterized by anoxic conditions and slow peat decay, and (ii) the temporally aerated acrotelm, where decomposition is generally fast and most of the initial plant mass is mineralized (Kuhry and Vitt, 1996; Malmer and Holm, 1984; Malmer and Wallén, 2004). It is therefore generally recognized that peat decomposition is low during times of high water table levels or wet climatic conditions, but high during times of low water table levels or dry climatic conditions (Clymo, 1984). Accordingly, the degree of peat decomposition in the historic profile reflects the hydrological conditions during the time of peat formation, or more exactly at the time of peat burial in the catotelm. It has to be noted, though, that repeated dry-falling and re-wetting cycles might also affect deeper and older peat layers and modify and superimpose the historic record of peat decomposition. This latter process, named secondary decomposition, can therefore result in a decoupling of the age vs. depth relationship between peat formation and peat decomposition.

A problem in the discussion and comparison of changes in peat decomposition addressed in different studies is that the chemical changes during decomposition are often not clearly defined. In many studies the term humification is used, which solely addresses the formation of humic acids containing UV absorbing aromatic components (Blackford and Chambers, 1993; Klavins et al., 2008). The term decomposition as used here encompasses mass loss during mineralization causing selective preservation of more resistant compounds, and transformations of the remaining OM.

Since early investigations of peat bogs as climate archives in the 1920s, several chemical and physical methods have been developed to determine the degree of peat decomposition. The von Post scale (von Post, 1922) divides the degree of peat decomposition into ten grades, based on the colour of the manually squeezed peat pore

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water and the remaining solid material. Later, a number of mostly alkaline extraction procedures have been developed which all aim to extract and quantify humic or fulvic acids from the peat as a measure of peat decomposition. Blackford and Chambers (1993) introduced an alkaline extraction (NaOH) procedure combined with UV absorption measurements to determine differences in the degree of peat humification based on the leachate's color intensity. Such methods are based on the assumption that increasing peat decomposition leads to an increase in solubilized or extractable humic acids. However, also other factors can influence the abundance of humic acids, such as botanical changes (Yeloff and Mauquoy, 2006) or fire incidence.

Peat decomposition has also been intensively investigated by coal petrologists (Outridge and Sanei, 2010) often using Rock Eval<sup>®</sup> pyrolysis to determine H/C and O/C ratios (Carrie et al., 2012; Lafargue et al., 1998; Langford and Blanc-Valleron, 1990). This approach relies on the assumption that during decomposition of OM, labile compounds such as polysaccharides are preferentially decomposed while refractory aromatic or aliphatic compounds become residually enriched, causing a decrease of O/C and H/C ratios (Klavins et al., 2008).

Another common proxy of peat decomposition is the use of C/N ratios (Kuhry and Vitt, 1996; Malmer and Holm, 1984). This approach is based on the observed residual enrichment of N relative to C during mineralization of OM, i.e. more narrow C/N ratios of more decomposed peat material. Thus changes in C/N ratios are postulated to indicate mainly changes in bog surface wetness and concomitant changes in peat decomposition. However, C/N ratios differ widely between peatland plants (Hornibrook et al., 2000), and thus shifts in vegetation upon changes in surface wetness affect C/N ratios in peat and may obscure the signal of decomposition.

Due to the turnover of OM and associated C losses in mires, also a relation between peat decomposition and stable isotope inventories of hydrogen (H), C, N, sulphur (S) and oxygen (O) would be expected. However, the fate of H, C, N, S, O stable isotopes during peat decomposition and their use as indicators is discussed controversially. Some studies used C, N, S, O stable isotopes to reconstruct past temperature

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or humidity (Skrzypek et al., 2009; Jones et al., 2010) and found support that the isotopic signature of the peat reflects that of the former living plant and is not affected by decomposition, at least if specific parts of plants are investigated (Skrzypek et al., 2007). Other studies found a rapid decrease in  $\delta^{13}\text{C}$  in the upper peat layers due to the preferential decay of cellulose over lignins, the latter being isotopically lighter (Benner et al., 1987). Methane formation causes a strong increase in  $^{13}\text{C}$  content of the residual peat (Charman et al., 1999; Quay et al., 1999; Jones et al., 2010). Therefore, the use of stable isotopes to elucidate decomposition processes is so far not fully constrained.

Klavins et al. (2008) compared a number of indicators for peat humification based on changes in some quality indices. They distinguished between a humification index, pyrophosphate index, humus quality, peat humification index, humification degree and several element ratios such as the H/C, O/C and the N/C ratio. Most of these indices are, however, operationally defined and it remains unresolved by which biogeochemical processes these indices are controlled.

A spectroscopic approach to overcome the limitations of operationally defined parameters of peat decomposition is the use of FTIR spectroscopy. This technique is used to distinguish the principal chemical classes in OM such as carbohydrates, lignin, cellulose, aliphatics, lipids and proteins through the vibrational characteristics of their chemical bonds (Artz et al., 2008; Coccozza et al., 2003; Senesi et al., 1991). Several studies used the relative intensities of FTIR absorption bands to evaluate the degree of peat decomposition (e.g. Artz et al., 2008; Broder et al., 2012; Kalbitz et al., 1999). An overview of FTIR absorption bands of peat humic acids and their assignment to defined chemical groups in peat can be found in Niemeyer et al. (1992).

Pyrolysis-GC-MS is a powerful method to characterize complex mixtures of OM such as peat (Buurman et al., 2006; Kracht and Gleixner, 2000; van der Heijden and Boon, 1994; van der Heijden et al., 1990; van Smeerdijk and Boon, 1987) and has been successfully applied to determine past environmental conditions from peat records (Boon et al., 1986; Kuder and Krüge, 1998; McClymont et al., 2011; Schellekens et al., 2011, 2012). Pyrolysis-GC-MS provides on one hand molecular vegetation markers, which

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impact mode, 70 eV) at 230 °C the quadrupole ion filter and the detector at 150 °C scanning a range between  $m/z$  50 and 500. Compounds were identified using the NIST '05 library.

For the eight analysed plant species all visible peaks were identified, of which six pyrolysis products appeared to be valuable as markers for one species or for a group of species. A marker is here defined as a pyrolysis product which is (i) exclusively found in one of the investigated species and (ii) well preserved in the peat. The term "marker" is used here for pyrolysis products that were specific within the analysed data set and in previous studies (Schellekens et al., 2009, 2011, 2013). Such markers may only be valid within the current study, as their presence in other species cannot be excluded, and thus the use of the term "marker" is provisional.

Based on frequency, abundance, and potential as vegetation marker, a selection of 84 pyrolysis products was quantified for all 42 peat samples of the KK core. Quantification was based on the peak area of characteristic ions ( $m/z$ ) from each pyrolysis products (Supplement Table S1). For each sample the sum of the quantified peak areas was set at 100% and amounts were calculated relative to this. According to probable origin and similarity, the pyrolysis products were grouped as follows: polysaccharides, aliphatics ( $n$ -alkanes,  $n$ -alkenes, methyl ketones, fatty acids, other aliphatic compounds), lignins, phenols, aromatics, polyaromatics, and nitrogen compounds.

## 2.2.2 C/N ratios and C and N stable isotope analyses

Concentrations of C and N were determined in the carbonate free, acidic peat by GC-TCD after thermal combustion in an elemental analyzer (Euro EA3000, Eurovector). Reproducibility of duplicates was always better than  $\sim 8\%$  RSD.

Ratios of  $^{13}\text{C}/^{12}\text{C}$  and  $^{15}\text{N}/^{14}\text{N}$  and total C and N were quantified in dried bulk plant material and peat on mass basis using an elemental analyser (CE instruments NA 1108, Milano, Italy), connected via ConFlo III interface to a delta S IR-MS (Thermo Finnigan MAT, Bremen, Germany). Isotope signatures are given in the common  $\delta$ -

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notation  $\delta\text{‰} = ((R_{\text{sample}}/R_{\text{standard}}) - 1) \times 10^3$ , relative to the V-PDB-standard or  $\text{N}_2$  in ambient air.

## 2.2.3 FTIR spectra

FTIR spectra of the ground peat samples were obtained using a Vector 22 FTIR spectrometer (Bruker Optik, Ettlingen, Germany; absorption mode, subsequent baseline subtraction) on KBr pellets (200 mg dried KBr and 2 mg sample). Measurements were recorded from 4500 to 300  $\text{cm}^{-1}$  using a resolution of 2  $\text{cm}^{-1}$ . A number of 32 scans per sample were averaged. Absorption peaks indicative of structural units in OM were identified according to Senesi et al. (1989) and Niemeyer et al. (1992) and used as an indicator of peat OM quality. Absorption at 1420  $\text{cm}^{-1}$  is ascribed to OH deformations and CO stretch of phenols or CH deformation of  $\text{CH}_2$  or  $\text{CH}_3$  groups (phenolic and aliphatic structures), at  $\sim 1510 \text{ cm}^{-1}$  to aromatic C=C or to CO of amide groups. The  $\sim 1630 \text{ cm}^{-1}$  region is indicative of aromatic C=C and asymmetric  $\text{COO}^-$  group vibrations (lignin and other aromatics and aromatic or aliphatic carboxylates) and  $\sim 1720 \text{ cm}^{-1}$  of CO stretch of carbonyl and carboxyl groups (carboxylic acids and aromatic esters). The 2920  $\text{cm}^{-1}$  and 2850  $\text{cm}^{-1}$  bands reflect C-H stretching vibrations, and 460  $\text{cm}^{-1}$  can be assigned to substituted aromatics and unsaturated aliphatic chains, which agrees with aromatic and aliphatic pyrolysis products. However, this band may be influenced by minerals such as clays and is thus not further considered here. Multiple signals at 1707  $\text{cm}^{-1}$  (C=O), 1620  $\text{cm}^{-1}$  (C=O, C=C), 1510  $\text{cm}^{-1}$  (C=C), 1420  $\text{cm}^{-1}$  ( $\text{CH}_2$ , heterocyclics), 1370  $\text{cm}^{-1}$  (C-H, phenolics) and 1265  $\text{cm}^{-1}$  (C-O, C-O-C) agree with lignin or alike compounds; strong bands at 3370  $\text{cm}^{-1}$  (O-H), 1151  $\text{cm}^{-1}$  (C-O, cellulose, carbohydrates), 1032  $\text{cm}^{-1}$  (C-O, cellulose, carbohydrates) and 891  $\text{cm}^{-1}$  (O-H, carbohydrates) bands agree with polysaccharides.

To determine relative abundances of functional groups as a humification index, the ratio between peak intensity at 1630  $\text{cm}^{-1}$  with respect to polysaccharides (1053  $\text{cm}^{-1}$ ) was calculated (Holmgren and Norden, 1988; Niemeyer et al., 1992).

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## 2.2.4 Oxygen and hydrogen indices using the Rock Eval® method

Rock Eval analyses were performed at the Alfred Wegener Institut für Polar- und Meeresforschung in Bremerhaven, Germany, using a Rock-Eval II (plus S3 unit). The method determines several operational defined groups of organic compounds based on pyrolysis at different temperatures and different H/C and O/C ratios. The obtained results allow distinguishing between labile and increasingly refractory OM fractions (Tissot and Welte, 1978). Pyrolysis is conducted on bulk peat samples (20 mg) to determine the amount of (i) hydrocarbons present in the sample (S1 peak in milligrams hydrocarbons per gram of peat, data not shown), (ii) the amount of hydrocarbons generated by pyrolytic degradation of the peat during heating up to 550 °C (S2 peak in mg hydrocarbon per gram sediment) defined as the Hydrogen Index (HI) when normalized to the total organic C content, and (iii) the amount of carbon dioxide generated during heating up to 390 °C (S3 peak in milligrams carbon dioxide per gram sediment), which gives the Oxygen Index (OI), when normalized to the total OM content (Lafargue et al., 1998).

## 2.2.5 Humic acids extraction and UV-absorption

The colorimetric method for characterizing the degree of decomposition was adopted by Blackford and Chambers (1993). For this study 0.1 g pulverized and freeze-dried peat samples were dissolved in 50 mL NaOH (8 %) and placed on a hot plate for 1 h to simmer. After cooling down the extract was filled up to 100 mL with deionized water and filtered through a Whatman No. 1 filter paper. Then 25 mL of the extract were diluted to 50 mL prior to the colorimetric measurement. The absorbance of light (in %) was measured with a Perkin-Elmer UV-Vis spectrometer at a wavelength of 540 nm.

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## 2.2.6 Statistical analyses

Statistical analyses were performed using PASW Statistics 18.0 (IBM Corporation). All data were tested for correlation with a level of significance ( $p$ ) of 0.01 and 0.05 (Pearson, two-tailed). Principal component analysis (PCA) was applied to extract the chemical signatures of the OM composition to derive the underlying factors accounting for them. The analysis was performed in the Z-scores transformation of the residuals (Z-score =  $(X_i - X_{avg})/X_{std}$ ; where  $X_i$  is a given value of a variable in a sample,  $X_{avg}$  is the average of that variable and  $X_{std}$  is its standard deviation). Z-scores allow keeping the relative variation of the original data while reducing all variables to a similar range of variation avoiding scaling effects (Eriksson et al., 1999). For the pyrolysis products, factor analysis without rotation was performed using Statistica 6 (StatSoft, Tulsa).

## 3 Results

Results of C and N analyses of plants show highest C/N ratios for *Sphagnum* as compared to *Calluna vulgaris* (Ericaceae) and *Eriophorum* (Cyperaceae), which are considered as the most important peat forming plants in the Harz bogs (Table 1).  $\delta^{13}\text{C}$  values range between  $-27.09\text{‰}$  and  $-29.21\text{‰}$  and are therefore similar in the three investigated species, but slightly lighter in *C. vulgaris*. In contrast to  $\delta^{13}\text{C}$ ,  $\delta^{15}\text{N}$  vary in a broad range between  $-0.87\text{‰}$  and  $-8.13\text{‰}$ , with *C. vulgaris* being lightest.

### 3.1 Records of decomposition proxies

#### 3.1.1 Königsmoor (KK)

C/N ratios in the KK core show lowest values in the surface layer and increase down to a depth of 50 cm where they reach their maximum (Fig. 1). Below 50 cm they show some variation but no more trends towards the end of the core. A similar trend of increasing values with depth is also observed for  $\delta^{13}\text{C}$  values as well as for UV-ABS

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and  $\delta^{15}\text{N}$ . HI and OI values both do not show a trend but intense changes throughout the core. The records of both values seem to show inverse relationship with highest OI and lowest HI values in the surface layer. Most intense variability is observed in the section between  $\sim 38$  and 50 cm depth corresponding to similar intense changes of the other proxies in the same section. The humification index ( $1630/1053\text{ cm}^{-1}$ ) based on FTIR also greatly varies throughout the core (Fig. 1). Values are comparatively high in the upper 40 cm and show a distinct low between 38 and 50 cm depth. UV-ABS is lowest in the surface layer and increases down to 40 cm by a factor of 2.5 after a pronounced low between 38 and 50 cm values again slightly increase towards the bottom of the core without larger changes. There are, however, little common features with any other indicator. Also  $\delta^{15}\text{N}$  values vary in a broad range throughout the profile with a weak trend of decreasing values towards the bottom of the core, but also did not seem to be related to the other indices under study.

Depth distribution of the FTIR spectra of the KK peat core is given in Fig. S1 in the Supplement. The distribution of FTIR band intensities can be roughly divided into two groups. Group I mainly reflects polysaccharides (band  $1053\text{ cm}^{-1}$ ) and show a slight decrease of values in the uppermost 38 cm, whereas Group II include aromatics (bands  $1420, 1510, 1630\text{ cm}^{-1}$ ) and aliphatics (bands  $2850, 2920\text{ cm}^{-1}$ ) and show generally higher intensities in the upper 38 cm.

### 3.1.2 Kleines Rotes Bruch (KRB)

C/N ratios in the KRB core exhibit a similar course than those found in the KK core. Values are higher in the top layer but decrease to minimum values between 5 and 10 cm (Fig. 2). From 10 to about 50 cm C/N ratios steadily increase without larger changes and reach maximum values between 50 and 70 cm. A remarkable sharp decrease of the C/N ratios occurs between 65 and 70 cm, also indicated by changes in the other decomposition proxies. Different from the KK profile  $\delta^{13}\text{C}$  values in the KRB core do not exhibit a trend, but show intense changes throughout the core. OI and HI

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indices show a distribution comparable to that observed in the KK profile. Again lowest HI values are observed in the upper 40 cm and the inverse course in the low part of the core interrupted by several short term changes. The development of the FTIR based humification index with depth is also comparable of what is found in the KK core. Highest values are found in the upper 40 cm followed by a section of low values between 40 and 65 cm and a strong sharp increase up to maximum values between 65 and 70 cm. UV-ABS values do not show a trend as observed in KK. They are lowest in the surface layer and steadily increase down to about 30 cm. Two pronounced maxima are observed between 30 and 45 cm and 65 to 70 cm where highest values occur (Fig. 2). No trend in  $\delta^{15}\text{N}$  could be observed but intense changes especially in the upper 60 cm of the core. As for the KK site, in most sections, changes in  $\delta^{15}\text{N}$  do not correspond to those of any other decomposition proxy.

FTIR band intensities of KRB peat show similar distributions than those of KK peat (Supplement Fig. S2). In the upper 45 cm, with the exception of the uppermost 6 cm which reflect the living plants, bands indicative for aliphatics ( $2920, 2860\text{ cm}^{-1}$ ) and aromatics ( $1630, 1510, 1420\text{ cm}^{-1}$ ) show higher intensities than in deeper peat sections. The uppermost 50 cm are characterized by a minimum between 18 and 25 cm followed by a pronounced peak of the aliphatic and especially the aromatic bands (25–35 cm). Another peak of aliphatic and aromatic band intensities is found at 65 cm. In contrast, bands indicating polysaccharides ( $1053\text{ cm}^{-1}$ ) show generally slightly higher values in the lower 50 cm and a trend of increasing values with depth. In some sections of the core, especially between 30 and 50 cm, polysaccharides band intensities appear to develop invers to the aliphatic and aromatic bands. High intensities of these bands are also found in the uppermost  $\sim 10$  cm where the high intensities of the polysaccharide band are attributed to the high cellulose and hemicellulose content in the living or only slightly decomposed plants.

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## 3.2 Molecular composition of KK peat determined by pyrolysis-GC-MS

Comparing the results for the traditional decomposition proxies and C and N isotopes in the two peat cores indicates that these proxies show similar relationship in both bogs. For that reason, molecular components were only determined in the KK core, assuming a similar relationship between the molecular components and the traditional decomposition proxies in both bogs.

### 3.2.1 Vegetation markers

Based on analysis of plant species (*Eriophorum vaginatum*, *Sphagnum* and *Calluna vulgaris*) and on previous plant analyses several markers were identified. 4-Isopropenylphenol is specific for *Sphagnum* moss (Ph5) (Rudolph and Samland, 1985; van der Heijden et al., 1997), additionally two biphenyl compounds were solely detected in *Sphagnum* (Ph7, Ph9; Supplement Table S1). 3-Methoxy-5-methylphenol (Ph6,  $m/z$  107, 109, 138) is characteristic for several lichen species (Schellekens et al., 2009, 2011). Ferulic acid methyl ester (Ph8,  $m/z$  145, 177, 208) was only detected in *E. vaginatum* and has been proposed as a marker of graminoids (Schellekens et al., 2011, 2013). A sesquiterpene (St1) was solely detected in the pyrolysate of *Pinus*. Three unidentified compounds with  $m/z$  86 and 114 were present in the peat OM, but were not detected in any of the analyzed plant species. All three probably originate from the same macromolecule as the difference between their molecular masses ( $M^+$  182, 210 and 238) point towards two  $\text{CH}_2$  groups. Only the compound with molecular mass 210 (unkn3) was quantified, as it showed the highest abundance.

### 3.2.2 Peat record

The mean abundance of groups of pyrolysis products for the KK peat record is given in Table 2. A detailed listing of all quantified pyrolysis products and their mean abundance in the peat OM is given in Supplement Table S1. As shown in Table 2, polysac-

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charides are by far the most abundant organic compound in the peat followed by much smaller amounts of aliphatics, lignins, phenols and aromatics. We selected five molecular proxies including plant-specific markers for *Sphagnum* and pine, and groups of pyrolysis products. Based on their distribution these proxies can roughly divided into two groups, where polysaccharides and the marker of *Sphagnum* (4-isopropenylphenol) on one hand and aliphatics and the marker of pine (sesquiterpene) on the other, showed similar changes (Fig. 3), whereas the distribution of lignin pyrolysis products shows smaller variations and less similarity to the distribution of aliphatics. All five groups of components except lignin show the highest variability in the upper 50 cm of the core. Polysaccharides and 4-isopropenylphenol show a trend of declining values towards the surface with pronounced minima between about 8 and 15 and 25 and 38 cm depth and a clear maximum between 38 and 50 cm depth (Fig. 3). Aliphatics and sesquiterpene show the opposite trend, also increasing below  $\sim 70$  cm.

## 4 Discussion

### 4.1 Comparison of decomposition proxies

Principal component analysis of decomposition proxies reveals minor differences if applied to the cores KK and KRB separately (Supplement Table S2). For both cores most of the variance (49 and 53.5%) is explained by the first component. All used decomposition proxies show high loadings on the first component in both cores except  $\delta^{15}\text{N}$  and UV-ABS, which both load significantly on a second component in the KRB core and OI, which loads higher on the second component of the KK PCA. KK peat shows three components with higher loadings of OI and  $\delta^{15}\text{N}$  on the second and high loadings of UV-ABS on the third component. PCA gives two components if the data of both cores are analyzed together, with C/N ratios, HI, OI, FTIR and  $\delta^{13}\text{C}$  loading high on the first and UV-ABS and  $\delta^{15}\text{N}$  on the second component.

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From this it is likely that variations in all proxies, except of  $\delta^{15}\text{N}$ , are predominately attributed to changes in the degree of peat decomposition and in vegetation. The occurrence of the C and N isotope data in a second component can be explained by additional fractionation processes, which are independent from or only indirectly linked to decomposition such as changes in temperature, humidity, plant species or N recycling. This seems to be especially true for  $\delta^{15}\text{N}$ , which we assume is mostly influenced by N-recycling, whereas  $\delta^{13}\text{C}$  appears to be mainly influenced by decomposition at least in the KRB core. This influence becomes more pronounced when the data of both cores are analyzed together and significant loadings of  $\delta^{13}\text{C}$  disappear from the second component. The appearance of UV-ABS in a second or third component independent from the other proxies was unexpected as UV-ABS of NaOH-extracts has been frequently used to determine the degree of peat humification. The missing correlation between UV-ABS and the other decomposition proxies indicates that changes in mass loss affecting C/N-ratios, FTIR band intensities, HI, OI and  $\delta^{13}\text{C}$  is not coincident with the formation of humic acids, the main process underlying changes in UV-ABS of NaOH peat extracts. However, changes in vegetation controlled by hydrology, i.e. shifts in vascular plant species (containing lignin) in peat which is dominated by *Sphagnum* (not containing lignin), during drier periods may also lead to changes in the abundance of humic acids (Yeloff and Mauquoy, 2006). Changes in humic acids may thus reflect more a signal of vegetation changes than of changes in humification or decomposition alone.

C/N ratios and the FTIR humification index have been used in a number of studies as indicator of peat decomposition or, more exactly, as a measure of mass loss caused by increased mineralization during water table lows, although most studies do not consider changes in the botanical setting. The influence of plants on one hand and decomposition on the other is a key issue of this study and will be discussed below. However, results from previous studies on trace element distribution in the same samples (Biester et al., 2012) show significant negative correlation between C/N ratios and trace element (e.g. Br, Ti, Zr) enrichment, confirming the major influence of mass

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loss on C/N in peat dominated by *Sphagnum*. Thus, C/N ratios in these *Sphagnum* dominated cores predominantly indicate mass loss attributed to peat decomposition and to a lesser extend shifts in vegetation. While a number of studies address this shortcoming by analyzing specific plant residues (e.g. remainders of *Sphagnum* only; Loisel et al., 2009; Skrzypek et al., 2007), this has not always been done and is also not always possible in more decomposed peat or if *Sphagnum* provides only a small proportion of predominant vegetation.

Outridge et al. (2010) investigated the relationship between mercury and peat decomposition in an arctic tundra mire. Although having not analyzed C/N ratios in their samples, they stated that the use of C/N ratios as indicator of peat decomposition might be misleading and proposed using the hydrogen index HI. In our study, C/N ratios instead correlated well and negative with hydrogen indices in both cores ( $R^2 = 0.74$  KK),  $R^2 = 0.74$  (KRB), but only weak with OI. The good correlation between C/N and HI is explained by a concurrent loss of mass (decreasing C/N) during decomposition of labile compounds and a relative enrichment of aliphatics (increasing HI indices). The concept of peat humification/coalification suggests increasing decarboxylation under anoxic conditions. C/N ratios, however, remain relatively stable under anoxic conditions and correlation between C/N ratios and OI indices could therefore not be expected. The weak correlation between C/N ratios and OI in our cores could be explained by oxidation of OM during low water tables, which does not generally lead to complete mineralization, but to an increase in carboxyl groups. Thus dry falling may promote two processes: an increase in OM associated oxygen groups and losses of (oxygenated) OM through mineralization; thus mass loss and losses of oxygen from OM do not necessarily have to be correlated, suggesting that the HI may be a better indicator for decomposition than the OI.

Correlations of C/N with intensities on FTIR band  $1510\text{ cm}^{-1}$  ( $R^2 = 0.69$ ) (lignins, phenols) and band  $1630\text{ cm}^{-1}$  ( $R^2 = 0.52$ ) (lignins, other aromatics, aliphatic carboxylates) were always negative and significant suggesting that lignins, aromatics and aliphatic carboxylates are enriched during peat mineralization. However, correlations

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between C/N ratios and pyrolysis products assigned to lignins or aromatics were significantly lower or absent, but, as will be discussed below, the intensities of the FTIR bands  $1510\text{ cm}^{-1}$  and  $1630\text{ cm}^{-1}$  are significantly correlated ( $R^2 = 0.7$  and  $R^2 = 0.48$ , respectively) with the first component (F1) obtained from the PCA of pyrolysis-GC-MS data. As F1 is mostly determined by variations in concentrations of polysaccharides and aliphatics it is most likely that both bands mainly reflect changes in the content of aliphatics and to a lesser extent variations in aromatics or lignins, here. This may be due to overlap of functional groups in the IR spectra (Coates, 2000), making an exact assignment of peaks difficult.

FTIR intensities, indicating lipids and waxes (bands  $2920\text{ cm}^{-1}$  and  $2850\text{ cm}^{-1}$ ), do not correlate with any of the other decomposition proxies significantly but show correlation with intensities of the bands  $1630\text{ cm}^{-1}$  and  $1420\text{ cm}^{-1}$  ( $R^2 = 0.74$  and  $0.62$ , respectively) which represent the group of lignins and other aromatics as well as the FTIR humification index ( $1630/1053$ ) ( $R^2 = 0.65$ ) (Supplement Table S3). This does probably indicate that the bands  $2920\text{ cm}^{-1}$  and  $2850\text{ cm}^{-1}$  do not solely absorb on fats and waxes as suggested previously (Cocozza et al., 2003; Niemeyer et al., 1992) but overlap with signals from polysaccharides and cellulose (Coates, 2000). HI solely correlate significantly with aromatics and lignin (band  $1510\text{ cm}^{-1}$  not shown) and F1 ( $R^2 = 0.6$ ) and the FTIR humification index ( $R^2 = 0.48$ ), which indicates that both, aliphatics and aromatics increase during decomposition due to the loss of polysaccharides and do both increase HI values. Such ambiguous information from FTIR data suggests that interpretation needs to be done with caution.

## 4.2 Interpretation of Pyrolysis-GC-MS data and PCA results

The main difficulty with the interpretation of decomposition proxies in peat is the fact that changes in bog hydrology simultaneously affect the botanical composition, both causing changes in the relative abundance of polysaccharides, lignin and aliphatics. For example, in *Sphagnum* dominated bogs a relatively high abundance of lignin indicates an increase of vascular plant species to the peat OM, as *Sphagnum* does not

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contain lignin. It also points to increased decomposition as in peat polysaccharides are preferentially degraded (Bracewell et al., 1980; Benner et al., 1984; van der Heijden et al., 1990) causing a relative increase of lignin and/or aliphatics. Accordingly, drier conditions simultaneously result in an increase of vascular plant species and an increase of aerobic decomposition. A relatively high abundance of lignin may thus be used as a proxy for hydrological reconstructions. However, if the relative abundance of lignin and polysaccharides are used as either vegetation or decomposition proxy it is problematic to separate between both factors. To disentangle the relationship between changes in plant species distribution on one side and decomposition changes on the other we applied a second factor analysis to all 82 quantified pyrolysis products of the KK core. To enable comparison with the decomposition proxies discussed above, they were included in the factor analysis. The first four factors explained 67.7% of the variance, of which the major part is explained by F1 and F2 (51.7%).

The loadings of the F1–F2 projection (Fig. 4) roughly distinguish three groups of pyrolysis products. The markers of *Sphagnum* (Ph5, Ph7 and Ph9) and most polysaccharides show negative loadings on F1 and positive loadings on F2. This indicates a high contribution of *Sphagnum* moss, which mainly yields polysaccharides and (poly)phenolic compounds upon pyrolysis. Most lignin pyrolysis products show negative loadings on F2 and positive loadings on F1; in *Sphagnum* dominated peat this points towards an increased input of vascular plant species as mosses do not contain lignin. This possibility to differentiate between contribution of *Sphagnum* and vascular plant residuals to the bulk peat is the particular value of the pyrolysis-GC-MS method. Aliphatics (except Al1), (poly)aromatics and nitrogen compounds show positive loadings on both F1 and F2 and indicate selective preservation and addition of microbial OM in peat (e.g. Schellekens et al., 2009).

The separation of polysaccharides, lignin and aliphatic pyrolysis products in F1–F2 projection suggests the following interpretation: F1 reflects the water table height at the time of plant death, with relatively dry conditions (positive on F1) causing an increase of vascular plants and aerobic decomposition. A lowering of the water table

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results in (i) a higher input of vascular plant material (negative on F2) and (ii) increased aerobic decomposition of *Sphagnum* moss (from negative on F1 to positive). F2 thus reflects the occurrence and degradation of vascular plants (lignin), (negative on F2), which increases under relative dry conditions (positive loadings on F1). A more detailed look confirms this interpretation: the marker of lichens (Ph6) is located with the lignin pyrolysis products, which confirms its preference for dryer habitats (Kleinebecker et al., 2007). Both lignin moieties with an acetyl side chain (Lg9, Lg17) show lowest loading on F2 and highest positive loading on F1 compared with other lignin pyrolysis products, which is in agreement with oxygenation of lignin alkyl side chains upon degradation (Kögel-Knabner, 2002; Schellekens et al., 2012). The main pyrolysis product of cellulose is levoglucosan (Ps13) (Pouwels et al., 1987); its neutral loadings on F2 (Fig. 4) agrees with the high abundance of cellulose in both, mosses and vascular plants. Xylose, which has a high abundance in hemicelluloses of grasses (Smith and Harris, 1999; Wende and Fry, 1997) and sedges (Bourdon et al., 2000), results in 4-hydroxy-5,6-dihydro-(2H)-pyran-2-one (Ps5) upon pyrolysis (Pouwels et al., 1987), which is the only polysaccharide pyrolysis product that show negative loadings on F2, as expected from high negative loadings of vascular plant derived lignins on F2.

### 4.3 Comparison of decomposition proxies and molecular peat components

PCA of the pyrolysis-GC-MS data shows that the hydrological controlled distribution of polysaccharides on one side and aliphatics and lignins on the other determine the variability in the molecular peat components through peat decomposition. Comparison of the decomposition proxies derived from KK peat records (Fig. 1) with the molecular compounds obtained by pyrolysis-GC-MS (Figs. 3, 4, and S3 in the Supplement) indicates that factor F1 and the group of aliphatics yield similar records as the hydrogen index and the FTIR-humification index. C/N ratios,  $\delta^{13}\text{C}$  and the oxygen index show similarity to the record of polysaccharides, which runs inversely to PCA factor F1. Under drier conditions, decomposition through mineralization of predominately polysaccharides is thus higher, but also the abundance of vascular plants, which causes an in-

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crease in lignins, HI values and a decrease of polysaccharides, C/N ratios,  $\delta^{13}\text{C}$  and OI values. Accordingly, wet conditions are indicated by the reverse trend in the distribution of the proxies attributed to higher abundance of *Sphagnum* in relation to lignin containing graminoids and less intense decomposition of polysaccharides (a detailed comparison of the proxy records is given in Supplement Fig. S3).

#### 4.3.1 C/N ratios

In detail, F1 is negatively correlated to the C/N ratio ( $R^2 = 0.74$ ; Supplement Table S2) and positively correlated to polysaccharides ( $R^2 = 0.66$ ; Table 2), which means that – not surprisingly – a higher content of polysaccharides correlates with a lower degree of peat decomposition. However, the C/N ratio is similarly affected by the effects of both botanical changes and aerobic decomposition upon changes in hydrology. The C/N ratio decreases with decomposition but also highly differs between *Sphagnum* spp. and vascular plants (e.g. Hornibrook et al., 2000). Table 1 shows slightly higher C/N ratios for the analyzed *Sphagnum* species compared to graminoids and ericoids. However, C/N ratios in *Sphagnum* from the Harz sites (Table 1) are much lower than those in *Sphagnum* from remote sites (Schmidt et al., 2010), which is most likely a result of the high atmospheric N deposition in the Harz Mountains in the past decades, as large parts of atmospheric N are captured by *Sphagnum* moss (Heijmans et al., 2002). *Sphagnum* from before the 20th century is likely to show similar C/N ratios than those from present day remote sites (e.g. 89–134 in Broder et al., 2012). Independent of the initial C/N ratio in the upper part, the highest negative loading of C/N on F1 and the loading of almost zero on F2 (Fig. 4) indicate that despite slight shifts in vegetation, C/N ratios here provide a reasonable estimate of the degree of decomposition. In a peat record from Tierra del Fuego major shifts in C/N ratio perfectly agreed with the main botanical transitions, from *Sphagnum* dominated to *Sphagnum-Juncus* to *Juncus* dominated peat (Schellekens and Buurman, 2011). In another peat record dominated by graminoids, an increase in lignin and aliphatics was related to decom-

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position (Schellekens et al., 2011, 2012), but no correlation with C/N ratio was found. However, within *Sphagnum*-dominated peat (as in our peat of the KK site), shifts in C/N ratio mainly reflect decomposition, as due to changes in hydrology. The fact that in our KK peat not lignin but aliphatics showed the better correlation with C/N ratios ( $R^2 = 0.60$ ; Supplement Table S2) further confirms this. Because vascular plants increase under relatively dry conditions in *Sphagnum*-dominated peat, this automatically indicates a higher degree of aerobic decomposition (positive loadings of lignin on F1, Fig. 4). A combination of vegetation and decomposition proxies is expected to best reflect changes in bog hydrology, as vegetation shifts are probably slower than decomposition (which is a continuous process); on the other hand secondary decomposition may disturb the hydrological interpretation when only decomposition proxies are used.

### 4.3.2 FTIR

Several bands of FTIR intensities show significant correlation with factor F1 of the Pyrolysis-GC-MS data (Fig. 5, Supplement Table S3). Best correlation was found between the bands 1510 and 1630 (aromatic structures in lignins or aliphatic carboxylates) and F1 (Supplement Table S3). Positive correlation with F1 was found for the bands 2920 ( $R^2 = 0.5$ ), 2850 ( $R^2 = 0.57$ ) (aliphatic structures) and 1420 ( $R^2 = 0.55$ ) (carboxylate structures), which was even higher if these bands are correlated to aliphatics, the major components behind F1. Negative correlations with F1 were found for the bands 891 ( $R^2 = 0.73$ ), 1151 ( $R^2 = 0.58$ ) and 1053 ( $R^2 = 0.36$ ) (polysaccharides), which were slightly weaker if directly correlated to aliphatics (negative) or polysaccharides (positive) determined by pyrolysis-GC-MS (Table 3). Significant correlation between the group of lignins determined by pyrolysis-GC-MS and FTIR-bands specific for lignin components (mainly phenolic and aromatic structures) could not be found. This could be related to the fact that also *Sphagnum* contains high amounts of lignin-like phenolic compounds. Comparable to what has been found by pyrolysis-GC/MS intensity changes of the FTIR bands mainly reflect the changes in the peat's concentrations of aliphatic and aromatic structural components and polysaccharides, while signals for 17373

lignin may be influenced by too much overlapping with other functional groups. Nevertheless, ratios of intensities on band 1630 (aromatic structures in lignins or aliphatic carboxylates) and those of band 1053 (0.36) (polysaccharides), which has been suggested as a FTIR humification index, are significantly correlated to F1 ( $R^2 = 0.67$ ), aliphatics ( $R^2 = 0.58$ ), polysaccharides ( $R^2 = 0.55$ ), 4-isopropenylphenol ( $R^2 = 0.48$ ) (*Sphagnum*) and a sesquiterpene (*Pinus*) ( $R^2 = 0.43$ ). A unique interpretation of the FTIR bands is obviously constrained by their overlap in complex OM such as peat, a shortcoming which may only partly be resolved by relating FTIR based indices to molecular markers. FTIR data, however, confirms the interpretation that decomposition on a molecular level in the KK peat record is mainly indicated by an increase in aliphatics and aromatic structures and a decrease in polysaccharides. A comparison of the FTIR intensity records and molecular components is given in Supplement Fig. S4.

### 4.3.3 Rock-Eval pyrolysis

The O/C ratio in peat is expected to be high in sections where decomposition is low and higher amounts of oxygen containing compounds occur (e.g. polysaccharides). According to the van Krevelen concept of coal petrology the oxygen index is expected to decrease during proceeding OM decomposition and coalification (Tissot and Welte, 1978). Although the OI index showed high values during the *Sphagnum* dominated period (38–48 cm, Fig. 1 and 3(4-isopropenylphenol)) and also in the uppermost sample, correlations with other pyrolysis-GC-MS proxies were rather low (Supplement Table S3). Significant correlation ( $R^2 = 0.41$ ) was only found for a group of polysaccharides ( $m/z$  57, 69, 82) and for 4-isopropenylphenol, which is specific for *Sphagnum*. Higher values of HI (equivalent to high H/C) in peat deposits have been interpreted as hydrogen-rich labile C compounds in unprocessed OM (Outridge and Sanei, 2010). However, the HI index is thought to reflect aliphatic structures in general and the HI record of KK peat was positively correlated ( $R^2 = 0.49$ ) with aliphatics determined by Py-GC-MS and with FTIR band 460 (aliphatics) ( $R^2 = 0.58$ ) (not shown), but only weakly ( $R^2 = 0.32$  and  $0.26$ ) to the bands  $2850\text{ cm}^{-1}$  and  $2920\text{ cm}^{-1}$ , respectively

(lipids, waxes, CH<sub>2</sub> stretching (2850 cm<sup>-1</sup>) (Supplement Table S3). Negative, but higher correlation ( $R^2 = 0.58$ ) was found with band 891 cm<sup>-1</sup> indicative of polysaccharides. Thus, the HI record in KK peat also reflects the relative enrichment of aliphatics compared to more labile polysaccharides such as cellulose as we have already concluded from the results of pyrolysis-GC-MS measurements. The H/C ratio is often found to decrease with burial depth and regarded to reflect a maturity enhanced conversion of aliphatic C to fixed aromatic C under Rock-Eval conditions (Schenk et al., 1986). However, such assumptions seem not to hold true for peats with a low diagenetic grade, in which decomposition of polysaccharides results in a relative increase of aliphatics. A transformation to aromatic compounds seems hardly relevant, which would explain the absence of a correlation between HI and aromatics and only a weak correlation between HI and polyaromatics in KK peat.

#### 4.3.4 C and N isotope ratios ( $\delta^{13}\text{C}$ , $\delta^{15}\text{N}$ )

The information derived from  $\delta^{13}\text{C}$  values in peat are discussed controversial. C isotopes in peat have been assigned to several processes, among which are the inherent isotopic signals of the plants due to e.g. the photosynthesis pathway, the temperature during plant formation at the bogs surface, bog surface wetness and by decomposition processes. Jedrysek and Skrzypek (2005) stated that peat diagenesis has generally no influence on  $\delta^{13}\text{C}$  in *Sphagnum* or extracted cellulose. Kaislahti et al. (2010) reported a dependency of  $\delta^{13}\text{C}$  in young peat on temperature and could also not find a relationship with peat decomposition patterns. Several studies stated that *Sphagnum*  $\delta^{13}\text{C}$  reflect surface moisture conditions (Rice and Giles, 1996; Price et al., 1997; Ménot and Burns, 2001; Loisel et al., 2009). While some studies found that  $\delta^{13}\text{C}$  values were lower during drier conditions (Loisel et al., 2009) or in more decomposed peat (Broder et al., 2012), the opposite effect was also reported (Hong et al., 2001). Nevertheless, this indicates that C isotope ratios may indeed be influenced by decomposition processes. Several publications indicate a decrease in  $\delta^{13}\text{C}$  due to preferential preservation of

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refractory C compounds depleted in <sup>13</sup>C (Akagi et al., 2004; Melillo et al., 1989), but uniform depth trends with no significant C isotope fraction or fractionation effects during CH<sub>4</sub> formation were also observed (Clymo and Bryant, 2008; Kracht and Gleixner, 2000). However, only very few studies have included analyses of peat decomposition or molecular proxies, so that the role of peat decomposition on C isotopes often remains speculative. Values of  $\delta^{13}\text{C}$  in peat from the KK bog range between -24.76‰ and -28.06‰ with a median of -26.1‰, which is typical for C3 plants (Hong et al., 2001), but slightly heavier than the peat forming plants of the Harz mires (-27.42 to -29.06‰) (Table 1). Correlations of the  $\delta^{13}\text{C}$  record with pyrolysis-GC/MS molecular components are comparatively weak. Levoglucosan, the pyrolysis product of cellulose as well as the group of polysaccharides show a positive correlation of  $R^2 = 0.41$  and  $R^2 = 0.37$ , respectively. Best correlation was found with aliphatics ( $R^2 = 0.59$ , negative), which means that  $\delta^{13}\text{C}$  becomes lighter with the enrichment of aliphatics. Moreover, an increase in lignin content during water table lows, as a result of an absolute increase in vascular plants and a relative decrease of polysaccharides might also lower  $\delta^{13}\text{C}$  values, although no correlation between lignin and  $\delta^{13}\text{C}$  has been found. The unusual low  $\delta^{13}\text{C}$  in the surface layer is probably attributed to the present day increase in grasses on the bogs surface or the Suess effect, which caused a decrease in the global atmospheric  $\delta^{13}\text{C}$  value from -6.5 to -8.0‰ since the Industrial Revolution (Francey et al., 1995).

Values of  $\delta^{15}\text{N}$  are not correlated with any of the Pyrolysis-GC-MS components or FTIR band intensities. As an essential nutrient, nitrogen undergoes intense recycling, which most likely is accompanied by intense isotope fractionation processes (Robinson, 2001). Moreover, N isotope variability in different peat forming plants will further add to the overall high variability, so that the fractionation attributed to peat decomposition seems to have a minor effect and thus, is mostly overwritten by other fractionation processes, especially N recycling.  $\delta^{15}\text{N}$  values in the Harz mire's plants (Table 1) were in agreement with other studies on peatland plants (Asada et al., 2005) and were significantly lower than those found in peat (mean -1.23‰ (KK) and 0.75‰, KRB) indi-

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cating that microbial processes, especially denitrification, have removed isotopic light N components from the peat (Jones et al., 2010). Ericoids (*C. vulgaris*) were typically low in  $^{15}\text{N}$  ( $-8.13\text{‰}$ ) due to their association with mycorrhiza (Gebauer and Dietrich, 1993) but this signal is not reflected in the peat due to inverse enrichment of  $^{13}\text{C}$  in the remaining fungal tissues in the peat. The reverse behavior of  $^{15}\text{N}$  with the other proxies in the part were *Sphagnum* dominated and lowest decomposition took place (38–45 cm) suggests that when *Sphagnum* is dominant the effect of vegetation is larger than that of decomposition, which was also found for C/N ratio.

#### 4.3.5 UV-ABS

Although some similarities in the records of UV-ABS values and F1 and aliphatics especially below 25 cm (Supplement Fig. S3) absorption data showed no significant correlations with other molecular components or decomposition indices (Supplement Table S3). However, low absorption values in the upper three samples and in the *Sphagnum* dominated period between 38 and 48 cm agreed with a low degree of decomposition of recent samples and during *Sphagnum* dominance (Figs. 1, 3 and S3 in the Supplement). The absence of good correlations with other proxies can be due to the fact that the peat, although dominated by *Sphagnum*, had a relatively high contribution and high variability of vascular plant species that differ in chemical composition and chemical changes during decay (e.g. graminoids, ericoids, pine, etc.) (Baumann, 2009). UV-ABS of alkaline peat extracts is based on the assumption that more extractable humic acids indicate a higher degree of peat decomposition, but the conditions under which increased “humification” occurs are poorly defined. Moreover, the molecular compounds extracted by alkaline leaching are loosely specified, although it is believed that mostly aromatic moieties such as lignin or phenols are preferentially extracted. However, lignins and aromatics in KK peat have been found not to be correlated to any of the decomposition proxies including the UV-ABS values and the reasons for this lack of significant correlation remains largely unresolved.

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## 5 Conclusions

The two investigated peat cores show similar progression and relationship of decomposition proxies indicating that the observed changes are attributed to external and at least regional forcing. Results from PCA reveal high correlation between C/N, HI and the FTIR-humification index and moderate correlation with  $\delta^{13}\text{C}$  and OI. Accordingly, all investigated proxies show changes upon peat decomposition reflecting changes in mass loss and related relative enrichment of more persistent components. Pyrolysis-GC-MS analyses of molecular components have shown that peat decomposition in KK peat is predominately characterized by preferential decomposition of polysaccharides and enrichment of aliphatics. Although lignins are also enriched during peat decay, their influence on decomposition patterns seems to be of minor importance, here. Although going hand in hand during water table changes, peat decomposition appears to overwrite most of the influence of changes in the contribution of vascular plants in the Harz mires and thus to allow the use of C/N ratio, FTIR index and the HI as bulk decomposition proxies. But in order to understand the processes and to ensure consistency of decomposition proxies, comparison of proxies with detailed molecular information (i.e. analytical pyrolysis) is recommended.

Despite slighter botanical changes, C/N ratios in the Harz peats reliably reflected changes in bog hydrology and mass loss during aerobic decomposition. Hydrogen indices appear to be mainly dependent on the preferential mineralization of polysaccharides and the enrichment of aliphatics, whereas oxygen indices are assumed to be additionally influenced by oxidative input of oxygen functional moieties during aerobic peat decay. FTIR bands give extensive information on the abundance of major groups of organic components and provide a broad picture of qualitative and quantitative changes in the OM structure during decomposition, although being constrained by overlap of defined bands of functional units. In conclusion, the FTIR humification index most reliably reflects peat decomposition in the Harz cores. Values of  $\delta^{13}\text{C}$  in bulk peat show intense changes with a clear trend towards high values with depth and sig-

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nificant correlation with C/N ratios, which hints to an influence of peat decomposition processes probably due to selective enrichment of isotopically lighter compounds such as lignins. The strong variation of  $\delta^{15}\text{N}$  values and the absence of any correlation with peat decomposition proxies indicates that variations in  $\delta^{15}\text{N}$  are mainly controlled by N recycling and plant uptake in the acrotelm and to a low extent by peat decomposition.

Although UV-ABS is frequently used as a measure of humification, its correlation with other decomposition proxies in the KK peat record was generally low. One reason might be that the impact of decomposition on the formation of humic acids is poorly constrained and the results obtained are chemically poorly defined or that the influence of mass loss and the relative enrichment of condensed organic components are more dominant than humification which is mainly a qualitative process. Moreover, the amount of UV-absorbing aromatic compounds which are e.g. abundant in lignin also depend on changes in vegetation, which does not necessarily correlate with decomposition and mass loss.

In conclusion, C/N ratios, Pyrolysis, and FTIR spectroscopy yielded a comprehensive picture of mass loss and the relative abundance of different OM components (e.g. carbohydrates vs. aliphatics), respectively, during peat decomposition at the investigated sites. Future studies need to show to which extent these results can be generalized for other sites and how decomposition proxies compare at sites with different botanical, hydrological or climatic background. However, despite a varying strength of correlation between the different proxies in the different bogs, they all indicated changes in the peat surface wetness, although this may include both, changes in decomposition and in predominant vegetation.

**Supplementary material related to this article is available online at**  
<http://www.biogeosciences-discuss.net/10/17351/2013/bgd-10-17351-2013-supplement.pdf>.

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**Table 1.** Average carbon and nitrogen isotope ratios and C/N ratios in plants from two Harz Mountain bogs (KK, KRB).

Plant species	C/N ratios	$\delta^{13}\text{C}$ [‰]	$\delta^{15}\text{N}$ [‰]
<i>Sphagnum magellanicum</i> ( $n = 3$ )	41.55	-27.67	-3.58
<i>Calluna vulgaris</i> ( $n = 3$ )	37.73	-29.21	-8.13
<i>Eriophorum</i> spec. ( $n = 3$ )	28.58	-27.09	-0.87

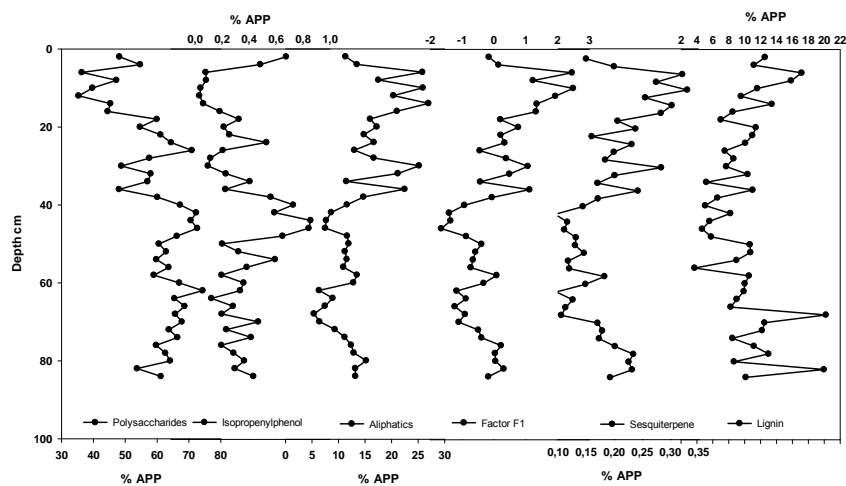
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**Table 2.** Groups of pyrolysis products in peat samples of KK. <sup>a</sup> Mean value for bulk sample relative to the sum of all quantified pyrolysis products ( $n = 42$ ); <sup>b</sup> standard deviation.

Group	avg. % <sup>a</sup>	SD <sup>b</sup>
Polysaccharides	59.2	9.76
Aliphatics	14.1	5.61
Lignins	10.1	3.63
Phenols	8.6	2.68
Aromatics	5.3	4.01
N-compounds	2.1	1.55
Other	0.3	0.34
Polyaromatics	0.2	0.06

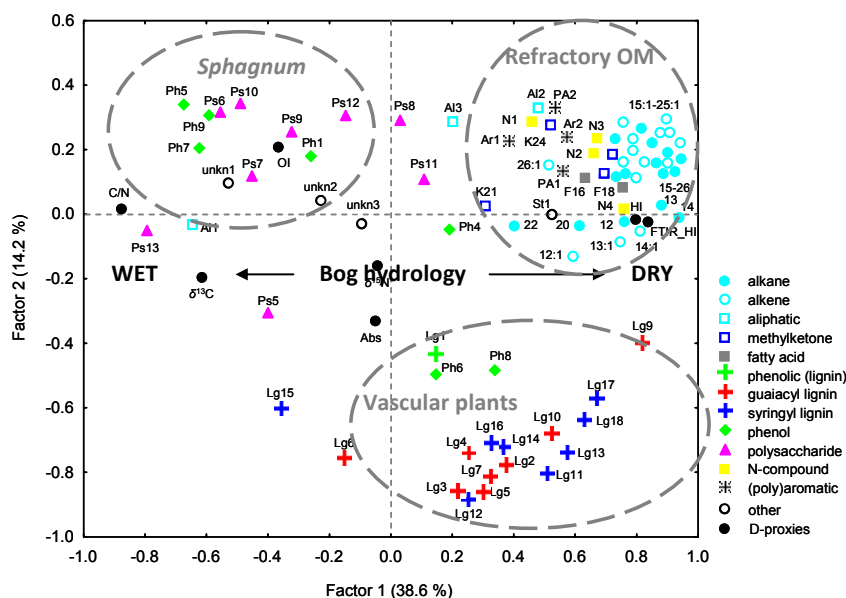
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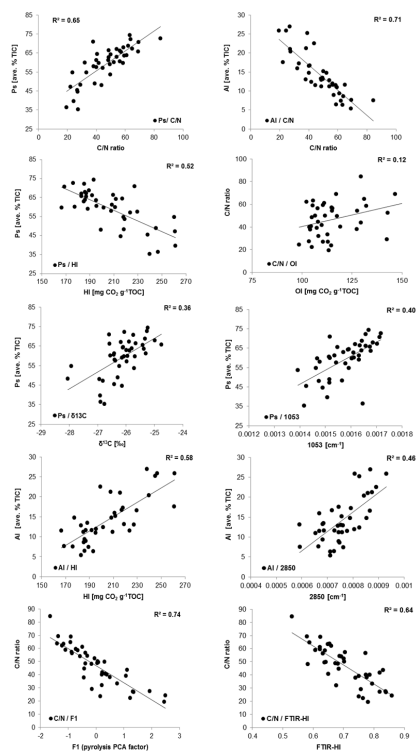
**Fig. 3.** Distribution of selected (groups of) pyrolysis products in the KK peat record indicative for polysaccharides, aliphatics and lignin, the markers of *Sphagnum* (4-Isopropenylphenol) and *Pinus* (sesquiterpene), and Factor F1 of the PCA of the pyrolysis data. Values show percentages for bulk samples relative to the sum of all quantified pyrolysis products (APP).

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**Fig. 4.** Factor loadings of the F1-F2 projection extracted from PCA of pyrolysis products from peat samples of the KK core. Note the opposite loadings of polysaccharides vs. alkanes, alkenes and aliphatics on F1, while lignins have high loadings on F1 and F2 resulting in “fields” separating *Sphagnum* moss from vascular plants and refractory OM. Thus, the projection of F1-F2 factor loadings could help to separate decomposition and vegetation effects.

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**Fig. 5.** Selected correlations between decomposition proxies in the KK core. C/N ratios, FTIR bands  $1053\text{ cm}^{-1}$  and  $2850\text{ cm}^{-1}$ , FTIR-Humification index (FTIR-HI,  $1630/1053$ ), HI and  $\delta^{13}\text{C}$  and molecular components determined by pyrolysis-GC-MS (polysaccharides (Ps), aliphatics (Al) and the PCA factor 1 (F1)).